

MOBT:009
38-21 (13589)

APPLICATION FOR UNITED STATES LETTERS PATENT

FOR

**HYBRID *BACILLUS THURINGIENSIS* δ -ENDOTOXINS WITH NOVEL
BROAD-SPECTRUM INSECTICIDAL ACTIVITY**

BY

THOMAS MALVAR

AND

AMY JELEN GILMER

CERTIFICATE OF EXPRESS MAILING

NUMBER: TB 688 298 831 US

DATE OF DEPOSIT: November 20, 1996

I hereby certify that this paper or fee is being deposited with the United States Postal Service "EXPRESS MAIL POST OFFICE TO ADDRESSEE" service under 37 C.F.R. 1.10 on the date indicated above and is addressed to: Assistant Commissioner for Patents, Washington, DC 20231.

Mark D. Moore
Mark D. Moore

1. BACKGROUND OF THE INVENTION

1.1 FIELD OF THE INVENTION

The present invention relates generally to the fields of molecular biology. More particularly, certain embodiments concern novel nucleic acid segments, and genetically-engineered recombinant δ -endotoxins derived from *Bacillus thuringiensis*. More particularly, it concerns novel chimeric crystal proteins and the chimeric *cry* gene segments which encode them. Various methods for making and using these DNA segments, methods of producing the encoded proteins, methods for making synthetically-modified chimeric crystal proteins, and methods of making and using the synthetic crystal proteins are disclosed, such as, for example, the use of nucleic acid segments as diagnostic probes and templates for protein production, and the use of proteins, fusion protein carriers and peptides in various immunological and diagnostic applications. Also disclosed is the use of these *cry* gene fusions and chimeric Cry proteins in the development of transgenic plants which express broad-spectrum insecticidal activity against a variety of coleopteran, dipteran, and lepidopteran insects.

1.2 DESCRIPTION OF RELATED ART

1.2.1 *Bacillus thuringiensis* Crystal Proteins

The Gram-positive soil bacterium *Bacillus thuringiensis* is well known for its production of proteinaceous parasporal crystals, or δ -endotoxins, that are toxic to a variety of lepidopteran, coleopteran, and dipteran larvae. *B. thuringiensis* produces crystal proteins during sporulation which are specifically toxic to certain species of insects. Many different strains of *B. thuringiensis* have been shown to produce insecticidal crystal proteins, and compositions comprising *B. thuringiensis* strains which produce proteins having insecticidal activity have been used commercially as environmentally-acceptable insecticides because of their toxicity to the specific target insect, and non-toxicity to plants and other non-targeted organisms.

Commercial formulations of naturally occurring *B. thuringiensis* isolates have long been used for the biological control of agricultural insect pests. In commercial production, the spores and crystals obtained from the fermentation process are

concentrated and formulated for foliar application according to conventional agricultural practices.

1.2.2 Nomenclature of Crystal Proteins

5 A review by Höfte *et al.*, (1989) describes the general state of the art with respect to the majority of insecticidal *B. thuringiensis* strains that have been identified which are active against insects of the Order Lepidoptera, *i.e.*, caterpillar insects. This treatise also describes *B. thuringiensis* strains having insecticidal activity against insects of the Orders Diptera (*i.e.* flies and mosquitoes) and Coleoptera (*i.e.* beetles). A number of genes encoding crystal proteins have been cloned from several strains of *B. thuringiensis*. Höfte
10 *et al.* (1989) discusses the genes and proteins that were identified in *B. thuringiensis* prior to 1990, and sets forth the nomenclature and classification scheme which has traditionally been applied to *B. thuringiensis* genes and proteins. *cry1* genes encode lepidopteran-toxic Cry1 proteins. *cry2* genes encode Cry2 proteins that are toxic to both lepidopterans and dipterans. *cry3* genes encode coleopteran-toxic Cry3 proteins, while *cry4* genes
15 encode dipteran-toxic Cry4 proteins, *etc.*

Recently a new nomenclature has been proposed which systematically classifies the Cry proteins based upon amino acid sequence homology rather than upon insect target specificities. This classification scheme is summarized in TABLE 1.

20

TABLE 1
Revised *B. thuringiensis* δ -Endotoxin Nomenclature^a

New	Old	GenBank Accession #
CryIAa	CryIA(a)	M11250
CryIAb	CryIA(b)	M13898
CryIAc	CryIA(c)	M11068
CryIAd	CryIA(d)	M73250
CryIAe	CryIA(e)	M65252
CryIBa	CryIB	X06711
CryIBb	ET5	L32020
CryIBc	PEG5	Z46442
CryICa	CryIC	X07518
CryICb	CryIC(b)	M97880
CryIDa	CryID	X54160
CryIDb	PrtB	Z22511
CryIEa	CryIE	X53985
CryIEb	CryIE(b)	M73253
CryIFa	CryIF	M63897
CryIFb	PrtD	Z22512
CryIG	PrtA	Z22510
CryIH	PrtC	Z22513
CryIHb		U35780
Cry2a	CryV	X62821
Cry2b	CryV	U07642
Cry2Ja	ET4	L32019
Cry1Jb	ET1	U31527
Cry1K		U28801
Cry2Aa	CryIIA	M31738
Cry2Ab	CryIIB	M23724
Cry2Ac	CryIIC	X57252
Cry3A	CryIIIA	M22472
Cry3Ba	CryIIIB	X17123
Cry3Bb	CryIIIB2	M89794
Cry3C	CryIIID	X59797
Cry4A	CryIVA	Y00423
Cry4B	CryIVB	X07423
Cry5Aa	CryVA(a)	L07025
Cry5Ab	CryVA(b)	L07026
Cry5B		U19725
Cry6A	CryVIA	L07022
Cry6B	CryVIB	L07024
Cry7Aa	CryIIIC	M64478

New	Old	GenBank Accession #
Cry7Ab	CryIIICb	U04367
Cry8A	CryIIIE	U04364
Cry8B	CryIIIG	U04365
Cry8C	CryIIIF	U04366
Cry9A	CryIG	X58120
Cry9B	CryIX	X75019
Cry9C	CryIH	Z37527
Cry10A	CryIVC	M12662
Cry11A	CryIVD	M31737
Cry11B	Jeg80	X86902
Cry12A	CryVB	L07027
Cry13A	CryVC	L07023
Cry14A	CryVD	U13955
Cry15A	34kDa	M76442
Cry16A	cbm71	X94146
Cyt1A	CytA	X03182
Cyt2A	CytB	Z14147

^aAdapted from: http://epunix.biols.susx.ac.uk/Home/Neil_Crickmore/Bt/index.html

1.2.3 Mode of Crystal Protein Toxicity

5 All δ -endotoxin crystals are toxic to insect larvae by ingestion. Solubilization of the crystal in the midgut of the insect releases the protoxin form of the δ -endotoxin which, in most instances, is subsequently processed to an active toxin by midgut protease. The activated toxins recognize and bind to the brush-border of the insect midgut epithelium through receptor proteins. Several putative crystal protein receptors have been isolated from certain insect larvae (Knight *et al.*, 1995; Gill *et al.*, 1995; Masson *et al.*, 10 1995). The binding of active toxins is followed by intercalation and aggregation of toxin molecules to form pores within the midgut epithelium. This process leads to osmotic imbalance, swelling, lysis of the cells lining the midgut epithelium, and eventual larvae mortality.

15 1.2.4 Molecular Biology of δ -Endotoxins

With the advent of molecular genetic techniques, various δ -endotoxin genes have been isolated and their DNA sequences determined. These genes have been used to construct certain genetically engineered *B. thuringiensis* products that have been

approved for commercial use. Recent developments have seen new δ -endotoxin delivery systems developed, including plants that contain and express genetically engineered δ -endotoxin genes.

5 The cloning and sequencing of a number of δ -endotoxin genes from a variety of *Bacillus thuringiensis* strains have been described and are summarized by Hofte and Whiteley, 1989. Plasmid shuttle vectors designed for the cloning and expression of δ -endotoxin genes in *E. coli* or *B. thuringiensis* are described by Gawron-Burke and Baum (1991). U. S. Patent No. 5,441,884 discloses a site-specific recombination system for constructing recombinant *B. thuringiensis* strains containing δ -endotoxin genes that
10 are free of DNA not native to *B. thuringiensis*.

The Cry1 family of crystal proteins, which are primarily active against lepidopteran pests, are the best studied class of δ -endotoxins. The pro-toxin form of Cry1 δ -endotoxins consist of two approximately equal sized segments. The carboxyl-half, or pro-toxin segment, is not toxic and is thought to be important for crystal formation
15 (Arvidson *et al.*, 1989). The amino-half of the protoxin comprises the active-toxin segment of the Cry1 molecule and may be further divided into three structural domains as determined by the recently described crystallographic structure for the active toxin segment of the Cry1Aa δ -endotoxin (Grochulski *et al.*, 1995). Domain 1 occupies the first third of the active toxin and is essential for channel formation (Thompson *et al.*,
20 1995). Domain 2 and domain 3 occupy the middle and last third of the active toxin, respectively. Both domains 2 and 3 have been implicated in receptor binding and insect specificity, depending on the insect and δ -endotoxin being examined (Thompson *et al.*, 1995).

25 1.2.5 Chimeric Crystal Proteins

In recent years, researchers have focused effort on the construction of hybrid δ -endotoxins with the hope of producing proteins with enhanced activity or improved properties. Advances in the art of molecular genetics over the past decade have facilitated a logical and orderly approach to engineering proteins with improved

properties. Site-specific and random mutagenesis methods, the advent of polymerase chain reaction (PCR™) methodologies, and the development of recombinant methods for generating gene fusions and constructing chimeric proteins have facilitated an assortment of methods for changing amino acid sequences of proteins, fusing portions of two or more proteins together in a single recombinant protein, and altering genetic sequences that
5 encode proteins of commercial interest.

Unfortunately, for crystal proteins, these techniques have only been exploited in limited fashion. The likelihood of arbitrarily creating a chimeric protein with enhanced properties from portions of the numerous native proteins which have been identified is
10 remote given the complex nature of protein structure, folding, oligomerization, activation, and correct processing of the chimeric protoxin to an active moiety. Only by careful selection of specific target regions within each protein, and subsequent protein engineering can toxins be synthesized which have improved insecticidal activity.

Some success in the area, however, has been reported in the literature. For example, the construction of a few hybrid δ -endotoxins is reported in the following
15 related art: Intl. Pat. Appl. Publ. No. WO 95/30753 discloses the construction of hybrid *B. thuringiensis* δ -endotoxins for production in *Pseudomonas fluorescens* in which the non-toxic protoxin fragment of Cry1F has been replaced by the non-toxic protoxin fragment from the Cry1Ac/Cry1Ab that is disclosed in U. S. Patent No. 5,128,130.

U. S. Patent No. 5,128,130 discloses the construction of hybrid *B. thuringiensis*
20 δ -endotoxins for production in *P. fluorescens* in which a portion of the non-toxic protoxin segment of Cry1Ac is replaced with the corresponding non-toxic protoxin fragment of Cry1Ab. U. S. Patent No. 5,055,294 discloses the construction of a specific hybrid δ -endotoxin between Cry1Ac (amino acid residues 1-466) and Cry1Ab (amino acid
25 residues 466-1155) for production in *P. fluorescens*. Although the aforementioned patent discloses the construction of a hybrid toxin within the active toxin segment, no specifics are presented in regard to the hybrid toxin's insecticidal activity. Intl. Pat. Appl. Publ. No. WO 95/30752 discloses the construction of hybrid *B. thuringiensis* δ -endotoxins for
30 production in *P. fluorescens* in which the non-toxic protoxin segment of Cry1C is replaced by the non-toxic protoxin segment from Cry1Ab. The aforementioned

application further discloses that the activity against *Spodoptera exigua* for the hybrid δ -endotoxin is improved over that of the parent active toxin, Cry1C.

5 Intl. Pat. Appl. Publ. No. WO 95/06730 discloses the construction of a hybrid *B. thuringiensis* δ -endotoxin consisting of domains 1 and 2 of Cry1E coupled to domain 3 and the non-toxic protoxin segment of Cry1C. Insect bioassays performed against *Manduca sexta* (sensitive to Cry1C and Cry1E), *Spodoptera exigua* (sensitive to Cry1C), and *Mamestra brassicae* (sensitive to Cry1C) show that the hybrid Cry1E/Cry1C hybrid toxin is active against *M. sexta*, *S. exigua*, and *M. brassicae*. The bioassay results were expressed as EC₅₀ values (toxin concentration giving a 50% growth reduction) rather than LC₅₀ values (toxin concentration giving 50% mortality). Although the δ -endotoxins used for bioassay were produced in *B. thuringiensis*, only artificially-generated active segments of the δ -endotoxins were used, not the naturally-produced crystals typically produced by *B. thuringiensis* that are present in commercial *B. thuringiensis* formulations. Bioassay results indicated that the LC₅₀ values for the hybrid Cry1E/Cry1C crystal against *S. frugiperda* were 1.5 to 1.7 fold lower (more active) than for native Cry1C. This art also discloses the construction of a hybrid *B. thuringiensis* δ -endotoxin between Cry1Ab (domains 1 and 2) and Cry1C (domain 3 and the non-toxic protoxin segment), although no data are given regarding the hybrid toxin's activity or usefulness.

15 Lee *et al.* (1995) report the construction of hybrid *B. thuringiensis* δ -endotoxins between Cry1Ac and Cry1Aa within the active toxin segment. Artificially generated active segments of the hybrid toxins were used to examine protein interactions in susceptible insect brush border membranes vesicles (BBMV). The bioactivity of the hybrid toxins was not reported.

20 Honee *et al.* (1991) report the construction of hybrid δ -endotoxins between Cry1C (domain 1) and Cry1Ab (domains 2 and 3) and the reciprocal hybrid between Cry1Ab (domain 1) and Cry1C (domains 2 and 3). These hybrids failed to show any significant increase in activity against susceptible insects. Furthermore, the Cry1C (domain 1)/Cry1Ab (domains 2 and 3) hybrid toxin was found to be hypersensitive to protease degradation. A report by Schnepf *et al.* (1990) discloses the construction of Cry1Ac hybrid toxin in which a small portion of domain 2 was replaced by the corresponding

25
30

region of CryIAa, although no significant increase in activity against susceptible insect larvae was observed.

1.3 Deficiencies in the Prior Art

5 The limited successes in producing chimeric crystal proteins which have improved activity have negatively impacted the field by thwarting efforts to produce recombinantly-engineered crystal protein for commercial development, and to extend the toxic properties and host specificities of the known endotoxins. Therefore, what is lacking in the prior art are reliable methods and compositions comprising recombinantly-
10 engineered crystal proteins which have improved insecticidal activity, broad-host-range specificities, and which are suitable for commercial production in *Bacillus thuringiensis*.

2. SUMMARY OF THE INVENTION

The present invention overcomes these and other limitations in the prior art by
15 providing novel chimeric δ -endotoxins which have improved insecticidal properties, and broad-range specificities.

Disclosed are methods for the construction of *B. thuringiensis* hybrid δ -endotoxins comprising amino acid sequences from native CryIAc and CryIF crystal proteins. These hybrid proteins, in which all or a portion of CryIAc domain 2, all or a
20 portion of CryIAc domain 3, and all or a portion of the CryIAc protoxin segment is replaced by the corresponding portions of CryIF, possess not only the insecticidal characteristics of the parent δ -endotoxins, but also have the unexpected and remarkable properties of enhanced broad-range specificity which is not proficiently displayed by either of the native δ -endotoxins from which the chimeric proteins were engineered.

25 Specifically, the present invention discloses and claims genetically-engineered hybrid δ -endotoxins which comprise a portion of a CryIAc crystal protein fused to a portion of a CryIF crystal protein. These chimeric endotoxins have broad-range specificity for the insect pests described herein.

In a further embodiment, the present invention also discloses and claims
30 recombinant *B. thuringiensis* hybrid δ -endotoxins which comprise a portion of CryIAb,

Cry1F, and Cry1Ac in which all or a portion of Cry1Ab domain 2 or all or a portion of Cry1Ab domain 3 is replaced by the corresponding portions of Cry1F and all or a portion of the Cry1Ab protoxin segment is replaced by the corresponding portions of Cry1Ac. Exemplary hybrid δ -endotoxins between Cry1Ab and Cry1F are identified in
5 SEQ ID NO:13 and SEQ ID NO:14.

One aspect of the present invention demonstrates the unexpected result that certain hybrid δ -endotoxins derived from Cry1Ac and Cry1F proteins exhibit not only the insecticidal characteristics of the parent δ -endotoxins, but also possess insecticidal activity which is not proficiently displayed by either of the parent δ -endotoxins.

10 Another aspect of the invention further demonstrates the unexpected result that certain chimeric Cry1Ab/Cry1F proteins maintain not only the insecticidal characteristics of the parent δ -endotoxins, but also exhibit insecticidal activity which is not displayed by either the native Cry1Ab or Cry1F endotoxins.

The present invention also encompasses Cry1Ac/Cry1F and Cry1Ab/Cry1F hybrid
15 δ -endotoxins that maintain the desirable characteristics needed for commercial production in *B. thuringiensis*. Specifically, the hybrid δ -endotoxins identified in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, and SEQ ID NO:30 can efficiently form proteinaceous parasporal inclusions in *B. thuringiensis* and have the favorable characteristics of solubility, protease susceptibility,
20 and insecticidal activity of the parent δ -endotoxins.

In a further embodiment, the present invention also discloses and claims recombinant *B. thuringiensis* hybrid δ -endotoxins which comprise a portion of Cry1Ac and Cry1C in which all or a portion of Cry1Ac domain 3 is replaced by the corresponding portions of Cry1C and all or a portion of the Cry1Ac protoxin segment is replaced by the
25 corresponding portion of Cry1C. Exemplary hybrid δ -endotoxins between Cry1Ac and Cry1C are identified in SEQ ID NO:29 and SEQ ID NO:30.

One aspect of the present invention demonstrates the unexpected result that, although neither Cry1Ac nor Cry1C possess *S. frugiperda* activity, the Cry1Ac/Cry1C hybrid δ -endotoxin identified by SEQ ID NO:29 and SEQ ID NO:30 has significant

activity against *S. frugiperda*. Furthermore, the Cry1Ac/Cry1C hybrid δ -endotoxin identified by SEQ ID NO:29 and SEQ ID NO:30 has significantly better activity against *S. exigua* than the Cry1C parental δ -endotoxin.

5 The present invention further pertains to the recombinant nucleic acid sequences which encode the novel crystal proteins disclosed herein. Specifically, the invention discloses and claims the nucleic acid sequences of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, and SEQ ID NO:29; nucleic acid sequences which are complementary to the nucleic acid sequences of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, and SEQ ID NO:29; and nucleic acid sequences which hybridize to the sequences of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, and SEQ ID NO:29.

10 The novel hybrid δ -endotoxins disclosed herein are useful in the control of a broad range of insect pests. These hybrid δ -endotoxins are described in FIG. 1 and are disclosed in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, and SEQ ID NO:30. The nucleic acid segments encoding these proteins are disclosed in SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, and SEQ ID NO:29. The insecticidal and biochemical properties of the hybrid δ -endotoxins are described by FIG. 2, FIG. 3, and TABLE 3, TABLE 4, TABLE 5, and TABLE 6. The broad host range of the improved δ -endotoxins specified in the present invention is useful in circumventing dilution effects caused by expressing multiple δ -endotoxin genes within a single *B. thuringiensis* strain. Expression of such a broad host range δ -endotoxin in plants is expected to impart protection against a wider variety of insect pests.

20 The impetus for constructing these and other hybrid δ -endotoxins is to create novel toxins with improved insecticidal activity, increased host-range specificity, and improved production characteristics. The DNA sequences listed in TABLE 5 define the exchange points for the hybrid δ -endotoxins pertinent to the present invention and as oligonucleotide primers, may be used to identify like or similar hybrid δ -endotoxins by Southern or colony hybridization under conditions of moderate to high stringency.

5 Researchers skilled in the art will recognize the importance of the exchange site chosen between two or more δ -endotoxins can be achieved using a number of *in vivo* or *in vitro* molecular genetic techniques. Small variations in the exchange region between two or more δ -endotoxins may yield similar results or, as demonstrated for EG11062 and EG11063, adversely affect desirable traits. Similarly, large variations in the exchange region between two or more δ -endotoxins may have no effect on desired traits, as demonstrated by EG11063 and EG11074, or may adversely affect desirable traits, as demonstrated by EG11060 and EG11063.

10 Favorable traits with regard to improved insecticidal activity, increased host range, and improved production characteristics may be achieved by other such hybrid δ -endotoxins including, but not limited to, the *cry1*, *cry2*, *cry3*, *cry4*, *cry5*, *cry6*, *cry7*, *cry8*, *cry9*, *cry10*, *cry11*, *cry12*, *cry13*, *cry14*, *cry15* class of δ -endotoxin genes and the *B. thuringiensis* cytolytic *cyt1* and *cyt2* genes. Members of these classes of *B. thuringiensis* insecticidal proteins include, but are not limited to Cry1Aa, Cry1Ab, Cry1Ac, Cry1Ad, Cry1Ae, Cry1Ba, Cry1Bb, Cry1Ca, Cry1Cb, Cry1Da, Cry1Db, Cry1Ea, Cry1Eb, Cry1Fa, Cry1Fb, Cry1Ga, Cry1Ha, Cry2a, Cry2b, Cry1Ja, Cry1Ka, Cry11Aa, Cry11Ab, Cry12Aa, Cry3Ba, Cry3Bb, Cry3C, Cry4a, Cry4Ba, Cry5a, Cry5Ab, Cry6Aa, Cry6Ba, Cry7Aa, Cry7Ab, Cry8Aa, Cry8Ba, Cry8Ca, Cry9Aa, Cry9Ba, Cry9Ca, Cry10Aa, Cry11Aa, Cry12Aa, Cry13Aa, Cry14Aa, Cry15Aa, Cyt1Aa, and Cyt2Aa. Related hybrid
20 δ -endotoxins would consist of the amino portion of one of the aforementioned δ -endotoxins, including all or part of domain 1 or domain 2, fused to all or part of domain 3 from another of the aforementioned δ -endotoxins. The non-active protoxin fragment of such hybrid δ -endotoxins may consist of the protoxin fragment from any of the aforementioned δ -endotoxins which may act to stabilize the hybrid δ -endotoxin as demonstrated by EG11087 and EG11091 (see *e.g.*, TABLE 3). Hybrid δ -endotoxins
25 possessing similar traits as those described in the present invention could be constructed by conservative, or "similar" replacements of amino acids within hybrid δ -endotoxins. Such substitutions would mimic the biochemical and biophysical properties of the native

amino acid at any position in the protein. Amino acids considered similar include for example, but are not limited to:

Ala, Ser, and Thr

Asp and Glu

5

Asn and Gln

Lys and Arg

Ile, Leu, Met, and Val

Phe, Tyr, and Trp

10 Researchers skilled in the art will recognize that improved insecticidal activity, increased host range, and improved production characteristics imparted upon hybrid δ -endotoxins may be further improved by altering the genetic code for one or more amino acid positions in the hybrid δ -endotoxin such that the position, or positions, is replaced by any other amino acid. This may be accomplished by targeting a region or regions of the protein for mutagenesis by any number of established mutagenic techniques, including
15 those procedures relevant to the present invention. Such techniques include site-specific mutagenesis (Kunkle, 1985; Kunkle *et al.*, 1987), DNA shuffling (Stemmer, 1994), and PCR™ overlap extension (Horton *et al.*, 1989). Since amino acids situated at or near the surface of a protein are likely responsible for its interaction with other proteinaceous or non-proteinaceous moieties, they may serve as "target" regions for mutagenesis. Such
20 surface exposed regions may consist of, but not be limited to, surface exposed amino acid residues within the active toxin fragment of the protein and include the inter- α -helical or inter- β -strand "loop" -regions of δ -endotoxins that separate α -helices within domain 1 and β -strands within domain 2 and domain 3. Such procedures may favorably change the protein's biochemical and biophysical characteristics or its mode of action as outlined in
25 the Section 1. These include, but are not limited to: 1) improved crystal formation, 2) improved protein stability or reduced protease degradation, 3) improved insect membrane receptor recognition and binding, 4) improved oligomerization or channel formation in the insect midgut endothelium, and 5) improved insecticidal activity or insecticidal specificity due to any or all of the reasons stated above.

2.1 Crystal Protein Transgenes and Transgenic Plants

In yet another aspect, the present invention provides methods for producing a transgenic plant which expresses a nucleic acid segment encoding the novel chimeric crystal proteins of the present invention. The process of producing transgenic plants is well-known in the art. In general, the method comprises transforming a suitable host cell with a DNA segment which contains a promoter operatively linked to a coding region that encodes a *B. thuringiensis* Cry1Ac-1F or Cry1Ab-Cry1F, or a Cry1Ab-1Ac-1F chimeric crystal protein. Such a coding region is generally operatively linked to a transcription-terminating region, whereby the promoter is capable of driving the transcription of the coding region in the cell, and hence providing the cell the ability to produce the recombinant protein *in vivo*. Alternatively, in instances where it is desirable to control, regulate, or decrease the amount of a particular recombinant crystal protein expressed in a particular transgenic cell, the invention also provides for the expression of crystal protein antisense mRNA. The use of antisense mRNA as a means of controlling or decreasing the amount of a given protein of interest in a cell is well-known in the art.

Another aspect of the invention comprises a transgenic plant which express a gene or gene segment encoding one or more of the novel polypeptide compositions disclosed herein. As used herein, the term "transgenic plant" is intended to refer to a plant that has incorporated DNA sequences, including but not limited to genes which are perhaps not normally present, DNA sequences not normally transcribed into RNA or translated into a protein ("expressed"), or any other genes or DNA sequences which one desires to introduce into the non-transformed plant, such as genes which may normally be present in the non-transformed plant but which one desires to either genetically engineer or to have altered expression. The construction and expression of synthetic *B. thuringiensis* genes in plants has been described in detail in U. S. Patent No. 5,500,365 and U. S. Patent No. 5,380,831 (each specifically incorporated herein by reference).

It is contemplated that in some instances the genome of a transgenic plant of the present invention will have been augmented through the stable introduction of one or more *cry1Ac-IF*, *cry1Ab-IF*, or *cry1Ab-1Ac-1F* transgenes, either native, synthetically-

modified, or further mutated. In some instances, more than one transgene will be incorporated into the genome of the transformed host plant cell. Such is the case when more than one crystal protein-encoding DNA segment is incorporated into the genome of such a plant. In certain situations, it may be desirable to have one, two, three, four, or even more *B. thuringiensis* crystal proteins (either native or recombinantly-engineered) incorporated and stably expressed in the transformed transgenic plant.

A preferred gene which may be introduced includes, for example, a crystal protein-encoding a DNA sequence from bacterial origin, and particularly one or more of those described herein which are obtained from *Bacillus* spp. Highly preferred nucleic acid sequences are those obtained from *B. thuringiensis*, or any of those sequences which have been genetically engineered to decrease or increase the insecticidal activity of the crystal protein in such a transformed host cell.

Means for transforming a plant cell and the preparation of a transgenic cell line are well-known in the art, and are discussed herein. Vectors, plasmids, cosmids, yeast artificial chromosomes (YACs) and nucleic acid segments for use in transforming such cells will, of course, generally comprise either the operons, genes, or gene-derived sequences of the present invention, either native, or synthetically-derived, and particularly those encoding the disclosed crystal proteins. These DNA constructs can further include structures such as promoters, enhancers, polylinkers, or even gene sequences which have positively- or negatively-regulating activity upon the particular genes of interest as desired. The DNA segment or gene may encode either a native or modified crystal protein, which will be expressed in the resultant recombinant cells, and/or which will impart an improved phenotype to the regenerated plant. Nucleic acid sequences optimized for expression in plants have been disclosed in Intl. Pat. Appl. Publ. No. WO 93/07278 (specifically incorporated herein by reference).

Such transgenic plants may be desirable for increasing the insecticidal resistance of a monocotyledonous or dicotyledonous plant, by incorporating into such a plant, a transgenic DNA segment encoding Cry1Ac-1F and/or Cry1Ab-1F and/or Cry1Ab-1Ac-1F crystal protein(s) which possess broad- insects. Particularly preferred plants such as grains, including but not limited to corn, wheat, oats, rice, maize, and barley; cotton;

soybeans and other legumes; trees, including but not limited to ornamental, shrub, fruit, and nut; vegetables, turf and pasture grasses, fruits, berries, citrus, other crops of commercial interest; including garden and houseplants.

5 In a related aspect, the present invention also encompasses a seed produced by the transformed plant, a progeny from such seed, and a seed produced by the progeny of the original transgenic plant, produced in accordance with the above process. Such progeny and seeds will have a stably crystal protein transgene stably incorporated into its genome, and such progeny plants will inherit the traits afforded by the introduction of a stable transgene in Mendelian fashion. All such transgenic plants having incorporated into their
10 genome transgenic DNA segments encoding one or more chimeric crystal proteins or polypeptides are aspects of this invention.

2.2 Crystal Protein Screening and Immunodetection Kits

15 The present invention contemplates methods and kits for screening samples suspected of containing crystal protein polypeptides or crystal protein-related polypeptides, or cells producing such polypeptides. Said kit can contain a nucleic acid segment or an antibody of the present invention. The kit can contain reagents for detecting an interaction between a sample and a nucleic acid or antibody of the present invention. The provided reagent can be radio-, fluorescently- or enzymatically-labeled.
20 The kit can contain a known radiolabeled agent capable of binding or interacting with a nucleic acid or antibody of the present invention.

The reagent of the kit can be provided as a liquid solution, attached to a solid support or as a dried powder. Preferably, when the reagent is provided in a liquid solution, the liquid solution is an aqueous solution. Preferably, when the reagent
25 provided is attached to a solid support, the solid support can be chromatograph media, a test plate having a plurality of wells, or a microscope slide. When the reagent provided is a dry powder, the powder can be reconstituted by the addition of a suitable solvent, that may be provided.

30 In still further embodiments, the present invention concerns immunodetection methods and associated kits. It is proposed that the crystal proteins or peptides of the

present invention may be employed to detect antibodies having reactivity therewith, or, alternatively, antibodies prepared in accordance with the present invention, may be employed to detect crystal proteins or crystal protein-related epitope-containing peptides. In general, these methods will include first obtaining a sample suspected of containing
5 such a protein, peptide or antibody, contacting the sample with an antibody or peptide in accordance with the present invention, as the case may be, under conditions effective to allow the formation of an immunocomplex, and then detecting the presence of the immunocomplex.

In general, the detection of immunocomplex formation is quite well known in the
10 art and may be achieved through the application of numerous approaches. For example, the present invention contemplates the application of ELISA, RIA, immunoblot (*e.g.*, dot blot), indirect immunofluorescence techniques and the like. Generally, immunocomplex formation will be detected through the use of a label, such as a radiolabel or an enzyme tag (such as alkaline phosphatase, horseradish peroxidase, or the like). Of course, one
15 may find additional advantages through the use of a secondary binding ligand such as a second antibody or a biotin/avidin ligand binding arrangement, as is known in the art.

For assaying purposes, it is proposed that virtually any sample suspected of comprising either a crystal protein or peptide or a crystal protein-related peptide or antibody sought to be detected, as the case may be, may be employed. It is contemplated
20 that such embodiments may have application in the titering of antigen or antibody samples, in the selection of hybridomas, and the like. In related embodiments, the present invention contemplates the preparation of kits that may be employed to detect the presence of crystal proteins or related peptides and/or antibodies in a sample. Samples may include cells, cell supernatants, cell suspensions, cell extracts, enzyme fractions,
25 protein extracts, or other cell-free compositions suspected of containing crystal proteins or peptides. Generally speaking, kits in accordance with the present invention will include a suitable crystal protein, peptide or an antibody directed against such a protein or peptide, together with an immunodetection reagent and a means for containing the antibody or antigen and reagent. The immunodetection reagent will typically comprise a
30 label associated with the antibody or antigen, or associated with a secondary binding

ligand. Exemplary ligands might include a secondary antibody directed against the first antibody or antigen or a biotin or avidin (or streptavidin) ligand having an associated label. Of course, as noted above, a number of exemplary labels are known in the art and all such labels may be employed in connection with the present invention.

5 The container will generally include a vial into which the antibody, antigen or detection reagent may be placed, and preferably suitably aliquotted. The kits of the present invention will also typically include a means for containing the antibody, antigen, and reagent containers in close confinement for commercial sale. Such containers may include injection or blow-molded plastic containers into which the desired vials are
10 retained.

2.3 ELISAs and Immunoprecipitation

ELISAs may be used in conjunction with the invention. In an ELISA assay, proteins or peptides incorporating crystal protein antigen sequences are immobilized onto
15 a selected surface, preferably a surface exhibiting a protein affinity such as the wells of a polystyrene microtiter plate. After washing to remove incompletely adsorbed material, it is desirable to bind or coat the assay plate wells with a nonspecific protein that is known to be antigenically neutral with regard to the test antisera such as bovine serum albumin (BSA), casein or solutions of milk powder. This allows for blocking of nonspecific
20 adsorption sites on the immobilizing surface and thus reduces the background caused by nonspecific binding of antisera onto the surface.

After binding of antigenic material to the well, coating with a non-reactive material to reduce background, and washing to remove unbound material, the immobilizing surface is contacted with the antisera or clinical or biological extract to be
25 tested in a manner conducive to immune complex (antigen/antibody) formation. Such conditions preferably include diluting the antisera with diluents such as BSA, bovine gamma globulin (BGG) and phosphate buffered saline (PBS)/Tween®. These added agents also tend to assist in the reduction of nonspecific background. The layered antisera is then allowed to incubate for from about 2 to about 4 hours, at temperatures
30 preferably on the order of about 25° to about 27°C. Following incubation, the antisera-

contacted surface is washed so as to remove non-immunocomplexed material. A preferred washing procedure includes washing with a solution such as PBS/Tween®, or borate buffer.

5 Following formation of specific immunocomplexes between the test sample and the bound antigen, and subsequent washing, the occurrence and even amount of immunocomplex formation may be determined by subjecting same to a second antibody having specificity for the first. To provide a detecting means, the second antibody will preferably have an associated enzyme that will generate a color development upon incubating with an appropriate chromogenic substrate. Thus, for example, one will desire
10 to contact and incubate the antisera-bound surface with a urease or peroxidase-conjugated anti-human IgG for a period of time and under conditions which favor the development of immunocomplex formation (e.g., incubation for 2 hours at room temperature in a PBS-containing solution such as PBS Tween®).

15 After incubation with the second enzyme-tagged antibody, and subsequent to washing to remove unbound material, the amount of label is quantified by incubation with a chromogenic substrate such as urea and bromocresol purple or 2, 2'-azino-di-(3-ethyl-benzthiazoline)-6-sulfonic acid (ABTS) and H₂O₂, in the case of peroxidase as the enzyme label. Quantitation is then achieved by measuring the degree of color generation, e.g., using a visible spectra spectrophotometer.

20 The anti-crystal protein antibodies of the present invention are particularly useful for the isolation of other crystal protein antigens by immunoprecipitation. Immunoprecipitation involves the separation of the target antigen component from a complex mixture, and is used to discriminate or isolate minute amounts of protein. For the isolation of membrane proteins cells must be solubilized into detergent micelles.
25 Nonionic salts are preferred, since other agents such as bile salts, precipitate at acid pH or in the presence of bivalent cations.

 In an alternative embodiment the antibodies of the present invention are useful for the close juxtaposition of two antigens. This is particularly useful for increasing the localized concentration of antigens, e.g. enzyme-substrate pairs.

2.4 Western Blots

The compositions of the present invention will find great use in immunoblot or western blot analysis. The anti-peptide antibodies may be used as high-affinity primary reagents for the identification of proteins immobilized onto a solid support matrix, such as nitrocellulose, nylon or combinations thereof. In conjunction with immunoprecipitation, followed by gel electrophoresis, these may be used as a single step reagent for use in detecting antigens against which secondary reagents used in the detection of the antigen cause an adverse background. This is especially useful when the antigens studied are immunoglobulins (precluding the use of immunoglobulins binding bacterial cell wall components), the antigens studied cross-react with the detecting agent, or they migrate at the same relative molecular weight as a cross-reacting signal.

Immunologically-based detection methods for use in conjunction with Western blotting include enzymatically-, radiolabel-, or fluorescently-tagged secondary antibodies against the toxin moiety are considered to be of particular use in this regard.

2.5 Epitopic Core Sequences

The present invention is also directed to protein or peptide compositions, free from total cells and other peptides, which comprise a purified protein or peptide which incorporates an epitope that is immunologically cross-reactive with one or more anti-crystal protein antibodies. In particular, the invention concerns epitopic core sequences derived from Cry proteins or peptides.

As used herein, the term "incorporating an epitope(s) that is immunologically cross-reactive with one or more anti-crystal protein antibodies" is intended to refer to a peptide or protein antigen which includes a primary, secondary or tertiary structure similar to an epitope located within a crystal protein or polypeptide. The level of similarity will generally be to such a degree that monoclonal or polyclonal antibodies directed against the crystal protein or polypeptide will also bind to, react with, or otherwise recognize, the cross-reactive peptide or protein antigen. Various immunoassay methods may be employed in conjunction with such antibodies, such as, for example,

Western blotting, ELISA, RIA, and the like, all of which are known to those of skill in the art.

The identification of Cry immunodominant epitopes, and/or their functional equivalents, suitable for use in vaccines is a relatively straightforward matter. For example, one may employ the methods of Hopp, as taught in U. S. Patent No. 4,554,101, incorporated herein by reference, which teaches the identification and preparation of epitopes from amino acid sequences on the basis of hydrophilicity. The methods described in several other papers, and software programs based thereon, can also be used to identify epitopic core sequences (see, for example, Jameson and Wolf, 1988; Wolf *et al.*, 1988; U. S. Patent No. 4,554,101). The amino acid sequence of these "epitopic core sequences" may then be readily incorporated into peptides, either through the application of peptide synthesis or recombinant technology.

Preferred peptides for use in accordance with the present invention will generally be on the order of about 8 to about 20 amino acids in length, and more preferably about 8 to about 15 amino acids in length. It is proposed that shorter antigenic crystal protein-derived peptides will provide advantages in certain circumstances, for example, in the preparation of immunologic detection assays. Exemplary advantages include the ease of preparation and purification, the relatively low cost and improved reproducibility of production, and advantageous biodistribution.

It is proposed that particular advantages of the present invention may be realized through the preparation of synthetic peptides which include modified and/or extended epitopic/immunogenic core sequences which result in a "universal" epitopic peptide directed to crystal proteins, and in particular Cry and Cry-related sequences. These epitopic core sequences are identified herein in particular aspects as hydrophilic regions of the particular polypeptide antigen. It is proposed that these regions represent those which are most likely to promote T-cell or B-cell stimulation, and, hence, elicit specific antibody production.

An epitopic core sequence, as used herein, is a relatively short stretch of amino acids that is "complementary" to, and therefore will bind, antigen binding sites on the crystal protein-directed antibodies disclosed herein. Additionally or alternatively, an

epitopic core sequence is one that will elicit antibodies that are cross-reactive with antibodies directed against the peptide compositions of the present invention. It will be understood that in the context of the present disclosure, the term "complementary" refers to amino acids or peptides that exhibit an attractive force towards each other. Thus, certain epitope core sequences of the present invention may be operationally defined in terms of their ability to compete with or perhaps displace the binding of the desired protein antigen with the corresponding protein-directed antisera.

In general, the size of the polypeptide antigen is not believed to be particularly crucial, so long as it is at least large enough to carry the identified core sequence or sequences. The smallest useful core sequence anticipated by the present disclosure would generally be on the order of about 8 amino acids in length, with sequences on the order of 10 to 20 being more preferred. Thus, this size will generally correspond to the smallest peptide antigens prepared in accordance with the invention. However, the size of the antigen may be larger where desired, so long as it contains a basic epitopic core sequence.

The identification of epitopic core sequences is known to those of skill in the art, for example, as described in U. S. Patent No. 4,554,101, incorporated herein by reference, which teaches the identification and preparation of epitopes from amino acid sequences on the basis of hydrophilicity. Moreover, numerous computer programs are available for use in predicting antigenic portions of proteins (see *e.g.*, Jameson and Wolf, 1988; Wolf *et al.*, 1988). Computerized peptide sequence analysis programs (*e.g.*, DNASTar[®] software, DNASTar, Inc., Madison, WI) may also be useful in designing synthetic peptides in accordance with the present disclosure.

Syntheses of epitopic sequences, or peptides which include an antigenic epitope within their sequence, are readily achieved using conventional synthetic techniques such as the solid phase method (*e.g.*, through the use of commercially available peptide synthesizer such as an Applied Biosystems Model 430A Peptide Synthesizer). Peptide antigens synthesized in this manner may then be aliquotted in predetermined amounts and stored in conventional manners, such as in aqueous solutions or, even more preferably, in a powder or lyophilized state pending use.

In general, due to the relative stability of peptides, they may be readily stored in aqueous solutions for fairly long periods of time if desired, *e.g.*, up to six months or more, in virtually any aqueous solution without appreciable degradation or loss of antigenic activity. However, where extended aqueous storage is contemplated it will generally be desirable to include agents including buffers such as Tris or phosphate buffers to maintain a pH of about 7.0 to about 7.5. Moreover, it may be desirable to include agents which will inhibit microbial growth, such as sodium azide or Merthiolate. For extended storage in an aqueous state it will be desirable to store the solutions at about 4°C, or more preferably, frozen. Of course, where the peptides are stored in a lyophilized or powdered state, they may be stored virtually indefinitely, *e.g.*, in metered aliquots that may be rehydrated with a predetermined amount of water (preferably distilled) or buffer prior to use.

2.6 Nucleic Acid Segments Encoding Crystal Protein Chimeras

The present invention also concerns DNA segments, both native, synthetic, and mutagenized, that can be synthesized, or isolated from virtually any source, that are free from total genomic DNA and that encode the novel chimeric peptides disclosed herein. DNA segments encoding these peptide species may prove to encode proteins, polypeptides, subunits, functional domains, and the like of crystal protein-related or other non-related gene products. In addition these DNA segments may be synthesized entirely *in vitro* using methods that are well-known to those of skill in the art.

As used herein, the term "DNA segment" refers to a DNA molecule that has been isolated free of total genomic DNA of a particular species. Therefore, a DNA segment encoding a crystal protein or peptide refers to a DNA segment that contains crystal protein coding sequences yet is isolated away from, or purified free from, total genomic DNA of the species from which the DNA segment is obtained, which in the instant case is the genome of the Gram-positive bacterial genus, *Bacillus*, and in particular, the species of *Bacillus* known as *B. thuringiensis*. Included within the term "DNA segment", are DNA segments and smaller fragments of such segments, and also recombinant

vectors, including, for example, plasmids, cosmids, phagemids, phage, viruses, and the like.

Similarly, a DNA segment comprising an isolated or purified crystal protein-encoding gene refers to a DNA segment which may include in addition to peptide-encoding sequences, certain other elements such as, regulatory sequences, isolated
5 substantially away from other naturally occurring genes or protein-encoding sequences. In this respect, the term "gene" is used for simplicity to refer to a functional protein-, polypeptide- or peptide-encoding unit. As will be understood by those in the art, this functional term includes both genomic sequences, operon sequences and smaller
10 engineered gene segments that express, or may be adapted to express, proteins, polypeptides or peptides.

"Isolated substantially away from other coding sequences" means that the gene of interest, in this case, a gene encoding a bacterial crystal protein, forms the significant part of the coding region of the DNA segment, and that the DNA segment does not contain
15 large portions of naturally-occurring coding DNA, such as large chromosomal fragments or other functional genes or operon coding regions. Of course, this refers to the DNA segment as originally isolated, and does not exclude genes, recombinant genes, synthetic linkers, or coding regions later added to the segment by the hand of man.

In particular embodiments, the invention concerns isolated DNA segments and
20 recombinant vectors incorporating DNA sequences that encode a Cry peptide species that includes within its amino acid sequence an amino acid sequence essentially as set forth in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, or SEQ ID NO:30.

The term "a sequence essentially as set forth in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, or SEQ ID NO:30" means that the
25 sequence substantially corresponds to a portion of the sequence of either SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, or SEQ ID NO:30 and has relatively few amino acids that are not identical to, or a biologically functional equivalent of, the amino acids of any of these sequences. The term "biologically
30 functional equivalent" is well understood in the art and is further defined in detail herein

(e.g., see Illustrative Embodiments). Accordingly, sequences that have between about 70% and about 80%, or more preferably between about 81% and about 90%, or even more preferably between about 91% and about 99% amino acid sequence identity or functional equivalence to the amino acids of SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, or SEQ ID NO:30 will be sequences
5 that are "essentially as set forth in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, or SEQ ID NO:30."

It will also be understood that amino acid and nucleic acid sequences may include additional residues, such as additional N- or C-terminal amino acids or 5' or 3' sequences,
10 and yet still be essentially as set forth in one of the sequences disclosed herein, so long as the sequence meets the criteria set forth above, including the maintenance of biological protein activity where protein expression is concerned. The addition of terminal sequences particularly applies to nucleic acid sequences that may, for example, include various non-coding sequences flanking either of the 5' or 3' portions of the coding region
15 or may include various internal sequences, i.e., introns, which are known to occur within genes.

The nucleic acid segments of the present invention, regardless of the length of the coding sequence itself, may be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other
20 coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol. For example, nucleic acid fragments may be prepared that include a short contiguous stretch encoding either of the peptide
25 sequences disclosed in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, or SEQ ID NO:30, or that are identical to or complementary to DNA sequences which encode any of the peptides disclosed in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, or SEQ ID NO:30, and particularly those DNA segments disclosed in SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13,
30 SEQ ID NO:25, SEQ ID NO:27, or SEQ ID NO:29. For example, DNA sequences such

as about 14 nucleotides, and that are up to about 10,000, about 5,000, about 3,000, about 2,000, about 1,000, about 500, about 200, about 100, about 50, and about 14 base pairs in length (including all intermediate lengths) are also contemplated to be useful.

5 It will be readily understood that "intermediate lengths", in these contexts, means any length between the quoted ranges, such as 14, 15, 16, 17, 18, 19, 20, *etc.*; 21, 22, 23, *etc.*; 30, 31, 32, *etc.*; 50, 51, 52, 53, *etc.*; 100, 101, 102, 103, *etc.*; 150, 151, 152, 153, *etc.*; including all integers through the 200-500; 500-1,000; 1,000-2,000; 2,000-3,000; 3,000-5,000; and up to and including sequences of about 10,000 nucleotides and the like.

10 It will also be understood that this invention is not limited to the particular nucleic acid sequences which encode peptides of the present invention, or which encode the amino acid sequences of SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, or SEQ ID NO:30, including those DNA sequences which are particularly disclosed in SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27 or SEQ ID NO:29. Recombinant vectors and isolated
15 DNA segments may therefore variously include the peptide-coding regions themselves, coding regions bearing selected alterations or modifications in the basic coding region, or they may encode larger polypeptides that nevertheless include these peptide-coding regions or may encode biologically functional equivalent proteins or peptides that have variant amino acids sequences.

20 The DNA segments of the present invention encompass biologically-functional, equivalent peptides. Such sequences may arise as a consequence of codon redundancy and functional equivalency that are known to occur naturally within nucleic acid sequences and the proteins thus encoded. Alternatively, functionally-equivalent proteins or peptides may be created via the application of recombinant DNA technology, in which
25 changes in the protein structure may be engineered, based on considerations of the properties of the amino acids being exchanged. Changes designed by man may be introduced through the application of site-directed mutagenesis techniques, *e.g.*, to introduce improvements to the antigenicity of the protein or to test mutants in order to examine activity at the molecular level.

If desired, one may also prepare fusion proteins and peptides, e.g., where the peptide-coding regions are aligned within the same expression unit with other proteins or peptides having desired functions, such as for purification or immunodetection purposes (e.g., proteins that may be purified by affinity chromatography and enzyme label coding regions, respectively).

Recombinant vectors form further aspects of the present invention. Particularly useful vectors are contemplated to be those vectors in which the coding portion of the DNA segment, whether encoding a full length protein or smaller peptide, is positioned under the control of a promoter. The promoter may be in the form of the promoter that is naturally associated with a gene encoding peptides of the present invention, as may be obtained by isolating the 5' non-coding sequences located upstream of the coding segment or exon, for example, using recombinant cloning and/or PCR™ technology, in connection with the compositions disclosed herein.

2.7 Recombinant Vectors and Protein Expression

In other embodiments, it is contemplated that certain advantages will be gained by positioning the coding DNA segment under the control of a recombinant, or heterologous, promoter. As used herein, a recombinant or heterologous promoter is intended to refer to a promoter that is not normally associated with a DNA segment encoding a crystal protein or peptide in its natural environment. Such promoters may include promoters normally associated with other genes, and/or promoters isolated from any bacterial, viral, eukaryotic, or plant cell. Naturally, it will be important to employ a promoter that effectively directs the expression of the DNA segment in the cell type, organism, or even animal, chosen for expression. The use of promoter and cell type combinations for protein expression is generally known to those of skill in the art of molecular biology, for example, see Sambrook *et al.*, 1989. The promoters employed may be constitutive, or inducible, and can be used under the appropriate conditions to direct high level expression of the introduced DNA segment, such as is advantageous in the large-scale production of recombinant proteins or peptides. Appropriate promoter systems

contemplated for use in high-level expression include, but are not limited to, the *Pichia* expression vector system (Pharmacia LKB Biotechnology).

In connection with expression embodiments to prepare recombinant proteins and peptides, it is contemplated that longer DNA segments will most often be used, with DNA segments encoding the entire peptide sequence being most preferred. However, it will be appreciated that the use of shorter DNA segments to direct the expression of crystal peptides or epitopic core regions, such as may be used to generate anti-crystal protein antibodies, also falls within the scope of the invention. DNA segments that encode peptide antigens from about 8 to about 50 amino acids in length, or more preferably, from about 8 to about 30 amino acids in length, or even more preferably, from about 8 to about 20 amino acids in length are contemplated to be particularly useful. Such peptide epitopes may be amino acid sequences which comprise contiguous amino acid sequences from SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, or SEQ ID NO:30; or any peptide epitope encoded by the nucleic acid sequences of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, or SEQ ID NO:29.

Methods for the recombinant expression of crystal proteins and vectors useful in the expression of DNA constructs encoding crystal proteins are described in Intl. Pat. Appl. Publ. No. WO 95/02058, specifically incorporated herein by reference.

2.8 DNA Segments as Hybridization Probes and Primers

In addition to their use in directing the expression of crystal proteins or peptides of the present invention, the nucleic acid sequences contemplated herein also have a variety of other uses. For example, they also have utility as probes or primers in nucleic acid hybridization embodiments. As such, it is contemplated that nucleic acid segments that comprise a sequence region that consists of at least a 14 nucleotide long contiguous sequence that has the same sequence as, or is complementary to, a 14 nucleotide long contiguous DNA segment of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, or SEQ ID NO:29 will find particular utility. Also, nucleic acid segments which encode at least a 6 amino acid contiguous sequence from

SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, or
SEQ ID NO:30, are also preferred. Longer contiguous identical or complementary
sequences, *e.g.*, those of about 20, 30, 40, 50, 100, 200, 500, 1000, 2000, 5000, 10000
etc. (including all intermediate lengths and up to and including full-length sequences will
5 also be of use in certain embodiments.

The ability of such nucleic acid probes to specifically hybridize to crystal protein-
encoding sequences will enable them to be of use in detecting the presence of
complementary sequences in a given sample. However, other uses are envisioned,
including the use of the sequence information for the preparation of mutant species
10 primers, or primers for use in preparing other genetic constructions.

Nucleic acid molecules having sequence regions consisting of contiguous
nucleotide stretches of 10-14, 15-20, 30, 50, or even of 100-200 nucleotides or so,
identical or complementary to DNA sequences of SEQ ID NO:9, SEQ ID NO:11,
SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, or SEQ ID NO:29, are particularly
15 contemplated as hybridization probes for use in, *e.g.*, Southern and Northern blotting.
Smaller fragments will generally find use in hybridization embodiments, wherein the
length of the contiguous complementary region may be varied, such as between about 10-
14 and about 100 or 200 nucleotides, but larger contiguous complementarity stretches
may be used, according to the length complementary sequences one wishes to detect.

Of course, fragments may also be obtained by other techniques such as, *e.g.*, by
20 mechanical shearing or by restriction enzyme digestion. Small nucleic acid segments or
fragments may be readily prepared by, for example, directly synthesizing the fragment by
chemical means, as is commonly practiced using an automated oligonucleotide
synthesizer. Also, fragments may be obtained by application of nucleic acid reproduction
25 technology, such as the PCR™ technology of U. S. Patent Nos. 4,683,195 and 4,683,202
(each specifically incorporated herein by reference), by introducing selected sequences
into recombinant vectors for recombinant production, and by other recombinant DNA
techniques generally known to those of skill in the art of molecular biology.

Accordingly, the nucleotide sequences of the invention may be used for their
30 ability to selectively form duplex molecules with complementary stretches of DNA

fragments. Depending on the application envisioned, one will desire to employ varying conditions of hybridization to achieve varying degrees of selectivity of probe towards target sequence. For applications requiring high selectivity, one will typically desire to employ relatively stringent conditions to form the hybrids, *e.g.*, one will select relatively low salt and/or high temperature conditions, such as provided by about 0.02 M to about 0.15 M NaCl at temperatures of about 50°C to about 70°C. Such selective conditions tolerate little, if any, mismatch between the probe and the template or target strand, and would be particularly suitable for isolating crystal protein-encoding DNA segments. Detection of DNA segments via hybridization is well-known to those of skill in the art, and the teachings of U. S. Patent Nos. 4,965,188 and 5,176,995 (each specifically incorporated herein by reference) are exemplary of the methods of hybridization analyses. Teachings such as those found in the texts of Maloy *et al.*, 1994; Segal 1976; Prokop, 1991; and Kuby, 1994, are particularly relevant.

Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template or where one seeks to isolate crystal protein-encoding sequences from related species, functional equivalents, or the like, less stringent hybridization conditions will typically be needed in order to allow formation of the heteroduplex. In these circumstances, one may desire to employ conditions such as about 0.15 M to about 0.9 M salt, at temperatures ranging from about 20°C to about 55°C. Cross-hybridizing species can thereby be readily identified as positively hybridizing signals with respect to control hybridizations. In any case, it is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide, which serves to destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired results.

In certain embodiments, it will be advantageous to employ nucleic acid sequences of the present invention in combination with an appropriate means, such as a label, for determining hybridization. A wide variety of appropriate indicator means are known in the art, including fluorescent, radioactive, enzymatic or other ligands, such as

avidin/biotin, which are capable of giving a detectable signal. In preferred embodiments, one will likely desire to employ a fluorescent label or an enzyme tag, such as urease, alkaline phosphatase or peroxidase, instead of radioactive or other environmental undesirable reagents. In the case of enzyme tags, colorimetric indicator substrates are known that can be employed to provide a means visible to the human eye or spectrophotometrically, to identify specific hybridization with complementary nucleic acid-containing samples.

In general, it is envisioned that the hybridization probes described herein will be useful both as reagents in solution hybridization as well as in embodiments employing a solid phase. In embodiments involving a solid phase, the test DNA (or RNA) is adsorbed or otherwise affixed to a selected matrix or surface. This fixed, single-stranded nucleic acid is then subjected to specific hybridization with selected probes under desired conditions. The selected conditions will depend on the particular circumstances based on the particular criteria required (depending, for example, on the G+C content, type of target nucleic acid, source of nucleic acid, size of hybridization probe, *etc.*). Following washing of the hybridized surface so as to remove nonspecifically bound probe molecules, specific hybridization is detected, or even quantitated, by means of the label.

2.9 Biological Functional Equivalents

Modification and changes may be made in the structure of the peptides of the present invention and DNA segments which encode them and still obtain a functional molecule that encodes a protein or peptide with desirable characteristics. The following is a discussion based upon changing the amino acids of a protein to create an equivalent, or even an improved, second-generation molecule. In particular embodiments of the invention, mutated crystal proteins are contemplated to be useful for increasing the insecticidal activity of the protein, and consequently increasing the insecticidal activity and/or expression of the recombinant transgene in a plant cell. The amino acid changes may be achieved by changing the codons of the DNA sequence, according to the codons given in TABLE 2.

TABLE 2

Amino Acid			Codons						
Alanine	Ala	A	GCA	GCC	GCG	GCU			
Cysteine	Cys	C	UGC	UGU					
Aspartic acid	Asp	D	GAC	GAU					
Glutamic acid	Glu	E	GAA	GAG					
Phenylalanine	Phe	F	UUC	UUU					
Glycine	Gly	G	GGA	GGC	GGG	GGU			
Histidine	His	H	CAC	CAU					
Isoleucine	Ile	I	AUA	AUC	AUU				
Lysine	Lys	K	AAA	AAG					
Leucine	Leu	L	UUA	UUG	CUA	CUC	CUG	CUU	
Methionine	Met	M	AUG						
Asparagine	Asn	N	AAC	AAU					
Proline	Pro	P	CCA	CCC	CCG	CCU			
Glutamine	Gln	Q	CAA	CAG					
Arginine	Arg	R	AGA	AGG	CGA	CGC	CGG	CGU	
Serine	Ser	S	AGC	AGU	UCA	UCC	UCG	UCU	
Threonine	Thr	T	ACA	ACC	ACG	ACU			
Valine	Val	V	GUA	GUC	GUG	GUU			
Tryptophan	Trp	W	UGG						
Tyrosine	Tyr	Y	UAC	UAU					

For example, certain amino acids may be substituted for other amino acids in a protein structure without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate

molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence substitutions can be made in a protein sequence, and, of course, its underlying DNA coding sequence, and nevertheless obtain a protein with like properties. It is thus contemplated by the inventors
5 that various changes may be made in the peptide sequences of the disclosed compositions, or corresponding DNA sequences which encode said peptides without appreciable loss of their biological utility or activity.

In making such changes, the hydropathic index of amino acids may be considered. The importance of the hydropathic amino acid index in conferring interactive biologic
10 function on a protein is generally understood in the art (Kyte and Doolittle, 1982, incorporate herein by reference). It is accepted that the relative hydropathic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes, substrates, receptors, DNA, antibodies, antigens, and the like.

Each amino acid has been assigned a hydropathic index on the basis of their hydrophobicity and charge characteristics (Kyte and Doolittle, 1982), these are: isoleucine
15 (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (-0.4); threonine (-0.7); serine (-0.8); tryptophan (-0.9); tyrosine (-1.3); proline (-1.6); histidine (-3.2); glutamate (-3.5);
20 glutamine (-3.5); aspartate (-3.5); asparagine (-3.5); lysine (-3.9); and arginine (-4.5).

It is known in the art that certain amino acids may be substituted by other amino acids having a similar hydropathic index or score and still result in a protein with similar biological activity, *i.e.*, still obtain a biological functionally equivalent protein. In making
25 such changes, the substitution of amino acids whose hydropathic indices are within ± 2 is preferred, those which are within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U. S. Patent No. 4,554,101, incorporated herein by reference, states that the greatest local average hydrophilicity of a protein, as

governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein.

As detailed in U. S. Patent No. 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate (+3.0 \pm 1); glutamate (+3.0 \pm 1); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0); threonine (-0.4); proline (-0.5 \pm 1); alanine (-0.5); histidine (-0.5); cysteine (-1.0); methionine (-1.3); valine (-1.5); leucine (-1.8); isoleucine (-1.8); tyrosine (-2.3); phenylalanine (-2.5); tryptophan (-3.4).

It is understood that an amino acid can be substituted for another having a similar hydrophilicity value and still obtain a biologically equivalent, and in particular, an immunologically equivalent protein. In such changes, the substitution of amino acids whose hydrophilicity values are within ± 2 is preferred, those which are within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

As outlined above, amino acid substitutions are generally therefore based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions which take various of the foregoing characteristics into consideration are well known to those of skill in the art and include: arginine and lysine; glutamate and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine.

2.10 Site-Specific Mutagenesis

Site-specific mutagenesis is a technique useful in the preparation of individual peptides, or biologically functional equivalent proteins or peptides, through specific mutagenesis of the underlying DNA. The technique further provides a ready ability to prepare and test sequence variants, for example, incorporating one or more of the foregoing considerations, by introducing one or more nucleotide sequence changes into the DNA. Site-specific mutagenesis allows the production of mutants through the use of specific oligonucleotide sequences which encode the DNA sequence of the desired mutation, as well as a sufficient number of adjacent nucleotides, to provide a primer sequence of sufficient size and sequence complexity to form a stable duplex on both sides

of the deletion junction being traversed. Typically, a primer of about 17 to 25 nucleotides in length is preferred, with about 5 to 10 residues on both sides of the junction of the sequence being altered.

In general, the technique of site-specific mutagenesis is well known in the art, as exemplified by various publications. As will be appreciated, the technique typically employs a phage vector which exists in both a single stranded and double stranded form. Typical vectors useful in site-directed mutagenesis include vectors such as the M13 phage. These phage are readily commercially available and their use is generally well known to those skilled in the art. Double stranded plasmids are also routinely employed in site directed mutagenesis which eliminates the step of transferring the gene of interest from a plasmid to a phage.

In general, site-directed mutagenesis in accordance herewith is performed by first obtaining a single-stranded vector or melting apart of two strands of a double stranded vector which includes within its sequence a DNA sequence which encodes the desired peptide. An oligonucleotide primer bearing the desired mutated sequence is prepared, generally synthetically. This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as *E. coli* polymerase I Klenow fragment, in order to complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is then used to transform appropriate cells, such as *E. coli* cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement.

The preparation of sequence variants of the selected peptide-encoding DNA segments using site-directed mutagenesis is provided as a means of producing potentially useful species and is not meant to be limiting as there are other ways in which sequence variants of peptides and the DNA sequences encoding them may be obtained. For example, recombinant vectors encoding the desired peptide sequence may be treated with mutagenic agents, such as hydroxylamine, to obtain sequence variants.

2.11 Crystal Protein Compositions As Insecticides and Methods of Use

The inventors contemplate that the chimeric crystal protein compositions disclosed herein will find particular utility as insecticides for topical and/or systemic application to field crops, grasses, fruits and vegetables, and ornamental plants. In a preferred embodiment, the bioinsecticide composition comprises an oil flowable suspension of bacterial cells which expresses a novel crystal protein disclosed herein. Preferably the cells are *B. thuringiensis* cells, however, any such bacterial host cell expressing the novel nucleic acid segments disclosed herein and producing a crystal protein is contemplated to be useful, such as *B. megaterium*, *B. subtilis*, *E. coli*, or *Pseudomonas* spp.

In another important embodiment, the bioinsecticide composition comprises a water dispersible granule. This granule comprises bacterial cells which expresses a novel crystal protein disclosed herein. Preferred bacterial cells are *B. thuringiensis* cells, however, bacteria such as *B. megaterium*, *B. subtilis*, *E. coli*, or *Pseudomonas* spp. cells transformed with a DNA segment disclosed herein and expressing the crystal protein are also contemplated to be useful.

In a third important embodiment, the bioinsecticide composition comprises a wettable powder, dust, pellet, or colloidal concentrate. This powder comprises bacterial cells which expresses a novel crystal protein disclosed herein. Preferred bacterial cells are *B. thuringiensis* cells, however, bacteria such as *B. megaterium*, *B. subtilis*, *E. coli*, or *Pseudomonas* spp. cells transformed with a DNA segment disclosed herein and expressing the crystal protein are also contemplated to be useful. Such dry forms of the insecticidal compositions may be formulated to dissolve immediately upon wetting, or alternatively, dissolve in a controlled-release, sustained-release, or other time-dependent manner.

In a fourth important embodiment, the bioinsecticide composition comprises an aqueous suspension of bacterial cells such as those described above which express the crystal protein. Such aqueous suspensions may be provided as a concentrated stock-solution which is diluted prior to application, or alternatively, as a diluted solution ready-to-apply.

For these methods involving application of bacterial cells, the cellular host containing the crystal protein gene(s) may be grown in any convenient nutrient medium, where the DNA construct provides a selective advantage, providing for a selective medium so that substantially all or all of the cells retain the *B. thuringiensis* gene. These cells may then be harvested in accordance with conventional ways. Alternatively, the cells can be treated prior to harvesting.

When the insecticidal compositions comprise intact *B. thuringiensis* cells expressing the protein of interest, such bacteria may be formulated in a variety of ways. They may be employed as wettable powders, granules or dusts, by mixing with various inert materials, such as inorganic minerals (phyllosilicates, carbonates, sulfates, phosphates, and the like) or botanical materials (powdered corncobs, rice hulls, walnut shells, and the like). The formulations may include spreader-sticker adjuvants, stabilizing agents, other pesticidal additives, or surfactants. Liquid formulations may be aqueous-based or non-aqueous and employed as foams, suspensions, emulsifiable concentrates, or the like. The ingredients may include rheological agents, surfactants, emulsifiers, dispersants, or polymers.

Alternatively, the novel chimeric Cry proteins may be prepared by recombinant bacterial expression systems *in vitro* and isolated for subsequent field application. Such protein may be either in crude cell lysates, suspensions, colloids, etc., or alternatively may be purified, refined, buffered, and/or further processed, before formulating in an active biocidal formulation. Likewise, under certain circumstances, it may be desirable to isolate crystals and/or spores from bacterial cultures expressing the crystal protein and apply solutions, suspensions, or colloidal preparations of such crystals and/or spores as the active bioinsecticidal composition.

Regardless of the method of application, the amount of the active component(s) are applied at an insecticidally-effective amount, which will vary depending on such factors as, for example, the specific coleopteran insects to be controlled, the specific plant or crop to be treated, the environmental conditions, and the method, rate, and quantity of application of the insecticidally-active composition.

The insecticide compositions described may be made by formulating either the bacterial cell, crystal and/or spore suspension, or isolated protein component with the desired agriculturally-acceptable carrier. The compositions may be formulated prior to administration in an appropriate means such as lyophilized, freeze-dried, desiccated, or in an aqueous carrier, medium or suitable diluent, such as saline or other buffer. The formulated compositions may be in the form of a dust or granular material, or a suspension in oil (vegetable or mineral), or water or oil/water emulsions, or as a wettable powder, or in combination with any other carrier material suitable for agricultural application. Suitable agricultural carriers can be solid or liquid and are well known in the art. The term "agriculturally-acceptable carrier" covers all adjuvants, *e.g.*, inert components, dispersants, surfactants, tackifiers, binders, *etc.* that are ordinarily used in insecticide formulation technology; these are well known to those skilled in insecticide formulation. The formulations may be mixed with one or more solid or liquid adjuvants and prepared by various means, *e.g.*, by homogeneously mixing, blending and/or grinding the insecticidal composition with suitable adjuvants using conventional formulation techniques.

The insecticidal compositions of this invention are applied to the environment of the target coleopteran insect, typically onto the foliage of the plant or crop to be protected, by conventional methods, preferably by spraying. The strength and duration of insecticidal application will be set with regard to conditions specific to the particular pest(s), crop(s) to be treated and particular environmental conditions. The proportional ratio of active ingredient to carrier will naturally depend on the chemical nature, solubility, and stability of the insecticidal composition, as well as the particular formulation contemplated.

Other application techniques, *e.g.*, dusting, sprinkling, soaking, soil injection, seed coating, seedling coating, spraying, aerating, misting, atomizing, and the like, are also feasible and may be required under certain circumstances such as *e.g.*, insects that cause root or stalk infestation, or for application to delicate vegetation or ornamental plants. These application procedures are also well-known to those of skill in the art.

The insecticidal composition of the invention may be employed in the method of the invention singly or in combination with other compounds, including and not limited to other pesticides. The method of the invention may also be used in conjunction with other treatments such as surfactants, detergents, polymers or time-release formulations. The insecticidal compositions of the present invention may be formulated for either systemic or topical use.

The concentration of insecticidal composition which is used for environmental, systemic, or foliar application will vary widely depending upon the nature of the particular formulation, means of application, environmental conditions, and degree of biocidal activity. Typically, the bioinsecticidal composition will be present in the applied formulation at a concentration of at least about 0.5% by weight and may be up to and including about 99% by weight. Dry formulations of the compositions may be from about 0.5% to about 99% or more by weight of the composition, while liquid formulations may generally comprise from about 0.5% to about 99% or more of the active ingredient by weight. Formulations which comprise intact bacterial cells will generally contain from about 10^4 to about 10^{12} cells/mg.

The insecticidal formulation may be administered to a particular plant or target area in one or more applications as needed, with a typical field application rate per hectare ranging on the order of from about 50 g to about 500 g of active ingredient, or of from about 500 g to about 1000 g, or of from about 1000 g to about 5000 g or more of active ingredient.

2.12 Chemical and Pharmaceutical Compositions

The inventors contemplate in addition to treating plants and their environments with the insecticidal compositions of the invention, the treatment of animals and their surroundings will also be possible with certain chimeric crystal proteins which have activity against insects which infest animals and their environment.

The inventors particularly contemplate the use of chimeric crystal proteins in the formulation of bioinsecticides comprising specific chimeric δ -endotoxin proteins which have insecticidal activity against one or more insects which infest pets, livestock, and

other domestic animals. The formulation of pharmaceutical compositions which may be given to animals as prophylaxis and/or treatment of infestation by such insects, and in particular mosquitoes, flies, fleas, and related insects will be particularly useful in treating such infestations. Insects as described in detail in U. S. Patent No. 5,449,681, incorporated herein by reference, may be particularly susceptible to treatment in this manner. Such insects include members of the Genera *Culex*, *Culiseta*, *Bovicola*, *Callitroga*, *Chrysops*, *Cimes*, *Ctenocephalis*, *Ctenocephaledes*, *Dermatophilus*, *Dermatobia*, and *Damalinia* among others.

Means for administering insecticidal compositions to an animal are well-known in the art. U. S. Patent No. 5,416,102 (specifically incorporated herein by reference) provides excellent teaching for methods and formulations for preventing insect infestation using an insecticidal composition. Veterinary-approved formulations may be delivered in a variety of methods depending upon the particular application.

It is further contemplated that in addition to topical administration of the pharmaceutical compositions disclosed, systemic administration may in some cases be preferable or desirable. For oral administration, the compositions may be formulated with an inert diluent or with an assimilatable edible carrier, or they may be enclosed in hard- or soft-shell gelatin capsule, or they may be compressed into tablets, or they may be incorporated directly with the food of the diet. For oral therapeutic administration, the active compounds may be incorporated with excipients and used in the form of ingestible tablets, buccal tables, troches, capsules, elixirs, suspensions, syrups, wafers, and the like. Such compositions and preparations should contain at least 0.1% of active compound. The percentage of the compositions and preparations may, of course, be varied and may conveniently be between about 2 to about 60% of the weight of the unit. The amount of active compounds in such therapeutically useful compositions is such that a suitable dosage will be obtained.

For oral prophylaxis, the crystal protein may be incorporated with excipients and used in the form of a gel, paste, powder, pill, tablet, capsule, or slurry which may be given to the animal for ingestion. Alternatively the compositions may be formulated as an additive to pet foods, treats, or other edible formulations. When formulated as a tablet or

capsule, or the like, the composition may also contain the following: a binder, as gum tragacanth, acacia, cornstarch, or gelatin; excipients, such as dicalcium phosphate; a disintegrating agent, such as corn starch, potato starch, alginic acid and the like; a lubricant, such as magnesium stearate; and a sweetening agent, such as sucrose, lactose or saccharin may be added or a flavoring agent to make the composition more palatable to the animal being treated. One such means for delivering prophylactics to an animal is a sauce as described in U. S. Patent No. 4,702,914 (specifically incorporated herein by reference).

When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar or both. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active compounds may be incorporated into sustained-release preparation and formulations.

Alternatively, the pharmaceutical compositions disclosed herein may be administered parenterally, intramuscularly, or even intraperitoneally. Solutions of the active compounds as free base or pharmacologically acceptable salts may be prepared in water suitably mixed with a surfactant, such as hydroxypropylcellulose. Dispersions may also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms. The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (e.g., glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and/or vegetable oils. Proper fluidity may be maintained, for example, by the use

of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

When systemic administration is desired, *e.g.*, parenteral administration in an aqueous solution, the solution should be suitably buffered if necessary and the liquid diluent first rendered isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intramuscular, subcutaneous and intraperitoneal administration. In this connection, sterile aqueous media which can be employed will be known to those of skill in the art in light of the present disclosure. Some variation in dosage will necessarily occur depending on the condition, size, and type of animal being treated. The person responsible for administration will, in any event, determine the appropriate dose for the individual subject. Moreover, for human administration, preparations should meet sterility, pyrogenicity, general safety and purity standards as required by FDA Office of Biologics standards.

Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the various sterilized active ingredients into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum-drying and freeze-drying techniques which yield a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

The compositions disclosed herein may be formulated in a neutral or salt form. Pharmaceutically-acceptable salts, include the acid addition salts (formed with the free

amino groups of the protein) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, histidine, procaine and the like. Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically effective. The formulations are easily administered in a variety of dosage forms such as creams, lotions, sprays, dips, emulsions, colloids, or alternatively, when systemic administration is desirable, injectable solutions, drug release capsules and the like.

As used herein, "carrier" includes any and all solvents, dispersion media, vehicles, coatings, diluents, antibacterial and antifungal agents, isotonic and absorption delaying agents, buffers, carrier solutions, suspensions, colloids, and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, its use in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions.

The phrase "pharmaceutically-acceptable" refers to molecular entities and compositions that do not produce an allergic or similar untoward reaction when administered to a animal. The preparation of an aqueous composition that contains a protein as an active ingredient is well understood in the art. Typically, such compositions are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid prior to injection can also be prepared. The preparation can also be emulsified.

The inventors further contemplate the use of chimeric crystal proteins of the present invention as active ingredients in pharmaceutical compositions for administration to the living areas and environment of an animal to prevent, lessen, or reduce the infestation of susceptible insects in such an area. The chimeric proteins or a suspension of cells which express the particular chimeric protein(s) may be formulated in a powder, spray, fog, granule, rinse, shampoo, dip, *etc.* suitable for administration to the living

quarters, bedding materials, or environment of the animal using techniques which are known to those of skill in the art of veterinary insecticide formulations. An example of oral formulation of veterinary insecticides is found in the teachings of U. S. Patent No. 5,416,102.

5

2.13 Antibody Compositions and Methods for Producing

In particular embodiments, the inventors contemplate the use of antibodies, either monoclonal or polyclonal which bind to the crystal proteins disclosed herein. Means for preparing and characterizing antibodies are well known in the art (See, *e.g.*, Harlow and Lane, 1988; incorporated herein by reference). The methods for generating monoclonal antibodies (mAbs) generally begin along the same lines as those for preparing polyclonal antibodies. Briefly, a polyclonal antibody is prepared by immunizing an animal with an immunogenic composition in accordance with the present invention and collecting antisera from that immunized animal. A wide range of animal species can be used for the production of antisera. Typically the animal used for production of anti-antisera is a rabbit, a mouse, a rat, a hamster, a guinea pig or a goat. Because of the relatively large blood volume of rabbits, a rabbit is a preferred choice for production of polyclonal antibodies.

As is well known in the art, a given composition may vary in its immunogenicity. It is often necessary therefore to boost the host immune system, as may be achieved by coupling a peptide or polypeptide immunogen to a carrier. Exemplary and preferred carriers are keyhole limpet hemocyanin (KLH) and bovine serum albumin (BSA). Other albumins such as ovalbumin, mouse serum albumin or rabbit serum albumin can also be used as carriers. Means for conjugating a polypeptide to a carrier protein are well known in the art and include glutaraldehyde, *m*-maleimidobencoyl-N-hydroxysuccinimide ester, carbodiimide and bis-biazotized benzidine.

As is also well known in the art, the immunogenicity of a particular immunogen composition can be enhanced by the use of non-specific stimulators of the immune response, known as adjuvants. Exemplary and preferred adjuvants include complete Freund's adjuvant (a non-specific stimulator of the immune response containing killed

Mycobacterium tuberculosis), incomplete Freund's adjuvants and aluminum hydroxide adjuvant.

5 The amount of immunogen composition used in the production of polyclonal antibodies varies upon the nature of the immunogen as well as the animal used for immunization. A variety of routes can be used to administer the immunogen (subcutaneous, intramuscular, intradermal, intravenous and intraperitoneal). The production of polyclonal antibodies may be monitored by sampling blood of the immunized animal at various points following immunization. A second booster injection may also be given. The process of boosting and titering is repeated until a
10 suitable titer is achieved. When a desired level of immunogenicity is obtained, the immunized animal can be bled and the serum isolated and stored, and/or the animal can be used to generate mAbs.

mAbs may be readily prepared through use of well-known techniques, such as those exemplified in U. S. Patent No. 4,196,265 (specifically incorporated herein by
15 reference). Typically, this technique involves immunizing a suitable animal with a selected immunogen composition, e.g., a purified or partially purified crystal protein, polypeptide or peptide. The immunizing composition is administered in a manner effective to stimulate antibody producing cells. Rodents such as mice and rats are preferred animals, however, the use of rabbit, sheep frog cells is also possible. The use of
20 rats may provide certain advantages (Goding, 1986, pp. 60-61), but mice are preferred, with the BALB/c mouse being most preferred as this is most routinely used and generally gives a higher percentage of stable fusions.

Following immunization, somatic cells with the potential for producing antibodies, specifically B lymphocytes (B cells), are selected for use in the mAb
25 generating protocol. These cells may be obtained from biopsied spleens, tonsils or lymph nodes, or from a peripheral blood sample. Spleen cells and peripheral blood cells are preferred, the former because they are a rich source of antibody-producing cells that are in the dividing plasmablast stage, and the latter because peripheral blood is easily accessible. Often, a panel of animals will have been immunized and the spleen of animal with the
30 highest antibody titer will be removed and the spleen lymphocytes obtained by

homogenizing the spleen with a syringe. Typically, a spleen from an immunized mouse contains approximately 5×10^7 to 2×10^8 lymphocytes.

5 The antibody-producing B lymphocytes from the immunized animal are then fused with cells of an immortal myeloma cell, generally one of the same species as the animal that was immunized. Myeloma cell lines suited for use in hybridoma-producing fusion procedures preferably are non-antibody-producing, have high fusion efficiency, and enzyme deficiencies that render them incapable of growing in certain selective media which support the growth of only the desired fused cells (hybridomas).

10 Any one of a number of myeloma cells may be used, as are known to those of skill in the art (Goding, pp. 65-66, 1986; Campbell, pp. 75-83, 1984). For example, where the immunized animal is a mouse, one may use P3-X63/Ag8, X63-Ag8.653, NS1/1.Ag 4 1, Sp210-Ag14, FO, NSO/U, MPC-11, MPC11-X45-GTG 1.7 and S194/5XX0 Bul; for rats, one may use R210.RCY3, Y3-Ag 1.2.3, IR983F and 4B210; and U-266, GM1500-GRG2, LICR-LON-HMy2 and UC729-6 are all useful in connection with human cell fusions.

15 One preferred murine myeloma cell is the NS-1 myeloma cell line (also termed P3-NS-1-Ag4-1), which is readily available from the NIGMS Human Genetic Mutant Cell Repository by requesting cell line repository number GM3573. Another mouse myeloma cell line that may be used is the 8-azaguanine-resistant mouse murine myeloma SP2/0 non-producer cell line.

20 Methods for generating hybrids of antibody-producing spleen or lymph node cells and myeloma cells usually comprise mixing somatic cells with myeloma cells in a 2:1 ratio, though the ratio may vary from about 20:1 to about 1:1, respectively, in the presence of an agent or agents (chemical or electrical) that promote the fusion of cell membranes. Fusion methods using Sendai virus have been described (Kohler and
25 Milstein, 1975; 1976), and those using polyethylene glycol (PEG), such as 37% (v/v) PEG, (Geftter *et al.*, 1977). The use of electrically induced fusion methods is also appropriate (Goding, 1986, pp. 71-74).

30 Fusion procedures usually produce viable hybrids at low frequencies, about 1×10^{-6} to 1×10^{-8} . However, this does not pose a problem, as the viable, fused hybrids are differentiated from the parental, unfused cells (particularly the unfused myeloma cells

that would normally continue to divide indefinitely) by culturing in a selective medium. The selective medium is generally one that contains an agent that blocks the *de novo* synthesis of nucleotides in the tissue culture media. Exemplary and preferred agents are aminopterin, methotrexate, and azaserine. Aminopterin and methotrexate block *de novo* synthesis of both purines and pyrimidines, whereas azaserine blocks only purine synthesis. Where aminopterin or methotrexate is used, the media is supplemented with hypoxanthine and thymidine as a source of nucleotides (HAT medium). Where azaserine is used, the media is supplemented with hypoxanthine.

The preferred selection medium is HAT. Only cells capable of operating nucleotide salvage pathways are able to survive in HAT medium. The myeloma cells are defective in key enzymes of the salvage pathway, *e.g.*, hypoxanthine phosphoribosyl transferase (HPRT), and they cannot survive. The B-cells can operate this pathway, but they have a limited life span in culture and generally die within about two weeks. Therefore, the only cells that can survive in the selective media are those hybrids formed from myeloma and B-cells.

This culturing provides a population of hybridomas from which specific hybridomas are selected. Typically, selection of hybridomas is performed by culturing the cells by single-clone dilution in microtiter plates, followed by testing the individual clonal supernatants (after about two to three weeks) for the desired reactivity. The assay should be sensitive, simple and rapid, such as radioimmunoassays, enzyme immunoassays, cytotoxicity assays, plaque assays, dot immunobinding assays, and the like.

The selected hybridomas would then be serially diluted and cloned into individual antibody-producing cell lines, which clones can then be propagated indefinitely to provide mAbs. The cell lines may be exploited for mAb production in two basic ways. A sample of the hybridoma can be injected (often into the peritoneal cavity) into a histocompatible animal of the type that was used to provide the somatic and myeloma cells for the original fusion. The injected animal develops tumors secreting the specific monoclonal antibody produced by the fused cell hybrid. The body fluids of the animal, such as serum or ascites fluid, can then be tapped to provide mAbs in high concentration.

The individual cell lines could also be cultured *in vitro*, where the mAbs are naturally secreted into the culture medium from which they can be readily obtained in high concentrations. mAbs produced by either means may be further purified, if desired, using filtration, centrifugation and various chromatographic methods such as HPLC or affinity chromatography.

3. BRIEF DESCRIPTION OF THE DRAWINGS

The following drawings form part of the present specification and are included to further demonstrate certain aspects of the present invention. The invention may be better understood by reference to one or more of these drawings in combination with the detailed description of specific embodiments presented herein.

FIG. 1. The wild-type δ -endotoxins and the relevant restriction sites that were used to construct the hybrid δ -endotoxins pertinent to the invention are diagrammed in FIG. 1A. Only the DNA encoding the δ -endotoxin that is contained on the indicated plasmid (identified by the "pEG" prefix) is shown. The *B. thuringiensis* strains containing the indicated plasmids are identified by the "EG" prefix. The hybrid δ -endotoxins described in the invention are diagrammed in FIG. 1B and are aligned with the wild-type δ -endotoxins in FIG. 1A.

FIG. 2. An equal amount of each washed sporulated *B. thuringiensis* culture was analyzed by SDS-PAGE. Lane a: control Cry1Ac producing *B. thuringiensis* strain EG11070, b: EG11060, c: EG11062, d: EG11063, e: EG11065, f: EG11067, g: EG11071, h: EG11073, i: EG11074, j: EG11088, k: EG11090, and l: EG11091.

FIG. 3. Solubilized hybrid δ -endotoxins were exposed to trypsin for 0, 15, 30, 60, and 120 minutes. The resulting material was analyzed by SDS-PAGE. The amount of active δ -endotoxin fragment remaining was quantitated by scanning densitometry using a Molecular Dynamics model 300A densitometer. The percent active toxin remaining was plotted versus time. Wild-type Cry1Ac δ -endotoxin (open box) served as the control.

4. BRIEF DESCRIPTION OF THE SEQUENCES

- SEQ ID NO:1 is oligonucleotide primer A.
- 5 SEQ ID NO:2 is oligonucleotide primer B.
- SEQ ID NO:3 is oligonucleotide primer C.
- SEQ ID NO:4 is oligonucleotide primer D.
- SEQ ID NO:5 is oligonucleotide primer E.
- SEQ ID NO:6 is oligonucleotide primer F.
- 10 SEQ ID NO:7 is oligonucleotide primer G.
- SEQ ID NO:8 is oligonucleotide primer H.
- SEQ ID NO:9 is the nucleotide and deduced amino acid sequences of the EG11063 hybrid δ -endotoxin.
- SEQ ID NO:10 denotes the three-letter abbreviation form of the amino acid sequence for
- 15 the hybrid δ -endotoxin specified in SEQ ID NO:9.
- SEQ ID NO:11 is the nucleotide and deduced amino acid sequences of the EG11074 hybrid δ -endotoxin.
- SEQ ID NO:12 denotes the three-letter abbreviation form of the amino acid sequence for
- the hybrid δ -endotoxin specified in SEQ ID NO:11.
- 20 SEQ ID NO:13 is the nucleotide and deduced amino acid sequences of the EG11735 hybrid δ -endotoxin.
- SEQ ID NO:14 denotes the three-letter abbreviation form of the amino acid sequence for
- the hybrid δ -endotoxin specified in SEQ ID NO:13.
- SEQ ID NO:15 is the 5' exchange site for pEG1065, pEG1070, and pEG1074.
- 25 SEQ ID NO:16 is the 5' exchange site for pEG1067, pEG1072, and pEG1076.
- SEQ ID NO:17 is the 5' exchange site for pEG1068, pEG1077, and pEG365.
- SEQ ID NO:18 is the 5' exchange site for pEG1088 and pEG1088.
- SEQ ID NO:19 is the 5' exchange site for pEG1089 and the 3' exchange site for pEG1070 and pEG1072.

SEQ ID NO:20 is the 5' exchange site for pEG1091.

SEQ ID NO:21 is the 3' exchange site for pEG1065, pEG1067, pEG1068, and pEG 365.

SEQ ID NO:22 is the 3' exchange site for pEG1088.

SEQ ID NO:23 is oligonucleotide Primer I.

5 SEQ ID NO:24 is oligonucleotide Primer J.

SEQ ID NO:25 is the nucleic acid sequence and deduced amino acid sequence of the hybrid crystal protein-encoding gene of EG11092.

SEQ ID NO:26 is the three-letter abbreviation form of the amino acid sequence of the hybrid crystal protein produced by strain EG11092 encoded by SEQ ID NO:25.

10 SEQ ID NO:27 is the nucleic acid sequence and the deduced amino acid sequence of the hybrid crystal protein-encoding gene of EG11751.

SEQ ID NO:28 is the three-letter abbreviation form of the amino acid sequence of the hybrid crystal protein produced by strain EG11751 encoded by SEQ ID NO:27.

15 SEQ ID NO:29 is the nucleic acid sequence and the deduced amino acid sequence of the hybrid crystal protein-encoding gene of EG11090.

SEQ ID NO:30 is the three-letter abbreviation form of the amino acid sequence of the hybrid crystal protein produced by strain EG11090 encoded by SEQ ID NO:29.

5. DESCRIPTION OF ILLUSTRATIVE EMBODIMENTS

20 5.1 Methods for Culturing *B. thuringiensis* to Produce the Novel Proteins

The *B. thuringiensis* strains described herein may be cultured using standard known media and fermentation techniques. Upon completion of the fermentation cycle, the bacteria may be harvested by first separating the *B. thuringiensis* spores and crystals from the fermentation broth by means well known in the art. The recovered *B.*
25 *thuringiensis* spores and crystals can be formulated into a wettable powder, a liquid concentrate, granules or other formulations by the addition of surfactants, dispersants, inert carriers and other components to facilitate handling and application for particular target pests. The formulation and application procedures are all well known in the art and are used with commercial strains of *B. thuringiensis* (HD-1) active against Lepidoptera,
30 e.g., caterpillars.

5.2 Recombinant Host Cells For Expression of the Novel Chimeric Genes

The nucleotide sequences of the subject invention can be introduced into a wide variety of microbial hosts. Expression of the toxin gene results, directly or indirectly, in the intracellular production and maintenance of the pesticide. With suitable hosts, e.g., *Pseudomonas*, the microbes can be applied to the sites of lepidopteran insects where they will proliferate and be ingested by the insects. The results is a control of the unwanted insects. Alternatively, the microbe hosting the toxin gene can be treated under conditions that prolong the activity of the toxin produced in the cell. The treated cell then can be applied to the environment of target pest(s). The resulting product retains the toxicity of the *B. thuringiensis* toxin.

Suitable host cells, where the pesticide-containing cells will be treated to prolong the activity of the toxin in the cell when the then treated cell is applied to the environment of target pest(s), may include either prokaryotes or eukaryotes, normally being limited to those cells which do not produce substances toxic to higher organisms, such as mammals. However, organisms which produce substances toxic to higher organisms could be used, where the toxin is unstable or the level of application sufficiently low as to avoid any possibility or toxicity to a mammalian host. As hosts, of particular interest will be the prokaryotes and the lower eukaryotes, such as fungi. Illustrative prokaryotes, both Gram-negative and Gram-positive, include *Enterobacteriaceae*, such as *Escherichia*, *Erwinia*, *Shigella*, *Salmonella*, and *Proteus*; *Bacillaceae*; *Rhizobiceae*, such as *Rhizobium*; *Spirillaceae*, such as photobacterium, *Zymomonas*, *Serratia*, *Aeromonas*, *Vibrio*, *Desulfovibrio*, *Spirillum*; *Lactobacillaceae*; *Pseudomonadaceae*, such as *Pseudomonas* and *Acetobacter*; *Azotobacteraceae*, *Actinomycetales*, and *Nitrobacteraceae*. Among eukaryotes are fungi, such as *Phycomycetes* and *Ascomycetes*, which includes yeast, such as *Saccharomyces* and *Schizosaccharomyces*; and *Basidiomycetes* yeast, such as *Rhodotorula*, *Aureobasidium*, *Sporobolomyces*, and the like.

Characteristics of particular interest in selecting a host cell for purposes of production include ease of introducing the *B. thuringiensis* gene into the host, availability of expression systems, efficiency of expression, stability of the pesticide in the host, and

the presence of auxiliary genetic capabilities. Characteristics of interest for use as a pesticide microcapsule include protective qualities for the pesticide, such as thick cell walls, pigmentation, and intracellular packaging or formation of inclusion bodies; leaf affinity; lack of mammalian toxicity; attractiveness to pests for ingestion; ease of killing and fixing without damage to the toxin; and the like. Other considerations include ease of formulation and handling, economics, storage stability, and the like.

Host organisms of particular interest include yeast, such as *Rhodotorula sp.*, *Aureobasidium sp.*, *Saccharomyces sp.*, and *Sporobolomyces sp.*; phylloplane organisms such as *Pseudomonas sp.*, *Erwinia sp.* and *Flavobacterium sp.*; or such other organisms as *Escherichia*, *Lactobacillus sp.*, *Bacillus sp.*, *Streptomyces sp.*, and the like. Specific organisms include *Pseudomonas aeruginosa*, *Pseudomonas fluorescens*, *Saccharomyces cerevisiae*, *Bacillus thuringiensis*, *Escherichia coli*, *Bacillus subtilis*, *Streptomyces lividans* and the like.

Treatment of the microbial cell, e.g., a microbe containing the *B. thuringiensis* toxin gene, can be by chemical or physical means, or by a combination of chemical and/or physical means, so long as the technique does not deleteriously affect the properties of the toxin, nor diminish the cellular capability in protecting the toxin. Examples of chemical reagents are halogenating agents, particularly halogens of atomic no. 17-80. More particularly, iodine can be used under mild conditions and for sufficient time to achieve the desired results. Other suitable techniques include treatment with aldehydes, such as formaldehyde and glutaraldehyde; anti-infectives, such as zephiran chloride and cetylpyridinium chloride; alcohols, such as isopropyl and ethanol; various histologic fixatives, such as Lugol's iodine, Bouin's fixative, and Helly's fixatives, (see e.g., Humason, 1967); or a combination of physical (heat) and chemical agents that preserve and prolong the activity of the toxin produced in the cell when the cell is administered to a suitable host. Examples of physical means are short wavelength radiation such as γ -radiation and X-radiation, freezing, UV irradiation, lyophilization, and the like. The cells employed will usually be intact and be substantially in the proliferative form when treated, rather than in a spore form, although in some instances spores may be employed.

Where the *B. thuringiensis* toxin gene is introduced via a suitable vector into a microbial host, and said host is applied to the environment in a living state, it is essential that certain host microbes be used. Microorganism hosts are selected which are known to occupy the "phytosphere" (phylloplane, phyllosphere, rhizosphere, and/or rhizoplane) of one or more crops of interest. These microorganisms are selected so as to be capable of successfully competing in the particular environment (crop and other insect habitats) with the wild-type microorganisms, provide for stable maintenance and expression of the gene expressing the polypeptide pesticide, and, desirably, provide for improved protection of the pesticide from environmental degradation and inactivation.

A large number of microorganisms are known to inhabit the phylloplane (the surface of the plant leaves) and/or the rhizosphere (the soil surrounding plant roots) of a wide variety of important crops. These microorganisms include bacteria, algae, and fungi. Of particular interest are microorganisms, such as bacteria, e.g., genera *Bacillus*, *Pseudomonas*, *Erwinia*, *Serratia*, *Klebsiella*, *Xanthomonas*, *Streptomyces*, *Rhizobium*, *Rhodopseudomonas*, *Methylophilus*, *Agrobacterium*, *Acetobacter*, *Lactobacillus*, *Arthrobacter*, *Azotobacter*, *Leuconostoc*, and *Alcaligenes*; fungi, particularly yeast, e.g., genera *Saccharomyces*, *Cryptococcus*, *Kluyveromyces*, *Sporobolomyces*, *Rhodotorula*, and *Aureobasidium*. Of particular interest are such phytosphere bacterial species as *Pseudomonas syringae*, *Pseudomonas fluorescens*, *Serratia marcescens*, *Acetobacter xylinum*, *Agrobacterium tumefaciens*, *Rhodobacter sphaeroides*, *Xanthomonas campestris*, *Rhizobium melioli*, *Alcaligenes eutrophus*, and *Azotobacter vinlandii*; and phytosphere yeast species such as *Rhodotorula rubra*, *R. glutinis*, *R. marina*, *R. aurantiaca*, *Cryptococcus albidus*, *C. diffluens*, *C. laurentii*, *Saccharomyces rosei*, *S. pretoriensis*, *S. cerevisiae*, *Sporobolomyces roseus*, *S. odoratus*, *Kluyveromyces veronae*, and *Aureobasidium pollulans*.

5.3 Definitions

The following words and phrases have the meanings set forth below.

Broad-Spectrum: refers to a wide range of insect species.

Broad-Spectrum Insecticidal Activity: toxicity towards a wide range of insect species.

Expression: The combination of intracellular processes, including transcription and translation undergone by a coding DNA molecule such as a structural gene to produce a polypeptide.

Insecticidal Activity: toxicity towards insects.

5 **Insecticidal Specificity:** the toxicity exhibited by a crystal protein towards multiple insect species.

Intraorder Specificity: the toxicity of a particular crystal protein towards insect species within an Order of insects (*e.g.*, Order Lepidoptera).

10 **Interorder Sspecificity:** the toxicity of a particular crystal protein towards insect species of different Orders (*e.g.*, Orders Lepidoptera and Diptera).

LC₅₀: the lethal concentration of crystal protein that causes 50% mortality of the insects treated.

LC₉₅: the lethal concentration of crystal protein that causes 95% mortality of the insects treated.

15 **Promoter:** A recognition site on a DNA sequence or group of DNA sequences that provide an expression control element for a structural gene and to which RNA polymerase specifically binds and initiates RNA synthesis (transcription) of that gene.

20 **Regeneration:** The process of growing a plant from a plant cell (*e.g.*, plant protoplast or explant).

Structural Gene: A gene that is expressed to produce a polypeptide.

Transformation: A process of introducing an exogenous DNA sequence (*e.g.*, a vector, a recombinant DNA molecule) into a cell or protoplast in which that exogenous DNA is incorporated into a chromosome or is capable of autonomous replication.

25 **Transformed Cell:** A cell whose DNA has been altered by the introduction of an exogenous DNA molecule into that cell.

Transgene: An exogenous gene which when introduced into the genome of a host cell through a process such as transformation, electroporation, particle bombardment, and the like, is expressed by the host cell and integrated into the cells genome

such that the trait or traits produced by the expression of the transgene is inherited by the progeny of the transformed cell.

5 **Transgenic Cell:** Any cell derived or regenerated from a transformed cell or derived from a transgenic cell. Exemplary transgenic cells include plant calli derived from a transformed plant cell and particular cells such as leaf, root, stem, *e.g.*, somatic cells, or reproductive (germ) cells obtained from a transgenic plant.

10 **Transgenic Plant:** A plant or progeny thereof derived from a transformed plant cell or protoplast, wherein the plant DNA contains an introduced exogenous DNA molecule not originally present in a native, non-transgenic plant of the same strain. The terms "transgenic plant" and "transformed plant" have sometimes been used in the art as synonymous terms to define a plant whose DNA contains an exogenous DNA molecule. However, it is thought more scientifically correct to refer to a regenerated plant or callus obtained from a transformed plant cell or protoplast as being a transgenic plant, and that usage will be followed herein.

15 **Vector:** A DNA molecule capable of replication in a host cell and/or to which another DNA segment can be operatively linked so as to bring about replication of the attached segment. A plasmid is an exemplary vector.

20 5.4 Probes And Primers

20 In another aspect, DNA sequence information provided by the invention allows for the preparation of relatively short DNA (or RNA) sequences having the ability to specifically hybridize to gene sequences of the selected polynucleotides disclosed herein. In these aspects, nucleic acid probes of an appropriate length are prepared based on a consideration of a selected crystal protein gene sequence, *e.g.*, a sequence such as that
25 shown in SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, or SEQ ID NO:29. The ability of such nucleic acid probes to specifically hybridize to a crystal protein-encoding gene sequence lends them particular utility in a variety of embodiments. Most importantly, the probes may be used in a variety of assays for detecting the presence of complementary sequences in a given sample.

In certain embodiments, it is advantageous to use oligonucleotide primers. The sequence of such primers is designed using a polynucleotide of the present invention for use in detecting, amplifying or mutating a defined segment of a crystal protein gene from *B. thuringiensis* using PCR™ technology. Segments of related crystal protein genes from other species may also be amplified by PCR™ using such primers.

To provide certain of the advantages in accordance with the present invention, a preferred nucleic acid sequence employed for hybridization studies or assays includes sequences that are complementary to at least a 14 to 30 or so long nucleotide stretch of a crystal protein-encoding sequence, such as that shown in SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, or SEQ ID NO:29. A size of at least 14 nucleotides in length helps to ensure that the fragment will be of sufficient length to form a duplex molecule that is both stable and selective. Molecules having complementary sequences over stretches greater than 14 bases in length are generally preferred, though, in order to increase stability and selectivity of the hybrid, and thereby improve the quality and degree of specific hybrid molecules obtained. One will generally prefer to design nucleic acid molecules having gene-complementary stretches of 14 to 20 nucleotides, or even longer where desired. Such fragments may be readily prepared by, for example, directly synthesizing the fragment by chemical means, by application of nucleic acid reproduction technology, such as the PCR™ technology of U. S. Patent Nos. 4,683,195, and 4,683,202, each specifically incorporated herein by reference, or by excising selected DNA fragments from recombinant plasmids containing appropriate inserts and suitable restriction sites.

5.5 Expression Vectors

The present invention contemplates an expression vector comprising a polynucleotide of the present invention. Thus, in one embodiment an expression vector is an isolated and purified DNA molecule comprising a promoter operatively linked to an coding region that encodes a polypeptide of the present invention, which coding region is operatively linked to a transcription-terminating region, whereby the promoter drives the transcription of the coding region.

As used herein, the term "operatively linked" means that a promoter is connected to an coding region in such a way that the transcription of that coding region is controlled and regulated by that promoter. Means for operatively linking a promoter to a coding region are well known in the art.

5 Promoters that function in bacteria are well known in the art. An exemplary and preferred promoter for the *Bacillus* crystal proteins include the *sigA*, *sigE*, and *sigK* gene promoters. Alternatively, the native, mutagenized, or recombinant crystal protein-encoding gene promoters themselves can be used.

10 Where an expression vector of the present invention is to be used to transform a plant, a promoter is selected that has the ability to drive expression in plants. Promoters that function in plants are also well known in the art. Useful in expressing the polypeptide in plants are promoters that are inducible, viral, synthetic, constitutive as described (Poszkowski *et al.*, 1989; Odell *et al.*, 1985), and temporally regulated, spatially regulated, and spatio-temporally regulated (Chau *et al.*, 1989).

15 A promoter is also selected for its ability to direct the transformed plant cell's or transgenic plant's transcriptional activity to the coding region. Structural genes can be driven by a variety of promoters in plant tissues. Promoters can be near-constitutive, such as the CaMV 35S promoter, or tissue-specific or developmentally specific promoters affecting dicots or monocots.

20 Where the promoter is a near-constitutive promoter such as CaMV 35S, increases in polypeptide expression are found in a variety of transformed plant tissues (*e.g.*, callus, leaf, seed and root). Alternatively, the effects of transformation can be directed to specific plant tissues by using plant integrating vectors containing a tissue-specific promoter.

25 An exemplary tissue-specific promoter is the lectin promoter, which is specific for seed tissue. The Lectin protein in soybean seeds is encoded by a single gene (*Lel*) that is only expressed during seed maturation and accounts for about 2 to about 5% of total seed mRNA. The lectin gene and seed-specific promoter have been fully characterized and used to direct seed specific expression in transgenic tobacco plants (Vodkin *et al.*, 1983; 30 Lindstrom *et al.*, 1990.)

An expression vector containing a coding region that encodes a polypeptide of interest is engineered to be under control of the lectin promoter and that vector is introduced into plants using, for example, a protoplast transformation method. (Dhir *et al.*, 1991). The expression of the polypeptide is directed specifically to the seeds of the transgenic plant.

A transgenic plant of the present invention produced from a plant cell transformed with a tissue specific promoter can be crossed with a second transgenic plant developed from a plant cell transformed with a different tissue specific promoter to produce a hybrid transgenic plant that shows the effects of transformation in more than one specific tissue.

Exemplary tissue-specific promoters are corn sucrose synthetase 1 (Yang *et al.*, 1990), corn alcohol dehydrogenase 1 (Vogel *et al.*, 1989), corn light harvesting complex (Simpson, 1986), corn heat shock protein (Odell *et al.*, 1985), pea small subunit RuBP carboxylase (Poulsen *et al.*, 1986; Cashmore *et al.*, 1983), Ti plasmid mannopine synthase (Langridge *et al.*, 1989), Ti plasmid nopaline synthase (Langridge *et al.*, 1989), petunia chalcone isomerase (Van Tunen *et al.*, 1988), bean glycine rich protein 1 (Keller *et al.*, 1989), CaMV 35s transcript (Odell *et al.*, 1985) and Potato patatin (Wenzler *et al.*, 1989). Preferred promoters are the cauliflower mosaic virus (CaMV 35S) promoter and the S-E9 small subunit RuBP carboxylase promoter.

The choice of which expression vector and ultimately to which promoter a polypeptide coding region is operatively linked depends directly on the functional properties desired, *e.g.*, the location and timing of protein expression, and the host cell to be transformed. These are well known limitations inherent in the art of constructing recombinant DNA molecules. However, a vector useful in practicing the present invention is capable of directing the expression of the polypeptide coding region to which it is operatively linked.

Typical vectors useful for expression of genes in higher plants are well known in the art and include vectors derived from the tumor-inducing (Ti) plasmid of *Agrobacterium tumefaciens* described (Rogers *et al.*, 1987). However, several other plant integrating vector systems are known to function in plants including pCaMVCN transfer

control vector described (Fromm *et al.*, 1985). Plasmid pCaMVCN (available from Pharmacia, Piscataway, NJ) includes the cauliflower mosaic virus CaMV 35S promoter.

5 In preferred embodiments, the vector used to express the polypeptide includes a selection marker that is effective in a plant cell, preferably a drug resistance selection marker. One preferred drug resistance marker is the gene whose expression results in kanamycin resistance; *i.e.*, the chimeric gene containing the nopaline synthase promoter, Tn5 neomycin phosphotransferase II (*nptII*) and nopaline synthase 3' non-translated region described (Rogers *et al.*, 1988).

10 RNA polymerase transcribes a coding DNA sequence through a site where polyadenylation occurs. Typically, DNA sequences located a few hundred base pairs downstream of the polyadenylation site serve to terminate transcription. Those DNA sequences are referred to herein as transcription-termination regions. Those regions are required for efficient polyadenylation of transcribed messenger RNA (mRNA).

15 Means for preparing expression vectors are well known in the art. Expression (transformation vectors) used to transform plants and methods of making those vectors are described in U. S. Patent Nos. 4,971,908, 4,940,835, 4,769,061 and 4,757,011, the disclosures of which are each specifically incorporated herein by reference. Those vectors can be modified to include a coding sequence in accordance with the present invention.

20 A variety of methods has been developed to operatively link DNA to vectors via complementary cohesive termini or blunt ends. For instance, complementary homopolymer tracts can be added to the DNA segment to be inserted and to the vector DNA. The vector and DNA segment are then joined by hydrogen bonding between the complementary homopolymeric tails to form recombinant DNA molecules.

25 A coding region that encodes a polypeptide having the ability to confer insecticidal activity to a cell is preferably a chimeric *B. thuringiensis* crystal protein-encoding gene. In preferred embodiments, such a polypeptide has the amino acid residue sequence of SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, or SEQ ID NO:30; or a functional equivalent of one or more of those
30 sequences. In accordance with such embodiments, a coding region comprising the DNA

sequence of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, or SEQ ID NO:29 is also preferred.

5.6 Transformed Or Transgenic Plant Cells

5 A bacterium, a yeast cell, or a plant cell or a plant transformed with an expression vector of the present invention is also contemplated. A transgenic bacterium, yeast cell, plant cell or plant derived from such a transformed or transgenic cell is also contemplated. Means for transforming bacteria and yeast cells are well known in the art. Typically, means of transformation are similar to those well known means used to transform other bacteria or yeast such as *E. coli* or *Saccharomyces cerevisiae*.

10 Methods for DNA transformation of plant cells include *Agrobacterium*-mediated plant transformation, protoplast transformation, gene transfer into pollen, injection into reproductive organs, injection into immature embryos and particle bombardment. Each of these methods has distinct advantages and disadvantages. Thus, one particular method of introducing genes into a particular plant strain may not necessarily be the most effective for another plant strain, but it is well known which methods are useful for a particular plant strain.

There are many methods for introducing transforming DNA segments into cells, but not all are suitable for delivering DNA to plant cells. Suitable methods are believed to include virtually any method by which DNA can be introduced into a cell, such as by *Agrobacterium* infection, direct delivery of DNA such as, for example, by PEG-mediated transformation of protoplasts (Omirulleh *et al.*, 1993), by desiccation/inhibition-mediated DNA uptake, by electroporation, by agitation with silicon carbide fibers, by acceleration of DNA coated particles, *etc.* In certain embodiments, acceleration methods are preferred and include, for example, microprojectile bombardment and the like.

25 Technology for introduction of DNA into cells is well-known to those of skill in the art. Four general methods for delivering a gene into cells have been described: (1) chemical methods (Graham and van der Eb, 1973); (2) physical methods such as microinjection (Capecchi, 1980), electroporation (Wong and Neumann, 1982; Fromm *et al.*, 1985) and the gene gun (Johnston and Tang, 1994; Fynan *et al.*, 1993); (3) viral

vectors (Clapp, 1993; Lu *et al.*, 1993; Eglitis and Anderson, 1988a, 1988b); and (4) receptor-mediated mechanisms (Curiel *et al.*, 1991; 1992; Wagner *et al.*, 1992).

5.6.1 Electroporation

5 The application of brief, high-voltage electric pulses to a variety of animal and plant cells leads to the formation of nanometer-sized pores in the plasma membrane. DNA is taken directly into the cell cytoplasm either through these pores or as a consequence of the redistribution of membrane components that accompanies closure of the pores. Electroporation can be extremely efficient and can be used both for transient
10 expression of clones genes and for establishment of cell lines that carry integrated copies of the gene of interest. Electroporation, in contrast to calcium phosphate-mediated transfection and protoplast fusion, frequently gives rise to cell lines that carry one, or at most a few, integrated copies of the foreign DNA.

 The introduction of DNA by means of electroporation, is well-known to those of
15 skill in the art. In this method, certain cell wall-degrading enzymes, such as pectin-degrading enzymes, are employed to render the target recipient cells more susceptible to transformation by electroporation than untreated cells. Alternatively, recipient cells are made more susceptible to transformation, by mechanical wounding. To effect transformation by electroporation one may employ either friable tissues such as a
20 suspension culture of cells, or embryogenic callus, or alternatively, one may transform immature embryos or other organized tissues directly. One would partially degrade the cell walls of the chosen cells by exposing them to pectin-degrading enzymes (pectolyases) or mechanically wounding in a controlled manner. Such cells would then be recipient to DNA transfer by electroporation, which may be carried out at this stage, and transformed
25 cells then identified by a suitable selection or screening protocol dependent on the nature of the newly incorporated DNA.

5.6.2 Microprojectile Bombardment

 A further advantageous method for delivering transforming DNA segments to
30 plant cells is microprojectile bombardment. In this method, particles may be coated with

nucleic acids and delivered into cells by a propelling force. Exemplary particles include those comprised of tungsten, gold, platinum, and the like.

5 An advantage of microprojectile bombardment, in addition to it being an effective means of reproducibly stably transforming monocots, is that neither the isolation of protoplasts (Cristou *et al.*, 1988) nor the susceptibility to *Agrobacterium* infection is required. An illustrative embodiment of a method for delivering DNA into maize cells by acceleration is a Biolistics Particle Delivery System, which can be used to propel particles coated with DNA or cells through a screen, such as a stainless steel or Nytex screen, onto a filter surface covered with corn cells cultured in suspension. The screen disperses the particles so that they are not delivered to the recipient cells in large aggregates. It is
10 believed that a screen intervening between the projectile apparatus and the cells to be bombarded reduces the size of projectiles aggregate and may contribute to a higher frequency of transformation by reducing damage inflicted on the recipient cells by projectiles that are too large.

15 For the bombardment, cells in suspension are preferably concentrated on filters or solid culture medium. Alternatively, immature embryos or other target cells may be arranged on solid culture medium. The cells to be bombarded are positioned at an appropriate distance below the macroprojectile stopping plate. If desired, one or more screens are also positioned between the acceleration device and the cells to be
20 bombarded. Through the use of techniques set forth herein one may obtain up to 1000 or more foci of cells transiently expressing a marker gene. The number of cells in a focus which express the exogenous gene product 48 hours post-bombardment often range from 1 to 10 and average 1 to 3.

25 In bombardment transformation, one may optimize the prebombardment culturing conditions and the bombardment parameters to yield the maximum numbers of stable transformants. Both the physical and biological parameters for bombardment are important in this technology. Physical factors are those that involve manipulating the DNA/microprojectile precipitate or those that affect the flight and velocity of either the macro- or microprojectiles. Biological factors include all steps involved in manipulation
30 of cells before and immediately after bombardment, the osmotic adjustment of target cells

to help alleviate the trauma associated with bombardment, and also the nature of the transforming DNA, such as linearized DNA or intact supercoiled plasmids. It is believed that pre-bombardment manipulations are especially important for successful transformation of immature embryos.

5 Accordingly, it is contemplated that one may wish to adjust various of the bombardment parameters in small scale studies to fully optimize the conditions. One may particularly wish to adjust physical parameters such as gap distance, flight distance, tissue distance, and helium pressure. One may also minimize the trauma reduction factors (TRFs) by modifying conditions which influence the physiological state of the
10 recipient cells and which may therefore influence transformation and integration efficiencies. For example, the osmotic state, tissue hydration and the subculture stage or cell cycle of the recipient cells may be adjusted for optimum transformation. The execution of other routine adjustments will be known to those of skill in the art in light of the present disclosure.

15 The methods of particle-mediated transformation is well-known to those of skill in the art. U. S. Patent No. 5,015,580 (specifically incorporated herein by reference) describes the transformation of soybeans using such a technique.

5.6.3 *Agrobacterium*-Mediated Transfer

20 *Agrobacterium*-mediated transfer is a widely applicable system for introducing genes into plant cells because the DNA can be introduced into whole plant tissues, thereby bypassing the need for regeneration of an intact plant from a protoplast. The use of *Agrobacterium*-mediated plant integrating vectors to introduce DNA into plant cells is well known in the art. See, for example, the methods described (Fraley *et al.*, 1985;
25 Rogers *et al.*, 1987). The genetic engineering of cotton plants using *Agrobacterium*-mediated transfer is described in U. S. Patent No. 5,004,863 (specifically incorporated herein by reference), while the transformation of lettuce plants is described in U. S. Patent No. 5,349,124 (specifically incorporated herein by reference). Further, the integration of the Ti-DNA is a relatively precise process resulting in few rearrangements. The region of
30 DNA to be transferred is defined by the border sequences, and intervening DNA is

usually inserted into the plant genome as described (Spielmann *et al.*, 1986; Jorgensen *et al.*, 1987).

Modern *Agrobacterium* transformation vectors are capable of replication in *E. coli* as well as *Agrobacterium*, allowing for convenient manipulations as described (Klee *et al.*, 1985). Moreover, recent technological advances in vectors for *Agrobacterium*-mediated gene transfer have improved the arrangement of genes and restriction sites in the vectors to facilitate construction of vectors capable of expressing various polypeptide coding genes. The vectors described (Rogers *et al.*, 1987), have convenient multi-linker regions flanked by a promoter and a polyadenylation site for direct expression of inserted polypeptide coding genes and are suitable for present purposes. In addition, *Agrobacterium* containing both armed and disarmed Ti genes can be used for the transformations. In those plant strains where *Agrobacterium*-mediated transformation is efficient, it is the method of choice because of the facile and defined nature of the gene transfer.

Agrobacterium-mediated transformation of leaf disks and other tissues such as cotyledons and hypocotyls appears to be limited to plants that *Agrobacterium* naturally infects. *Agrobacterium*-mediated transformation is most efficient in dicotyledonous plants. Few monocots appear to be natural hosts for *Agrobacterium*, although transgenic plants have been produced in asparagus using *Agrobacterium* vectors as described (Bytebier *et al.*, 1987). Therefore, commercially important cereal grains such as rice, corn, and wheat must usually be transformed using alternative methods. However, as mentioned above, the transformation of asparagus using *Agrobacterium* can also be achieved (see, for example, Bytebier *et al.*, 1987).

A transgenic plant formed using *Agrobacterium* transformation methods typically contains a single gene on one chromosome. Such transgenic plants can be referred to as being heterozygous for the added gene. However, inasmuch as use of the word "heterozygous" usually implies the presence of a complementary gene at the same locus of the second chromosome of a pair of chromosomes, and there is no such gene in a plant containing one added gene as here, it is believed that a more accurate name for such a

plant is an independent segregant, because the added, exogenous gene segregates independently during mitosis and meiosis.

5 More preferred is a transgenic plant that is homozygous for the added structural gene; i.e., a transgenic plant that contains two added genes, one gene at the same locus on each chromosome of a chromosome pair. A homozygous transgenic plant can be obtained by sexually mating (selfing) an independent segregant transgenic plant that contains a single added gene, germinating some of the seed produced and analyzing the resulting plants produced for enhanced carboxylase activity relative to a control (native, non-transgenic) or an independent segregant transgenic plant.

10 It is to be understood that two different transgenic plants can also be mated to produce offspring that contain two independently segregating added, exogenous genes. Selfing of appropriate progeny can produce plants that are homozygous for both added, exogenous genes that encode a polypeptide of interest. Back-crossing to a parental plant and out-crossing with a non-transgenic plant are also contemplated.

15 Transformation of plant protoplasts can be achieved using methods based on calcium phosphate precipitation, polyethylene glycol treatment, electroporation, and combinations of these treatments (see, e.g., Potrykus *et al.*, 1985; Lorz *et al.*, 1985; Fromm *et al.*, 1985; Uchimiya *et al.*, 1986; Callis *et al.*, 1987; Marcotte *et al.*, 1988).

20 Application of these systems to different plant strains depends upon the ability to regenerate that particular plant strain from protoplasts. Illustrative methods for the regeneration of cereals from protoplasts are described (Fujimura *et al.*, 1985; Toriyama *et al.*, 1986; Yamada *et al.*, 1986; Abdullah *et al.*, 1986).

25 To transform plant strains that cannot be successfully regenerated from protoplasts, other ways to introduce DNA into intact cells or tissues can be utilized. For example, regeneration of cereals from immature embryos or explants can be effected as described (Vasil, 1988). In addition, "particle gun" or high-velocity microprojectile technology can be utilized (Vasil, 1992).

Using that latter technology, DNA is carried through the cell wall and into the cytoplasm on the surface of small metal particles as described (Klein *et al.*, 1987; Klein *et*

al., 1988; McCabe *et al.*, 1988). The metal particles penetrate through several layers of cells and thus allow the transformation of cells within tissue explants.

5.7 Production of Insect-Resistant Transgenic Plants

5 Thus, the amount of a gene coding for a polypeptide of interest (*i.e.*, a bacterial crystal protein or polypeptide having insecticidal activity against one or more insect species) can be increased in plant such as corn by transforming those plants using particle bombardment methods (Maddock *et al.*, 1991). By way of example, an expression vector containing a coding region for a *B. thuringiensis* crystal protein and an appropriate
10 selectable marker is transformed into a suspension of embryonic maize (corn) cells using a particle gun to deliver the DNA coated on microprojectiles. Transgenic plants are regenerated from transformed embryonic calli that express the disclosed insecticidal crystal proteins. Particle bombardment has been used to successfully transform wheat (Vasil *et al.*, 1992).

15 DNA can also be introduced into plants by direct DNA transfer into pollen as described (Zhou *et al.*, 1983; Hess, 1987; Luo *et al.*, 1988). Expression of polypeptide coding genes can be obtained by injection of the DNA into reproductive organs of a plant as described (Pena *et al.*, 1987). DNA can also be injected directly into the cells of immature embryos and the rehydration of desiccated embryos as described (Neuhaus *et al.*, 1987; Benbrook *et al.*, 1986).

20 The development or regeneration of plants from either single plant protoplasts or various explants is well known in the art (Weissbach and Weissbach, 1988). This regeneration and growth process typically includes the steps of selection of transformed cells, culturing those individualized cells through the usual stages of embryonic
25 development through the rooted plantlet stage. Transgenic embryos and seeds are similarly regenerated. The resulting transgenic rooted shoots are thereafter planted in an appropriate plant growth medium such as soil.

The development or regeneration of plants containing the foreign, exogenous gene that encodes a polypeptide of interest introduced by *Agrobacterium* from leaf explants
30 can be achieved by methods well known in the art such as described (Horsch *et al.*, 1985).

5 In this procedure, transformants are cultured in the presence of a selection agent and in a medium that induces the regeneration of shoots in the plant strain being transformed as described (Fraley *et al.*, 1983). In particular, U. S. Patent No. 5,349,124 details the creation of genetically transformed lettuce cells and plants resulting therefrom which express hybrid crystal proteins conferring insecticidal activity against Lepidopteran larvae to such plants.

10 This procedure typically produces shoots within two to four months and those shoots are then transferred to an appropriate root-inducing medium containing the selective agent and an antibiotic to prevent bacterial growth. Shoots that rooted in the presence of the selective agent to form plantlets are then transplanted to soil or other media to allow the production of roots. These procedures vary depending upon the particular plant strain employed, such variations being well known in the art.

15 Preferably, the regenerated plants are self-pollinated to provide homozygous transgenic plants, as discussed before. Otherwise, pollen obtained from the regenerated plants is crossed to seed-grown plants of agronomically important, preferably inbred lines. Conversely, pollen from plants of those important lines is used to pollinate regenerated plants. A transgenic plant of the present invention containing a desired polypeptide is cultivated using methods well known to one skilled in the art.

20 A transgenic plant of this invention thus has an increased amount of a coding region (*e.g.*, a *cry* gene) that encodes the Cry polypeptide of interest. A preferred transgenic plant is an independent segregant and can transmit that gene and its activity to its progeny. A more preferred transgenic plant is homozygous for that gene, and transmits that gene to all of its offspring on sexual mating. Seed from a transgenic plant may be grown in the field or greenhouse, and resulting sexually mature transgenic plants are self-pollinated to generate true breeding plants. The progeny from these plants become true breeding lines that are evaluated for, by way of example, increased insecticidal capacity against Coleopteran insects, preferably in the field, under a range of environmental conditions. The inventors contemplate that the present invention will find particular utility in the creation of transgenic corn, wheat, oats, barley, other grains, vegetables, fruits, fruit trees, berries, turf grass, ornamentals, shrubs and trees.

30

6. EXAMPLES

The following examples are included to demonstrate preferred embodiments of the invention. It should be appreciated by those of skill in the art that the techniques disclosed in the examples which follow represent techniques discovered by the inventor to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those of skill in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the spirit and scope of the invention.

6.1 Example 1 -- Construction of Hybrid *B. thuringiensis* δ -Endotoxins

The *B. thuringiensis* shuttle vectors pEG853, pEG854, and pEG857 which are used in the present invention are described by Baum *et al.*, 1990. The plasmid pEG857 contains the *CryI*Ac gene cloned into pEG853 as an *SphI*-*Bam*HI DNA fragment. pEG1064 was constructed for purposes of the present invention in such a way that the *KpnI* site within the *CryI*Ac gene was preserved and the *KpnI* site in the pEG857 multiple cloning site (MCS) was eliminated. This was accomplished by sequentially subjecting pEG857 DNA to limited *KpnI* digestion so that only one *KpnI* site is cut, filling in the *KpnI* 5' overhang by Klenow fragment of DNA polymerase I to create blunt DNA ends, and joining the blunt ends of DNA by T4 DNA ligase. pEG318 contains the *CryI*F gene (Chambers *et al.*, 1991) cloned into the *XhoI* site of pEG854 as a *XhoI*-*SalI* DNA fragment. pEG315 contains the *CryI*C gene from strain EG6346 (Chambers *et al.*, 1991) cloned into the *XhoI*-*Bam*HI sites of pEG854 as a *SalI*-*Bam*HI DNA fragment.

FIG. 1A shows a schematic representation of the DNA encoding the complete *CryI*Ac, *CryI*Ab, *CryI*C, and *CryI*F genes contained on pEG854/pEG1064, pEG20, pEG315, and pEG318, respectively. Unique restriction sites that were used in constructing certain hybrid genes are also shown. FIG. 1B shows a schematic representation of hybrid genes pertaining to the present invention. In some cases standard PCRTM amplification with mutagenic oligonucleotide primers were used to incorporate

appropriate restriction sites into DNA fragments used for hybrid gene construction. Certain hybrid gene constructions could not be accomplished by restriction fragment subcloning. In those instances, PCR™ overlap extension (POE) was used to construct the desired hybrid gene (Horton *et al.*, 1989). The following oligonucleotide primers (purchased from Integrated DNA Technologies Inc., Coralville, IA) were used:

Primer A

5'-GGATAGCACTCATCAAAGGTACC-3' (SEQ ID NO:1)

Primer B

5'-GAAGATATCCAATTCGAACAGTTTCCC-3' (SEQ ID NO:2)

Primer C

5'-CATATTCTGCCTCGAGTGTTGCAGTAAC-3' (SEQ ID NO:3)

Primer D

5'-CCCGATCGGCCGCATGC-3' (SEQ ID NO:4)

Primer E

5'-CATTGGAGCTCTCCATG-3' (SEQ ID NO:5)

Primer F

5'-GCACTACGATGTATCC-3' (SEQ ID NO:6)

Primer G

5'-CATCGTAGTGCAACTCTTAC-3' (SEQ ID NO:7)

Primer H

5'-CCAAGAAAATACTAGAGCTCTTGTTAAAAAAGGTGTTCC-3' (SEQ ID NO:8)

Primer I

5'-ATTTGAGTAATACTATCC-3' (SEQ ID NO:23)

Primer J

5'-ATTACTCAAATACCATTGG-3' (SEQ ID NO:24)

The plasmids described in FIG. 1B containing the hybrid δ -endotoxin genes pertinent to this invention are described below. Isolation or purification of DNA fragments generated by restriction of plasmid DNA, PCR™ amplification, or POE refers

to the sequential application of agarose-TAE gel electrophoresis and use of the GeneClean Kit (Bio 101) following the manufacturer's recommendation. pEG1065 was constructed by PCR™ amplification of the *CryIF* DNA fragment using primer pair A and B and pEG318 as the DNA template. The resulting PCR™ product was isolated, cut with *AsuII* and *KpnI*, and used to replace the corresponding *AsuII-KpnI* DNA fragment in pEG857. 5 Plasmid pEG1067 was constructed using POE and DNA fragments *SauI-KpnI* of *CryIF* and *AsuII-ClaI* of *CryIAC* that were isolated from pEG318 and pEG857, respectively. The resulting POE product was PCR™ amplified with primer pair A and B, cut with *AsuII* and *KpnI*, and used to replace the corresponding *AsuII-KpnI* fragment in pEG857.

10 pEG1068 was constructed by replacing the *SacI-KpnI* DNA fragment of *CryIAC* isolated from pEG857 with the corresponding *SacI-KpnI* DNA fragment isolated from *CryIF* (pEG318). pEG1070 was constructed by replacing the *SacI-KpnI* DNA fragment isolated from pEG1065 with the corresponding *SacI-KpnI* DNA fragment isolated from *CryIAC* (pEG857). pEG1072 was constructed by replacing the *SacI-KpnI* DNA fragment 15 isolated from pEG1067 with the corresponding *SacI-KpnI* DNA fragment isolated from *CryIAC* (pEG857). pEG1074, pEG1076, and pEG1077 were constructed by replacing the *SphI-XhoI* DNA fragment from pEG1064 with the PCR™ amplified *SphI-XhoI* DNA fragment from pEG1065, pEG1067, pEG1068, respectively, using primer pairs C and D. pEG1089 was constructed by replacing the *SphI-SacI* DNA fragment of pEG1064 with 20 the isolated and *SphI* and *SacI* cut PCR™ product of *CryIF* that was generated using primer pair D and E and the template pEG318.

pEG1091 was constructed by replacing the *SphI-SacI* DNA fragment of pEG1064 with the isolated and *SphI* and *SacI* cut PCR™ product of *CryIC* that was generated using primer pair D and H and the template pEG315.

25 pEG1088 was constructed by POE using a *CryIAC* DNA fragment generated using primer pair B and F and a *CryIC* DNA fragment generated using primer pair A and G. The *SacI-KpnI* fragment was isolated from the resulting POE product and used to replace the corresponding *SacI-KpnI* fragment in pEG1064.

30 pEG365 was constructed by first replacing the *SphI-KpnI* DNA fragment from pEG1065 with the corresponding *CryIAb* DNA fragment isolated from pEG20 to give

pEG364. The *SacI-KpnI* DNA fragment from pEG364 was then replaced with the corresponding *CryIF* DNA fragment isolated from pEG318.

pEG1092 was constructed by replacing the *KpnI-BamHI* DNA fragment from pEG1088 with the corresponding DNA fragment isolated from pEG315. pEG1092 is distinct from the *cryIAb/cryIC* hybrid δ -endotoxin gene disclosed in Intl. Pat. Appl. Publ. No. WO 95/06730.

pEG1093 was constructed by replacing the *SphI-AsuII* DNA fragment from pEG1068 with the corresponding *SphI-AsuII* DNA fragment isolated from pEG20.

pEG378 was constructed by POE using a *cryIAC* DNA fragment generated using primer pair B and I using pEG857 as the template and a *cryIF* DNA fragment generated using primer pair A and J using pEG318 as the template. The resulting POE product was cut with *AsuII* and *KpnI* and the resulting isolated DNA fragment used to replace the corresponding *AsuII-KpnI* DNA fragment in pEG1064.

6.2 Example 2 -- Production of the Hybrid Toxins In *B. thuringiensis*

The plasmids encoding the hybrid toxins described in Example 1 were transformed into *B. thuringiensis* as described by Mettus and Macaluso, 1990. The resulting *B. thuringiensis* strains were grown in 50 ml of C-2 medium until the culture was fully sporulated and lysed (approximately 48 hr.). Since crystal formation is a prerequisite for efficient commercial production of δ -endotoxins in *B. thuringiensis*, microscopic analysis was used to identify crystals in the sporulated cultures (TABLE 3).

TABLE 3
Crystal Formation by the Hybrid δ -Endotoxins

Strain	NRRL ^a Accession Number	NRRL Deposit Date	Plasmid	Parent δ -Endotoxins	Crystal Formation
EG11060			pEG1065	Cry1Ac + Cry1F	+
EG11062			pEG1067	Cry1Ac + Cry1F	+
EG11063	B-21579	May 24, 1996	pEG1068	Cry1Ac + Cry1F	+
EG11065			pEG1070	Cry1Ac + Cry1F	-
EG11067			pEG1072	Cry1Ac + Cry1F	-
EG11071			pEG1074	Cry1Ac + Cry1F	+
EG11073			pEG1076	Cry1Ac + Cry1F	+
EG11074	B-21580	May 24, 1996	pEG1077	Cry1Ac + Cry1F	+
EG11087			pEG1088	Cry1Ac + Cry1C	-
EG11088			pEG1089	Cry1F + Cry1Ac	-
EG11090			pEG1091	Cry1C + Cry1Ac	-
EG11091			pEG1092	Cry1Ac + Cry1C	+
EG11092	B-21635	Oct. 21, 1996	pEG1093	Cry1Ab + Cry1Ac + Cry1F	+
EG11735	B-21581	May 24, 1996	pEG365	Cry1Ab + Cry1F + Cry1Ac	+
EG11751	B-21636	Oct. 21, 1996	pEG378	Cry1Ac + Cry1F	+

^aThe subject cultures have been deposited under conditions that assure that access to the cultures will be available during the pendency of this patent application to one determined by the Commissioner of Patents and Trademarks to be entitled thereto under 37 C.F.R. §1.14 and 35 U.S.C. §122. The deposits are available as required by foreign patent laws in countries wherein counterparts of the subject application, or its progeny, are filed. However, it should be understood that the availability of a deposit does not constitute a license to practice the subject invention in derogation of patent rights granted by governmental action. The subject culture deposits will be stored and made available to the public in accord with the provisions of the Budapest Treaty for the Deposit of Microorganisms, *i.e.*, they will be stored with all the care necessary to keep them viable and uncontaminated for a period of at least five years after the most recent request for the finishing of a sample of the deposit, and in any case, for a period of at least 30 (thirty) years after the date of deposit or for the enforceable life of any patent which may issue disclosing the cultures. The depositor acknowledges the duty to replace the deposits should the depository be unable to furnish a sample when requested, due to the condition of the deposits.

All restrictions on the availability to the public of the subject culture deposits will be irrevocably removed upon the granting of a patent disclosing them.

Cultures shown in Table 3 were deposited in the permanent collection of the Agricultural Research Service Culture Collection, Northern Regional Research Laboratory (NRRL), located at 1815 N. University Street, Peoria, IL 61604, under the terms of the Budapest Treaty.--

The δ -endotoxin production for some of the *B. thuringiensis* strains specified in TABLE 3 was examined by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) as described by Baum *et al.*, 1990. Equal volume cultures of each *B. thuringiensis* strain were grown in C-2 medium until fully sporulated and lysed. The cultures were centrifuged and the spore/crystal pellet was washed twice with equal volumes of distilled deionized water. The final pellet was suspended in half the culture volume of 0.005% Triton X-100[®]. An equal volume of each washed culture was analyzed by SDS-PAGE as shown in FIG. 2.

The majority of hybrids involving Cry1Ac and Cry1F formed stable crystals in *B. thuringiensis*. A notable exception is EG11088 in which the active toxin fragment would be the reciprocal exchange of EG11063. Two of the three hybrids involving Cry1Ac and Cry1C, EG11087 and EG11090, failed to produce crystal in *B. thuringiensis* even though these reciprocal hybrids mimic the activated toxin fragments of crystal-forming EG11063 and EG11074.

Every strain that was examined by SDS-PAGE produced some level of δ -endotoxin. As expected, however, those cultures identified as crystal negative produced very little protein (*e.g.*, lane e: EG11065, lane f: EG11067, lane j: EG11088, and lane k: EG11090). For reference, typical yields from a crystal forming δ -endotoxin is shown for Cry1Ac (lane a). Several hybrid δ -endotoxins produce comparable levels of protein including EG11060 (lane b), EG11062 (lane c), EG11063 (lane d; SEQ ID NO:10), and EG11074 (lane i; SEQ ID NO:12). The data clearly show that efficient hybrid δ -endotoxin production in *B. thuringiensis* is unpredictable and varies depending on the parent δ -endotoxins used to construct the hybrid.

6.3 Example 3 -- Proteolytic Processing of the Hybrid δ -Endotoxins

Proteolytic degradation of the protoxin form of the δ -endotoxin to a stable active toxin occurs once δ -endotoxin crystals are solubilized in the larval midgut. One measure of the

potential activity of δ -endotoxins is the stability of the active δ -endotoxin in a proteolytic environment. To test the proteolytic sensitivity of the hybrid δ -endotoxins, solubilized toxin was subjected to trypsin digestion. The δ -endotoxins were purified from sporulated *B. thuringiensis* cultures and quantified as described by Chambers *et al.*, 1991. Exactly 250 μ g of each hybrid δ -endotoxin crystal was solubilized in 30 mM NaHCO₃, 10 mM DTT (total volume 0.5 ml). Trypsin was added to the solubilized toxin at a 1:10 ratio. At appropriate time points 50 μ l aliquots were removed to 50 μ l Laemmli buffer, heated to 100°C for 3 min., and frozen in a dry-ice ethanol bath for subsequent analysis. The trypsin digests of the solubilized toxins were analyzed by SDS-PAGE and the amount of active δ -endotoxin at each time point was quantified by densitometry. A graphic representation of the results from these studies are shown in FIG. 3.

The wild-type Cry1Ac is rapidly processed to the active δ -endotoxin fragment that is stable for the duration of the study. The hybrid δ -endotoxins from EG11063 and EG11074 are also processed to active δ -endotoxin fragments which are stable for the duration of the study. The processing of the EG11063 δ -endotoxin occurs at a slower rate and a higher percentage of this active δ -endotoxin fragment remains at each time point. Although the hybrid δ -endotoxins from EG11060 and EG11062 are processed to active δ -endotoxin fragments, these fragments are more susceptible to further cleavage and degrade at various rates during the course of the study. The 5' exchange points between *Cry1Ac* and *Cry1F* for the EG11062 and EG11063 δ -endotoxins result in toxins that differ by only 21 amino acid residues (see FIG. 1). However, the importance of maintaining *Cry1Ac* sequences at these positions is evident by the more rapid degradation of the EG11062 δ -endotoxin. These data demonstrate that different hybrid δ -endotoxins constructed using the same parental δ -endotoxins can vary significantly in biochemical characteristics such as proteolytic stability.

6.4 Example 4 -- Bioactivity of the Hybrid δ -Endotoxins

B. thuringiensis cultures expressing the desired δ -endotoxin were grown until fully sporulated and lysed and washed as described in Example 2. The δ -endotoxin levels for each culture were quantified by SDS-PAGE as described by Baum *et al.*, 1990. In the case of bioassay screens, a single appropriate concentration of each washed δ -endotoxin culture was

topically applied to 32 wells containing 1.0 ml artificial diet per well (surface area of 175 mm²). A single neonate larvae was placed in each of the treated wells and the tray covered by a clear perforated mylar sheet. Larvae mortality was scored after 7 days of feeding and percent mortality expressed as the ratio of the number of dead larvae to the total number of larvae treated (32).

In the case of LC₅₀ determinations (δ -endotoxin concentration giving 50% mortality), δ -endotoxins were purified from the *B. thuringiensis* cultures and quantified as described by Chambers *et al.*, 1991. Eight concentrations of the δ -endotoxins were prepared by serial dilution in 0.005% Triton X-100[®] and each concentration was topically applied to wells containing 1.0 ml of artificial diet. Larvae mortality was scored after 7 days of feeding (32 larvae for each δ -endotoxin concentration). In all cases the diluent served as the control.

A comparison of the Cry1A/Cry1F hybrid toxins by bioassay screens is shown in TABLE 4. The hybrid δ -endotoxins from strains EG11063 and EG11074 maintain the activities of the parental Cry1Ac and Cry1F δ -endotoxins. Furthermore, the hybrid δ -endotoxin from EG11735 maintains the activity of its parental Cry1Ab and Cry1F δ -endotoxins. The δ -endotoxins produced by strains EG11061, EG11062, EG11071, and EG11073 have no insecticidal activity on the insect larvae tested despite 1) being comprised of at least one parental δ -endotoxin that is active against the indicated larvae and 2) forming stable, well-defined crystals in *B. thuringiensis*. These results demonstrate the unpredictable nature of hybrid toxin constructions.

For the data in TABLE 4. All strains were tested as washed sporulated cultures. For each insect tested, equivalent amounts of δ -endotoxins were used and insecticidal activity was based on the strain showing the highest percent mortality (++++).

TABLE 4
Bioassay Screens of Hybrid Cry1A/Cry1F δ -Endotoxins

Strain	<i>S. frugiperda</i>	<i>S. exigua</i>	<i>H. virescens</i>	<i>H. zea</i>	<i>O. nubilalis</i>
Cry1Ac	-	-	++++	++++	+++
Cry1F	++++	++	++	++	++
Cry1Ab	++	+	+++	++	+++
EG11060	-	-	-	-	-
EG11062	-	-	-	-	-
EG11063	++++	++++	+++	+++	++++
EG11071	-	-	-	-	-
EG11073	-	-	-	-	-
EG11074	++++	++++	+++	+++	++++
EG11090	-	+++	-	-	-
EG11091	++++	++++	-	-	N.D. ^a
EG11092	++++	++++	+++	+++	N.D. ^a
EG11735	++++	++++	+++	+++	N.D. ^a
EG11751	N.D. ^a	++++	N.D. ^a	++++	N.D. ^a

^aN.D. = not determined.

5 The δ -endotoxins described in FIG. 1 and that demonstrated insecticidal activity in bioassay screens were tested as purified crystals to determine their LC₅₀ (see TABLE 5). The δ -endotoxins purified from strains EG11063, EG11074, EG11091, and EG11735 all show increased armyworm (*S. frugiperda* and *S. exigua*) activity compared to any of the wild-type δ -endotoxins tested. The EG11063 and EG11074 δ -endotoxins would yield identical active toxin fragments (refer to FIG. 1B) which is evident by their similar LC₅₀ values on the insects examined. An unexpected result evident from these data is that a hybrid δ -endotoxin such as EG11063, EG11092, EG11074, EG11735, or EG11751 can retain the activity of their respective parental δ -endotoxins, and, against certain insects such as *S. exigua*, can have activity far better than either parental δ -endotoxin. This broad range of insecticidal activity at doses close to or lower than the parental δ -endotoxins,

5 along with the wild-type level of toxin production (see Example 2), make these proteins particularly suitable for production in *B. thuringiensis*. Although the EG11091 derived δ -endotoxin has better activity against *S. frugiperda* and *S. exigua* than its parental δ -endotoxins, it has lost the *H. virescens* and *H. zea* activity attributable to its Cry1Ac parent. This restricted host range along with lower toxin yield observed for the EG11091 δ -endotoxin (see Example 2) make it less amenable to production in *B. thuringiensis*

TABLE 5
LC₅₀ Values for the Purified Hybrid δ -Endotoxin^a

Toxin	<i>S. frugiperda</i>	<i>S. exigua</i>	<i>H. virescens</i>	<i>H. zea</i>	<i>O. nubilalis</i>
Cry1Ac	>10000	>10000	9	100	23
Cry1Ab	1435	4740	118	400	17
Cry1C	>10000	490	>10000	>10000	>10000
Cry1F	1027	3233	54	800	51
EG11063	550	114	33	80	7
(Cry1Ac/1F)					
EG11074	468	77	25	76	9
(Cry1Ac/1F)					
EG11090	21	21	219	>10000	nd
(Cry1Ac/1C)					

10 In TABLE 5, the LC₅₀ values are expressed in nanograms of purified δ -endotoxin per well (175 mm²) and are the composite values for 2 to 6 replications. nd = not determined.

TABLE 6
DNA Exchange Sites for Cry1 Hybrid δ -Endotoxins

Plasmid	SEQ ID N	5' Exchange Site	SEQ ID NO	3' Exchange Site
	O:			
pEG1065	15	TATCCAATTCTGAACGTCATC	21	ACTACCAGGTACCTTTGATG
pEG1067	16	TTTAGTCATCGATTAAATCA	21	ACTACCAGGTACCTTTGATG
pEG1068	17	ATAATAAGAGCTCCAATGTT	21	ACTACCAGGTACCTTTGATG
pEG1070	15	TATCCAATTCTGAACGTCATC	19	TCATGGAGAGCTCCTATGTT
pEG1072	16	TTTAGTCATCGATTAAATCA	19	TCATGGAGAGCTCCTATGTT
pEG1074	15	TATCCAATTCTGAACGTCATC		TGCAACACTCGAGGCTGAAT
pEG1076	16	TTTAGTCATCGATTAAATCA		TGCAACACTCGAGGCTGAAT
pEG1077	17	ATAATAAGAGCTCCAATGTT		TGCAACACTCGAGGCTGAAT
pEG1088	18	TACATCGTAGTGCAACTCTT	22	ACTACCGGGTACCTTTGATA
pEG1089	19	TCATGGAGAGCTCCTATGTT		-
pEG1091	20	TTAACAAGAGCTCCTATGTT		-
pEG1092	18	TACATCGTAGTGCAACTCTT		-
pEG11093		-	21	ACTACCAGGTACCTTTGATG
pEG365	17	ATAATAAGAGCTCCAATGTT	21	ACTACCAGGTACCTTTGATG
pEG378		-	21	ACTACCAGGTACCTTTGATG

TABLE 6 describes the DNA surrounding the 5' and 3' exchange points for the hybrid δ -endotoxins which are pertinent to the present invention. As evident by the SEQ ID NO, certain hybrid δ -endotoxins share exchange sites. In certain instances, the exchange site is indistinguishable from one of the parent δ -endotoxins. This is the case for the 3' exchange site for pEG1074, pEG1076, and pEG1077 and therefore, they have not been assigned a separate sequence identifier (SEQ ID NO.).

To examine the effect of other small changes in the exchange site chosen for hybrid endotoxin construction, the activity of EG11751 and EG11063 on *S. exigua* and *H. zea* were compared (TABLE 7). The data clearly show that hybrid δ -endotoxin improvements can be made by altering the exchange site between the two parental δ -endotoxins. In this example, the exchange site in the EG11751 δ -endotoxin was moved 75 base pairs 3' compared to the EG11063 δ -endotoxin and results in improved insecticidal activity. Although no significant improvement in *S. exigua* activity is observed between EG11063 and EG11751, a significant improvement in *H. zea* activity

of almost 4-fold is observed for EG11751. It is important to note that improvements in hybrid δ -endotoxin bioactivity by altering exchange sites is unpredictable. In the case of EG11062, moving the exchange site 63 base pairs 5' of the EG11063 exchange site abolishes insecticidal activity as shown in TABLE 5.

TABLE 7

Bioactivity of EG11063 and EG11751

B.t. Strain	LC ₅₀ Values for Washed Sporulated Cultures	
	<i>S. exigua</i>	<i>H. zea</i>
EG11063	106	38
EG11751	90	10

6.5 EXAMPLE 5 -- AMINO ACID SEQUENCES OF THE NOVEL CRYSTAL PROTEINS

10 6.5.1 AMINO ACID SEQUENCE OF THE EG11063 CRYSTAL PROTEIN (SEQ ID NO:10)

MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeu
 SerAsnProGluValGluValLeuGlyGlyGluArgIleGluThrGly
 TyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
 GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIle
 15 TrpGlyIlePheGlyProSerGlnTrpAspAlaPheLeuValGlnIle
 GluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
 IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGlu
 SerPheArgGluTrpGluAlaAspProThrAsnProAlaLeuArgGlu
 GluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla
 20 IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerVal
 TyrValGlnAlaAlaAsnLeuHisLeuSerValLeuArgAspValSer
 ValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
 TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspTyrAlaVal
 ArgTrpTyrAsnThrGlyLeuGluArgValTrpGlyProAspSerArg
 25 AspTrpValArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
 LeuAspIleValAlaLeuPheProAsnTyrAspSerArgArgTyrPro
 ileArgThrValSerGlnLeuThrArgGluIleTyrThrAsnProVal

LeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
 ArgSerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThr
 IleTyrThrAspAlaHisArgGlyTyrTyrTyrTrpSerGlyHisGln
 IleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro
 5 LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAla
 GlnLeuGlyGlnGlyValTyrArgThrLeuSerSerThrLeuTyrArg
 ArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp
 GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaVal
 TyrArgLysSerGlyThrValAspSerLeuAspGluIleProProGln
 10 AsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
 ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIle
 ArgAlaProMetPheSerTrpThrHisArgSerAlaThrProThrAsn
 ThrIleAspProGluArgIleThrGlnIleProLeuValLysAlaHis
 ThrLeuGlnSerGlyThrThrValValArgGlyProGlyPheThrGly
 15 GlyAspIleLeuArgArgThrSerGlyGlyProPheAlaTyrThrIle
 ValAsnIleAsnGlyGlnLeuProGlnArgTyrArgAlaArgIleArg
 TyrAlaSerThrThrAsnLeuArgIleTyrValThrValAlaGlyGlu
 ArgIlePheAlaGlyGlnPheAsnLysThrMetAspThrGlyAspPro
 LeuThrPheGlnSerPheSerTyrAlaThrIleAsnThrAlaPheThr
 20 PheProMetSerGlnSerSerPheThrValGlyAlaAspThrPheSer
 SerGlyAsnGluValTyrIleAspArgPheGluLeuIleProValThr
 AlaThrPheGluAlaGluTyrAspLeuGluArgAlaGlnLysAlaVal
 AsnAlaLeuPheThrSerIleAsnGlnIleGlyIleLysThrAspVal
 ThrAspTyrHisIleAspGlnValSerAsnLeuValAspCysLeuSer
 25 AspGluPheCysLeuAspGluLysArgGluLeuSerGluLysValLys
 HisAlaLysArgLeuSerAspGluArgAsnLeuLeuGlnAspProAsn
 PheLysGlyIleAsnArgGlnLeuAspArgGlyTrpArgGlySerThr
 AspIleThrIleGlnArgGlyAspAspValPheLysGluAsnTyrVal
 ThrLeuProGlyThrPheAspGluCysTyrProThrTyrLeuTyrGln
 30 LysIleAspGluSerLysLeuLysAlaPheThrArgTyrGlnLeuArg
 GlyTyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
 AsnAlaLysHisGluThrValAsnValProGlyThrGlySerLeuTrp
 ProLeuSerAlaGlnSerProIleGlyLysCysGlyGluProAsnArg

CysAlaProHisLeuGluTrpAsnProAspLeuAspCysSerCysArg
 AspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspIle
 AspValGlyCysThrAspLeuAsnGluAspLeuGlyValTrpValIle
 PheLysIleLysThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGlu
 PheLeuGluGluLysProLeuValGlyGluAlaLeuAlaArgValLys
 5 ArgAlaGluLysLysTrpArgAspLysArgGluLysLeuGluTrpGlu
 ThrAsnIleValTyrLysGluAlaLysGluSerValAspAlaLeuPhe
 ValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsnIleAlaMet
 IleHisAlaAlaAspLysArgValHisSerIleArgGluAlaTyrLeu
 10 ProGluLeuSerValIleProGlyValAsnAlaAlaIlePheGluGlu
 LeuGluGlyArgIlePheThrAlaPheSerLeuTyrAspAlaArgAsn
 ValIleLysAsnGlyAspPheAsnAsnGlyLeuSerCysTrpAsnVal
 LysGlyHisValAspValGluGluGlnAsnAsnGlnArgSerValLeu
 ValValProGluTrpGluAlaGluValSerGlnGluValArgValCys
 15 ProGlyArgGlyTyrIleLeuArgValThrAlaTyrLysGluGlyTyr
 GlyGluGlyCysValThrIleHisGluIleGluAsnAsnThrAspGlu
 LeuLysPheSerAsnCysValGluGluGluIleTyrProAsnAsnThr
 ValThrCysAsnAspTyrThrValAsnGlnGluGluTyrGlyGlyAla
 TyrThrSerArgAsnArgGlyTyrAsnGluAlaProSerValProAla
 20 AspTyrAlaSerValTyrGluGluLysSerTyrThrAspGlyArgArg
 GluAsnProCysGluPheAsnArgGlyTyrArgAspTyrThrProLeu
 ProValGlyTyrValThrLysGluLeuGluTyrPheProGluThrAsp
 LysValTrpIleGluIleGlyGluThrGluGlyThrPheIleValAsp
 SerValGluLeuLeuLeuMetGluGlu

25 6.5.2 AMINO ACID SEQUENCE OF THE EG11074 CRYSTAL PROTEIN (SEQ ID NO:12)

MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeu
 SerAsnProGluValGluValLeuGlyGlyGluArgIleGluThrGly
 TyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
 30 GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIle
 TrpGlyIlePheGlyProSerGlnTrpAspAlaPheLeuValGlnIle
 GluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
 IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGlu

SerPheArgGluTrpGluAlaAspProThrAsnProAlaLeuArgGlu
 GluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla
 IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerVal
 TyrValGlnAlaAlaAsnLeuHisLeuSerValLeuArgAspValSer
 5 ValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
 TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspTyrAlaVal
 ArgTrpTyrAsnThrGlyLeuGluArgValTrpGlyProAspSerArg
 AspTrpValArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
 LeuAspIleValAlaLeuPheProAsnTyrAspSerArgArgTyrPro
 10 IleArgThrValSerGlnLeuThrArgGluIleTyrThrAsnProVal
 LeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
 ArgSerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThr
 IleTyrThrAspAlaHisArgGlyTyrTyrTyrTrpSerGlyHisGln
 IleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro
 15 LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAla
 GlnLeuGlyGlnGlyValTyrArgThrLeuSerSerThrLeuTyrArg
 ArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp
 GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaVal
 TyrArgLysSerGlyThrValAspSerLeuAspGluIleProProGln
 20 AsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
 ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIle
 ArgAlaProMetPheSerTrpThrHisArgSerAlaThrProThrAsn
 ThrIleAspProGluArgIleThrGlnIleProLeuValLysAlaHis
 ThrLeuGlnSerGlyThrThrValValArgGlyProGlyPheThrGly
 25 GlyAspIleLeuArgArgThrSerGlyGlyProPheAlaTyrThrIle
 ValAsnIleAsnGlyGlnLeuProGlnArgTyrArgAlaArgIleArg
 TyrAlaSerThrThrAsnLeuArgIleTyrValThrValAlaGlyGlu
 ArgIlePheAlaGlyGlnPheAsnLysThrMetAspThrGlyAspPro
 LeuThrPheGlnSerPheSerTyrAlaThrIleAsnThrAlaPheThr
 30 PheProMetSerGlnSerSerPheThrValGlyAlaAspThrPheSer
 SerGlyAsnGluValTyrIleAspArgPheGluLeuIleProValThr
 AlaThrLeuGluAlaGluTyrAsnLeuGluArgAlaGlnLysAlaVal
 AsnAlaLeuPheThrSerThrAsnGlnLeuGlyLeuLysThrAsnVal

ThrAspTyrHisIleAspGlnValSerAsnLeuValThrTyrLeuSer
 AspGluPheCysLeuAspGluLysArgGluLeuSerGluLysValLys
 HisAlaLysArgLeuSerAspGluArgAsnLeuLeuGlnAspSerAsn
 PheLysAspIleAsnArgGlnProGluArgGlyTrpGlyGlySerThr
 5 GlyIleThrIleGlnGlyGlyAspAspValPheLysGluAsnTyrVal
 ThrLeuSerGlyThrPheAspGluCysTyrProThrTyrLeuTyrGln
 LysIleAspGluSerLysLeuLysAlaPheThrArgTyrGlnLeuArg
 GlyTyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
 AsnAlaLysHisGluThrValAsnValProGlyThrGlySerLeuTrp
 10 ProLeuSerAlaGlnSerProIleGlyLysCysGlyGluProAsnArg
 CysAlaProHisLeuGluTrpAsnProAspLeuAspCysSerCysArg
 AspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspIle
 AspValGlyCysThrAspLeuAsnGluAspLeuGlyValTrpValIle
 PheLysIleLysThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGlu
 15 PheLeuGluGluLysProLeuValGlyGluAlaLeuAlaArgValLys
 ArgAlaGluLysLysTrpArgAspLysArgGluLysLeuGluTrpGlu
 ThrAsnIleValTyrLysGluAlaLysGluSerValAspAlaLeuPhe
 ValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsnIleAlaMet
 IleHisAlaAlaAspLysArgValHisSerIleArgGluAlaTyrLeu
 20 ProGluLeuSerValIleProGlyValAsnAlaAlaIlePheGluGlu
 LeuGluGlyArgIlePheThrAlaPheSerLeuTyrAspAlaArgAsn
 ValIleLysAsnGlyAspPheAsnAsnGlyLeuSerCysTrpAsnVal
 LysGlyHisValAspValGluGluGlnAsnAsnGlnArgSerValLeu
 ValValProGluTrpGluAlaGluValSerGlnGluValArgValCys
 25 ProGlyArgGlyTyrIleLeuArgValThrAlaTyrLysGluGlyTyr
 GlyGluGlyCysValThrIleHisGluIleGluAsnAsnThrAspGlu
 LeuLysPheSerAsnCysValGluGluGluIleTyrProAsnAsnThr
 ValThrCysAsnAspTyrThrValAsnGlnGluGluTyrGlyGlyAla
 TyrThrSerArgAsnArgGlyTyrAsnGluAlaProSerValProAla
 30 AspTyrAlaSerValTyrGluGluLysSerTyrThrAspGlyArgArg
 GluAsnProCysGluPheAsnArgGlyTyrArgAspTyrThrProLeu
 ProValGlyTyrValThrLysGluLeuGluTyrPheProGluThrAsp
 LysValTrpIleGluIleGlyGluThrGluGlyThrPheIleValAsp

SerValGluLeuLeuLeuMetGluGlu

6.5.3 AMINO ACID SEQUENCE OF THE EG11735 CRYSTAL PROTEIN (SEQ ID NO:14)

5 MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeu
SerAsnProGluValGluValLeuGlyGlyGluArgIleGluThrGly
TyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIle
TrpGlyIlePheGlyProSerGlnTrpAspAlaPheLeuValGlnIle
10 GluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGlu
SerPheArgGluTrpGluAlaAspProThrAsnProAlaLeuArgGlu
GluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla
IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerVal
15 TyrValGlnAlaAlaAsnLeuHisLeuSerValLeuArgAspValSer
ValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspHisAlaVal
ArgTrpTyrAsnThrGlyLeuGluArgValTrpGlyProAspSerArg
AspTrpIleArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
20 LeuAspIleValSerLeuPheProAsnTyrAspSerArgThrTyrPro
IleArgThrValSerGlnLeuThrArgGluIleTyrThrAsnProVal
LeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
GlySerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThr
IleTyrThrAspAlaHisArgGlyGluTyrTyrTrpSerGlyHisGln
25 IleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro
LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAla
GlnLeuGlyGlnGlyValTyrArgThrLeuSerSerThrLeuTyrArg
ArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp
GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaVal
TyrArgLysSerGlyThrValAspSerLeuAspGluIleProProGln
30 AsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIle
ArgAlaProMetPheSerTrpThrHisArgSerAlaThrProThrAsn

ThrIleAspProGluArgIleThrGlnIleProLeuValLysAlaHis
 ThrLeuGlnSerGlyThrThrValValArgGlyProGlyPheThrGly
 GlyAspIleLeuArgArgThrSerGlyGlyProPheAlaTyrThrIle
 ValAsnIleAsnGlyGlnLeuProGlnArgTyrArgAlaArgIleArg
 5 TyrAlaSerThrThrAsnLeuArgIleTyrValThrValAlaGlyGlu
 ArgIlePheAlaGlyGlnPheAsnLysThrMetAspThrGlyAspPro
 LeuThrPheGlnSerPheSerTyrAlaThrIleAsnThrAlaPheThr
 PheProMetSerGlnSerSerPheThrValGlyAlaAspThrPheSer
 SerGlyAsnGluValTyrIleAspArgPheGluLeuIleProValThr
 10 AlaThrPheGluAlaGluTyrAspLeuGluArgAlaGlnLysAlaVal
 AsnAlaLeuPheThrSerIleAsnGlnIleGlyIleLysThrAspVal
 ThrAspTyrHisIleAspGlnValSerAsnLeuValAspCysLeuSer
 AspGluPheCysLeuAspGluLysArgGluLeuSerGluLysValLys
 HisAlaLysArgLeuSerAspGluArgAsnLeuLeuGlnAspProAsn
 15 PheLysGlyIleAsnArgGlnLeuAspArgGlyTrpArgGlySerThr
 AspIleThrIleGlnArgGlyAspAspValPheLysGluAsnTyrVal
 ThrLeuProGlyThrPheAspGluCysTyrProThrTyrLeuTyrGln
 LysIleAspGluSerLysLeuLysAlaPheThrArgTyrGlnLeuArg
 GlyTyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
 20 AsnAlaLysHisGluThrValAsnValProGlyThrGlySerLeuTrp
 ProLeuSerAlaGlnSerProIleGlyLysCysGlyGluProAsnArg
 CysAlaProHisLeuGluTrpAsnProAspLeuAspCysSerCysArg
 AspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspIle
 AspValGlyCysThrAspLeuAsnGluAspLeuGlyValTrpValIle
 25 PheLysIleLysThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGlu
 PheLeuGluGluLysProLeuValGlyGluAlaLeuAlaArgValLys
 ArgAlaGluLysLysTrpArgAspLysArgGluLysLeuGluTrpGlu
 ThrAsnIleValTyrLysGluAlaLysGluSerValAspAlaLeuPhe
 ValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsnIleAlaMet
 30 IleHisAlaAlaAspLysArgValHisSerIleArgGluAlaTyrLeu
 ProGluLeuSerValIleProGlyValAsnAlaAlaIlePheGluGlu
 LeuGluGlyArgIlePheThrAlaPheSerLeuTyrAspAlaArgAsn
 ValIleLysAsnGlyAspPheAsnAsnGlyLeuSerCysTrpAsnVal

LysGlyHisValAspValGluGluGlnAsnAsnGlnArgSerValLeu
 ValValProGluTrpGluAlaGluValSerGlnGluValArgValCys
 ProGlyArgGlyTyrIleLeuArgValThrAlaTyrLysGluGlyTyr
 GlyGluGlyCysValThrIleHisGluIleGluAsnAsnThrAspGlu
 5 LeuLysPheSerAsnCysValGluGluGluIleTyrProAsnAsnThr
 ValThrCysAsnAspTyrThrValAsnGlnGluGluTyrGlyGlyAla
 TyrThrSerArgAsnArgGlyTyrAsnGluAlaProSerValProAla
 AspTyrAlaSerValTyrGluGluLysSerTyrThrAspGlyArgArg
 GluAsnProCysGluPheAsnArgGlyTyrArgAspTyrThrProLeu
 10 ProValGlyTyrValThrLysGluLeuGluTyrPheProGluThrAsp
 LysValTrpIleGluIleGlyGluThrGluGlyThrPheIleValAsp
 SerValGluLeuLeuLeuMetGluGlu

6.5.4 AMINO ACID SEQUENCE OF THE EG11092 CRYSTAL PROTEIN (SEQ ID NO:26)

15 MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeu
 SerAsnProGluValGluValLeuGlyGlyGluArgIleGluThrGly
 TyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
 GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIle
 TrpGlyIlePheGlyProSerGlnTrpAspAlaPheLeuValGlnIle
 20 GluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
 IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGlu
 SerPheArgGluTrpGluAlaAspProThrAsnProAlaLeuArgGlu
 GluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla
 IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerVal
 25 TyrValGlnAlaAlaAsnLeuHisLeuSerValLeuArgAspValSer
 ValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
 TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspHisAlaVal
 ArgTrpTyrAsnThrGlyLeuGluArgValTrpGlyProAspSerArg
 AspTrpIleArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
 30 LeuAspIleValSerLeuPheProAsnTyrAspSerArgThrTyrPro
 IleArgThrValSerGlnLeuThrArgGluIleTyrThrAsnProVal
 LeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
 ArgSerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThr

IleTyrThrAspAlaHisArgGlyTyrTyrTyrTrpSerGlyHisGln
 IleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro
 LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAla
 GlnLeuGlyGlnGlyValTyrArgThrLeuSerSerThrLeuTyrArg
 5 ArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp
 GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaVal
 TyrArgLysSerGlyThrValAspSerLeuAspGluIleProProGln
 AsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
 ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIle
 10 ArgAlaProMetPheSerTrpThrHisArgSerAlaThrProThrAsn
 ThrIleAspProGluArgIleThrGlnIleProLeuValLysAlaHis
 ThrLeuGlnSerGlyThrThrValValArgGlyProGlyPheThrGly
 GlyAspIleLeuArgArgThrSerGlyGlyProPheAlaTyrThrIle
 ValAsnIleAsnGlyGlnLeuProGlnArgTyrArgAlaArgIleArg
 15 TyrAlaSerThrThrAsnLeuArgIleTyrValThrValAlaGlyGlu
 ArgIlePheAlaGlyGlnPheAsnLysThrMetAspThrGlyAspPro
 LeuThrPheGlnSerPheSerTyrAlaThrIleAsnThrAlaPheThr
 PheProMetSerGlnSerSerPheThrValGlyAlaAspThrPheSer
 SerGlyAsnGluValTyrIleAspArgPheGluLeuIleProValThr
 20 AlaThrPheGluAlaGluTyrAspLeuGluArgAlaGlnLysAlaVal
 AsnAlaLeuPheThrSerIleAsnGlnIleGlyIleLysThrAspVal
 ThrAspTyrHisIleAspGlnValSerAsnLeuValAspCysLeuSer
 AspGluPheCysLeuAspGluLysArgGluLeuSerGluLysValLys
 HisAlaLysArgLeuSerAspGluArgAsnLeuLeuGlnAspProAsn
 25 PheLysGlyIleAsnArgGlnLeuAspArgGlyTrpArgGlySerThr
 AspIleThrIleGlnArgGlyAspAspValPheLysGluAsnTyrVal
 ThrLeuProGlyThrPheAspGluCysTyrProThrTyrLeuTyrGln
 LysIleAspGluSerLysLeuLysAlaPheThrArgTyrGlnLeuArg
 GlyTyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
 AsnAlaLysHisGluThrValAsnValProGlyThrGlySerLeuTrp
 30 ProLeuSerAlaGlnSerProIleGlyLysCysGlyGluProAsnArg
 CysAlaProHisLeuGluTrpAsnProAspLeuAspCysSerCysArg
 AspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspIle

AspValGlyCysThrAspLeuAsnGluAspLeuGlyValTrpValIle
 PheLysIleLysThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGlu
 PheLeuGluGluLysProLeuValGlyGluAlaLeuAlaArgValLys
 ArgAlaGluLysLysTrpArgAspLysArgGluLysLeuGluTrpGlu
 5 ThrAsnIleValTyrLysGluAlaLysGluSerValAspAlaLeuPhe
 ValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsnIleAlaMet
 IleHisAlaAlaAspLysArgValHisSerIleArgGluAlaTyrLeu
 ProGluLeuSerValIleProGlyValAsnAlaAlaIlePheGluGlu
 LeuGluGlyArgIlePheThrAlaPheSerLeuTyrAspAlaArgAsn
 10 ValIleLysAsnGlyAspPheAsnAsnGlyLeuSerCysTrpAsnVal
 LysGlyHisValAspValGluGluGlnAsnAsnGlnArgSerValLeu
 ValValProGluTrpGluAlaGluValSerGlnGluValArgValCys
 ProGlyArgGlyTyrIleLeuArgValThrAlaTyrLysGluGlyTyr
 GlyGluGlyCysValThrIleHisGluIleGluAsnAsnThrAspGlu
 15 LeuLysPheSerAsnCysValGluGluGluIleTyrProAsnAsnThr
 ValThrCysAsnAspTyrThrValAsnGlnGluGluTyrGlyGlyAla
 TyrThrSerArgAsnArgGlyTyrAsnGluAlaProSerValProAla
 AspTyrAlaSerValTyrGluGluLysSerTyrThrAspGlyArgArg
 GluAsnProCysGluPheAsnArgGlyTyrArgAspTyrThrProLeu
 20 ProValGlyTyrValThrLysGluLeuGluTyrPheProGluThrAsp
 LysValTrpIleGluIleGlyGluThrGluGlyThrPheIleValAsp
 SerValGluLeuLeuLeuMetGluGlu

6.5.5 AMINO ACID SEQUENCE OF THE EG11751 CRYSTAL PROTEIN (SEQ ID NO:28)

25 MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeu
 SerAsnProGluValGluValLeuGlyGlyGluArgIleGluThrGly
 TyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
 GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIle
 TrpGlyIlePheGlyProSerGlnTrpAspAlaPheLeuValGlnIle
 30 GluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
 IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGlu
 SerPheArgGluTrpGluAlaAspProThrAsnProAlaLeuArgGlu
 GluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla

IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerVal
 TyrValGlnAlaAlaAsnLeuHisLeuSerValLeuArgAspValSer
 ValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
 TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspTyrAlaVal
 5 ArgTrpTyrAsnThrGlyLeuGluArgValTrpGlyProAspSerArg
 AspTrpValArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
 LeuAspIleValAlaLeuPheProAsnTyrAspSerArgArgTyrPro
 IleArgThrValSerGlnLeuThrArgGluIleTyrThrAsnProVal
 LeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
 10 ArgSerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThr
 IleTyrThrAspAlaHisArgGlyTyrTyrTyrTrpSerGlyHisGln
 IleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro
 LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAla
 GlnLeuGlyGlnGlyValTyrArgThrLeuSerSerThrLeuTyrArg
 15 ArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp
 GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaVal
 TyrArgLysSerGlyThrValAspSerLeuAspGluIleProProGln
 AsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
 ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIle
 20 ArgAlaProMetPheSerTrpIleHisArgSerAlaGluPheAsnAsn
 IleIleAlaSerAspSerIleThrGlnIleProLeuValLysAlaHis
 ThrLeuGlnSerGlyThrThrValValArgGlyProGlyPheThrGly
 GlyAspIleLeuArgArgThrSerGlyGlyProPheAlaTyrThrIle
 ValAsnIleAsnGlyGlnLeuProGlnArgTyrArgAlaArgIleArg
 25 TyrAlaSerThrThrAsnLeuArgIleTyrValThrValAlaGlyGlu
 ArgIlePheAlaGlyGlnPheAsnLysThrMetAspThrGlyAspPro
 LeuThrPheGlnSerPheSerTyrAlaThrIleAsnThrAlaPheThr
 PheProMetSerGlnSerSerPheThrValGlyAlaAspThrPheSer
 SerGlyAsnGluValTyrIleAspArgPheGluLeuIleProValThr
 30 AlaThrPheGluAlaGluTyrAspLeuGluArgAlaGlnLysAlaVal
 AsnAlaLeuPheThrSerIleAsnGlnIleGlyIleLysThrAspVal
 ThrAspTyrHisIleAspGlnValSerAsnLeuValAspCysLeuSer
 AspGluPheCysLeuAspGluLysArgGluLeuSerGluLysValLys

HisAlaLysArgLeuSerAspGluArgAsnLeuLeuGlnAspProAsn
 PheLysGlyIleAsnArgGlnLeuAspArgGlyTrpArgGlySerThr
 AspIleThrIleGlnArgGlyAspAspValPheLysGluAsnTyrVal
 ThrLeuProGlyThrPheAspGluCysTyrProThrTyrLeuTyrGln
 5 LysIleAspGluSerLysLeuLysAlaPheThrArgTyrGlnLeuArg
 GlyTyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
 AsnAlaLysHisGluThrValAsnValProGlyThrGlySerLeuTrp
 ProLeuSerAlaGlnSerProIleGlyLysCysGlyGluProAsnArg
 CysAlaProHisLeuGluTrpAsnProAspLeuAspCysSerCysArg
 10 AspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspIle
 AspValGlyCysThrAspLeuAsnGluAspLeuGlyValTrpValIle
 PheLysIleLysThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGlu
 PheLeuGluGluLysProLeuValGlyGluAlaLeuAlaArgValLys
 ArgAlaGluLysLysTrpArgAspLysArgGluLysLeuGluTrpGlu
 15 ThrAsnIleValTyrLysGluAlaLysGluSerValAspAlaLeuPhe
 ValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsnIleAlaMet
 IleHisAlaAlaAspLysArgValHisSerIleArgGluAlaTyrLeu
 ProGluLeuSerValIleProGlyValAsnAlaAlaIlePheGluGlu
 LeuGluGlyArgIlePheThrAlaPheSerLeuTyrAspAlaArgAsn
 20 ValIleLysAsnGlyAspPheAsnAsnGlyLeuSerCysTrpAsnVal
 LysGlyHisValAspValGluGluGlnAsnAsnGlnArgSerValLeu
 ValValProGluTrpGluAlaGluValSerGlnGluValArgValCys
 ProGlyArgGlyTyrIleLeuArgValThrAlaTyrLysGluGlyTyr
 GlyGluGlyCysValThrIleHisGluIleGluAsnAsnThrAspGlu
 25 LeuLysPheSerAsnCysValGluGluGluIleTyrProAsnAsnThr
 ValThrCysAsnAspTyrThrValAsnGlnGluGluTyrGlyGlyAla
 TyrThrSerArgAsnArgGlyTyrAsnGluAlaProSerValProAla
 AspTyrAlaSerValTyrGluGluLysSerTyrThrAspGlyArgArg
 GluAsnProCysGluPheAsnArgGlyTyrArgAspTyrThrProLeu
 30 ProValGlyTyrValThrLysGluLeuGluTyrPheProGluThrAsp
 LysValTrpIleGluIleGlyGluThrGluGlyThrPheIleValAsp
 SerValGluLeuLeuLeuMetGluGlu

6.5.6 AMINO ACID SEQUENCE OF THE EG11090 CRYSTAL PROTEIN (SEQ ID NO:30)

MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCys
 LeuSerAsnProGluValGluValLeuGlyGlyGluArgIleGlu
 5 ThrGlyTyrThrProIleAspIleSerLeuSerLeuThrGlnPhe
 LeuLeuSerGluPheValProGlyAlaGlyPheValLeuGlyLeu
 ValAspIleIleTrpGlyIlePheGlyProSerGlnTrpAspAla
 PheLeuValGlnIleGluGlnLeuIleAsnGlnArgIleGluGlu
 PheAlaArgAsnGlnAlaIleSerArgLeuGluGlyLeuSerAsn
 10 LeuTyrGlnIleTyrAlaGluSerPheArgGluTrpGluAlaAsp
 ProThrAsnProAlaLeuArgGluGluMetArgIleGlnPheAsn
 AspMetAsnSerAlaLeuThrThrAlaIleProLeuPheAlaVal
 GlnAsnTyrGlnValProLeuLeuSerValTyrValGlnAlaAla
 AsnLeuHisLeuSerValLeuArgAspValSerValPheGlyGln
 15 ArgTrpGlyPheAspAlaAlaThrIleAsnSerArgTyrAsnAsp
 LeuThrArgLeuIleGlyAsnTyrThrAspTyrAlaValArgTrp
 TyrAsnThrGlyLeuGluArgValTrpGlyProAspSerArgAsp
 TrpValArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
 LeuAspIleValAlaLeuPheProAsnTyrAspSerArgArgTyr
 20 ProIleArgThrValSerGlnLeuThrArgGluIleTyrThrAsn
 ProValLeuGluAsnPheAspGlySerPheArgGlySerAlaGln
 GlyIleGluArgSerIleArgSerProHisLeuMetAspIleLeu
 AsnSerIleThrIleTyrThrAspAlaHisArgGlyTyrTyrTyr
 TrpSerGlyHisGlnIleMetAlaSerProValGlyPheSerGly
 25 ProGluPheThrPheProLeuTyrGlyThrMetGlyAsnAlaAla
 ProGlnGlnArgIleValAlaGlnLeuGlyGlnGlyValTyrArg
 ThrLeuSerSerThrLeuTyrArgArgProPheAsnIleGlyIle
 AsnAsnGlnGlnLeuSerValLeuAspGlyThrGluPheAlaTyr
 GlyThrSerSerAsnLeuProSerAlaValTyrArgLysSerGly
 30 ThrValAspSerLeuAspGluIleProProGlnAsnAsnAsnVal
 ProProArgGlnGlyPheSerHisArgLeuSerHisValSerMet
 PheArgSerGlyPheSerAsnSerSerValSerIleIleArgAla
 ProMetPheSerTrpIleHisArgSerAlaThrLeuThrAsnThr

IleAspProGluArgIleAsnGlnIleProLeuValLysGlyPhe
 ArgValTrpGlyGlyThrSerValIleThrGlyProGlyPheThr
 GlyGlyAspIleLeuArgArgAsnThrPheGlyAspPheValSer
 LeuGlnValAsnIleAsnSerProIleThrGlnArgTyrArgLeu
 5 ArgPheArgTyrAlaSerSerArgAspAlaArgValIleValLeu
 ThrGlyAlaAlaSerThrGlyValGlyGlyGlnValSerValAsn
 MetProLeuGlnLysThrMetGluIleGlyGluAsnLeuThrSer
 ArgThrPheArgTyrThrAspPheSerAsnProPheSerPheArg
 AlaAsnProAspIleIleGlyIleSerGluGlnProLeuPheGly
 10 AlaGlySerIleSerSerGlyGluLeuTyrIleAspLysIleGlu
 IleIleLeuAlaAspAlaThrPheGluAlaGluSerAspLeuGlu
 ArgAlaGlnLysAlaValAsnAlaLeuPheThrSerSerAsnGln
 IleGlyLeuLysThrAspValThrAspTyrHisIleAspGlnVal
 SerAsnLeuValAspCysLeuSerAspGluPheCysLeuAspGlu
 15 LysArgGluLeuSerGluLysValLysHisAlaLysArgLeuSer
 AspGluArgAsnLeuLeuGlnAspProAsnPheArgGlyIleAsn
 ArgGlnProAspArgGlyTrpArgGlySerThrAspIleThrIle
 GlnGlyGlyAspAspValPheLysGluAsnTyrValThrLeuPro
 GlyThrValAspGluCysTyrProThrTyrLeuTyrGlnLysIle
 20 AspGluSerLysLeuLysAlaTyrThrArgTyrGluLeuArgGly
 TyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
 AsnAlaLysHisGluIleValAsnValProGlyThrGlySerLeu
 TrpProLeuSerAlaGlnSerProIleGlyLysCysGlyGluPro
 AsnArgCysAlaProHisLeuGluTrpAsnProAspLeuAspCys
 25 SerCysArgAspGlyGluLysCysAlaHisHisSerHisHisPhe
 ThrLeuAspIleAspValGlyCysThrAspLeuAsnGluAspLeu
 GlyValTrpValIlePheLysIleLysThrGlnAspGlyHisAla
 ArgLeuGlyAsnLeuGluPheLeuGluGluLysProLeuLeuGly
 GluAlaLeuAlaArgValLysArgAlaGluLysLysTrpArgAsp
 30 LysArgGluLysLeuGlnLeuGluThrAsnIleValTyrLysGlu
 AlaLysGluSerValAspAlaLeuPheValAsnSerGlnTyrAsp
 ArgLeuGlnValAspThrAsnIleAlaMetIleHisAlaAlaAsp
 LysArgValHisArgIleArgGluAlaTyrLeuProGluLeuSer

ValIleProGlyValAsnAlaAlaIlePheGluGluLeuGluGly
 ArgIlePheThrAlaTyrSerLeuTyrAspAlaArgAsnValIle
 LysAsnGlyAspPheAsnAsnGlyLeuLeuCysTrpAsnValLys
 GlyHisValAspValGluGluGlnAsnAsnHisArgSerValLeu
 5 ValIleProGluTrpGluAlaGluValSerGlnGluValArgVal
 CysProGlyArgGlyTyrIleLeuArgValThrAlaTyrLysGlu
 GlyTyrGlyGluGlyCysValThrIleHisGluIleGluAspAsn
 ThrAspGluLeuLysPheSerAsnCysValGluGluGluValTyr
 ProAsnAsnThrValThrCysAsnAsnTyrThrGlyThrGlnGlu
 10 GluTyrGluGlyThrTyrThrSerArgAsnGlnGlyTyrAspGlu
 AlaTyrGlyAsnAsnProSerValProAlaAspTyrAlaSerVal
 TyrGluGluLysSerTyrThrAspGlyArgArgGluAsnProCys
 GluSerAsnArgGlyTyrGlyAspTyrThrProLeuProAlaGly
 TyrValThrLysAspLeuGluTyrPheProGluThrAspLysVal
 15 TrpIleGluIleGlyGluThrGluGlyThrPheIleValAspSer
 ValGluLeuLeuLeuMetGluGlu

6.6 EXAMPLE 6 -- DNA SEQUENCES ENCODING THE NOVEL CRYSTAL PROTEINS

6.6.1 DNA SEQUENCE ENCODING THE EG11063 CRYSTAL PROTEIN (SEQ ID NO:9)

20	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
25	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
30	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
35	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816

	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
5	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
10	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT ACA AAT	1392
	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
15	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
20	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG	1872
	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG	1920
	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA	1968
25	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC	2064
	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG	2112
	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC	2160
	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
30	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
35	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
40	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736

	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
5	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
10	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312
	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3360
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA	3408
15	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504
	AGC GTG GAA TTA CTC CTT ATG GAG GAA	3531

6.6.2 DNA SEQUENCE ENCODING THE EG11074 CRYSTAL PROTEIN (SEQ ID NO:11)

20	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
25	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
30	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
35	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
40	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008

	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
5	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT ACA AAT	1392
	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
10	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
15	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	GCA ACA CTC GAG GCT GAA TAT AAT CTG GAA AGA GCG CAG AAG GCG GTG	1872
	AAT GCG CTG TTT ACG TCT ACA AAC CAA CTA GGG CTA AAA ACA AAT GTA	1920
20	ACG GAT TAT CAT ATT GAT CAA GTG TCC AAT TTA GTT ACG TAT TTA TCG	1968
	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
	CAT GCG AAG CGA CTC AGT GAT GAA CGC AAT TTA CTC CAA GAT TCA AAT	2064
	TTC AAA GAC ATT AAT AGG CAA CCA GAA CGT GGG TGG GGC GGA AGT ACA	2112
	GGG ATT ACC ATC CAA GGA GGG GAT GAC GTA TTT AAA GAA AAT TAC GTC	2160
25	ACA CTA TCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
30	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
35	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
40	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928

	GTC	ATT	AAA	AAT	GGT	GAT	TTT	AAT	AAT	GGC	TTA	TCC	TGC	TGG	AAC	GTG	2976
	AAA	GGG	CAT	GTA	GAT	GTA	GAA	GAA	CAA	AAC	AAC	CAA	CGT	TCG	GTC	CTT	3024
	GTT	GTT	CCG	GAA	TGG	GAA	GCA	GAA	GTG	TCA	CAA	GAA	GTT	CGT	GTC	TGT	3072
	CCG	GGT	CGT	GGC	TAT	ATC	CTT	CGT	GTC	ACA	GCG	TAC	AAG	GAG	GGA	TAT	3120
5	GGA	GAA	GGT	TGC	GTA	ACC	ATT	CAT	GAG	ATC	GAG	AAC	AAT	ACA	GAC	GAA	3168
	CTG	AAG	TTT	AGC	AAC	TGC	GTA	GAA	GAG	GAA	ATC	TAT	CCA	AAT	AAC	ACG	3216
	GTA	ACG	TGT	AAT	GAT	TAT	ACT	GTA	AAT	CAA	GAA	GAA	TAC	GGA	GGT	GCG	3264
	TAC	ACT	TCT	CGT	AAT	CGA	GGA	TAT	AAC	GAA	GCT	CCT	TCC	GTA	CCA	GCT	3312
	GAT	TAT	GCG	TCA	GTC	TAT	GAA	GAA	AAA	TCG	TAT	ACA	GAT	GGA	CGA	AGA	3360
10	GAG	AAT	CCT	TGT	GAA	TTT	AAC	AGA	GGG	TAT	AGG	GAT	TAC	ACG	CCA	CTA	3408
	CCA	GTT	GGT	TAT	GTG	ACA	AAA	GAA	TTA	GAA	TAC	TTC	CCA	GAA	ACC	GAT	3456
	AAG	GTA	TGG	ATT	GAG	ATT	GGA	GAA	ACG	GAA	GGA	ACA	TTT	ATC	GTG	GAC	3504
	AGC	GTG	GAA	TTA	CTC	CTT	ATG	GAG	GAA								3531

15 6.6.3 DNA SEQUENCE ENCODING THE EG11735 CRYSTAL PROTEIN (SEQ ID NO:13)

	ATG	GAT	AAC	AAT	CCG	AAC	ATC	AAT	GAA	TGC	ATT	CCT	TAT	AAT	TGT	TTA	48
	AGT	AAC	CCT	GAA	GTA	GAA	GTA	TTA	GGT	GGA	GAA	AGA	ATA	GAA	ACT	GGT	96
	TAC	ACC	CCA	ATC	GAT	ATT	TCC	TTG	TCG	CTA	ACG	CAA	TTT	CTT	TTG	AGT	144
	GAA	TTT	GTT	CCC	GGT	GCT	GGA	TTT	GTG	TTA	GGA	CTA	GTT	GAT	ATA	ATA	192
20	TGG	GGA	ATT	TTT	GGT	CCC	TCT	CAA	TGG	GAC	GCA	TTT	CTT	GTA	CAA	ATT	240
	GAA	CAG	TTA	ATT	AAC	CAA	AGA	ATA	GAA	GAA	TTC	GCT	AGG	AAC	CAA	GCC	288
	ATT	TCT	AGA	TTA	GAA	GGA	CTA	AGC	AAT	CTT	TAT	CAA	ATT	TAC	GCA	GAA	336
	TCT	TTT	AGA	GAG	TGG	GAA	GCA	GAT	CCT	ACT	AAT	CCA	GCA	TTA	AGA	GAA	384
	GAG	ATG	CGT	ATT	CAA	TTC	AAT	GAC	ATG	AAC	AGT	GCC	CTT	ACA	ACC	GCT	432
25	ATT	CCT	CTT	TTT	GCA	GTT	CAA	AAT	TAT	CAA	GTT	CCT	CTT	TTA	TCA	GTA	480
	TAT	GTT	CAA	GCT	GCA	AAT	TTA	CAT	TTA	TCA	GTT	TTG	AGA	GAT	GTT	TCA	528
	GTG	TTT	GGA	CAA	AGG	TGG	GGA	TTT	GAT	GCC	GCG	ACT	ATC	AAT	AGT	CGT	576
	TAT	AAT	GAT	TTA	ACT	AGG	CTT	ATT	GGC	AAC	TAT	ACA	GAT	CAT	GCT	GTA	624
	CGC	TGG	TAC	AAT	ACG	GGA	TTA	GAG	CGT	GTA	TGG	GGA	CCG	GAT	TCT	AGA	672
30	GAT	TGG	ATA	AGA	TAT	AAT	CAA	TTT	AGA	AGA	GAA	TTA	ACA	CTA	ACT	GTA	720
	TTA	GAT	ATC	GTT	TCT	CTA	TTT	CCG	AAC	TAT	GAT	AGT	AGA	ACG	TAT	CCA	768
	ATT	CGA	ACA	GTT	TCC	CAA	TTA	ACA	AGA	GAA	ATT	TAT	ACA	AAC	CCA	GTA	816
	TTA	GAA	AAT	TTT	GAT	GGT	AGT	TTT	CGA	GGC	TCG	GCT	CAG	GGC	ATA	GAA	864
	GGA	AGT	ATT	AGG	AGT	CCA	CAT	TTG	ATG	GAT	ATA	CTT	AAC	AGT	ATA	ACC	912
35	ATC	TAT	ACG	GAT	GCT	CAT	AGA	GGA	GAA	TAT	TAT	TGG	TCA	GGG	CAT	CAA	960
	ATA	ATG	GCT	TCT	CCT	GTA	GGG	TTT	TCG	GGG	CCA	GAA	TTC	ACT	TTT	CCG	1008
	CTA	TAT	GGA	ACT	ATG	GGA	AAT	GCA	GCT	CCA	CAA	CAA	CGT	ATT	GTT	GCT	1056
	CAA	CTA	GGT	CAG	GGC	GTG	TAT	AGA	ACA	TTA	TCG	TCC	ACT	TTA	TAT	AGA	1104
	AGA	CCT	TTT	AAT	ATA	GGG	ATA	AAT	AAT	CAA	CAA	CTA	TCT	GTT	CTT	GAC	1152
40	GGG	ACA	GAA	TTT	GCT	TAT	GGA	ACC	TCC	TCA	AAT	TTG	CCA	TCC	GCT	GTA	1200

	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT AGA AAT	1392
5	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
10	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG	1872
15	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG	1920
	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA	1968
	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC	2064
	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG	2112
20	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC	2160
	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
25	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
30	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
35	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
40	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120

5 GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA 3153
 CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG 3213
 GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG 3273
 TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT 3333
 GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA 3393
 GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA 3453
 CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT 3513
 AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC 3573
 AGC GTG GAA TTA CTC CTT ATG GAG GAA 3633

10

6.6.4 DNA SEQUENCE ENCODING THE EG11092 CRYSTAL PROTEIN (SEQ ID NO:25)

15 ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA 43
 AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT 49
 TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT 55
 GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA 61
 TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT 67
 GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC 73
 ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA 79
 TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA 85
 20 GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT 91
 ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA 97
 TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA 103
 GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT 109
 TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT CAT GCT GTA 115
 25 CGC TGG TAC AAT ACG GGA TTA GAG CGT GTA TGG GGA CCG GAT TCT AGA 121
 GAT TGG ATA AGA TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA 127
 TTA GAT ATC GTT TCT CTA TTT CCG AAC TAT GAT AGT AGA ACG TAT CCA 133
 ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA 139
 TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA 145
 30 AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC 151
 ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA 157
 ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG 163
 CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT 169
 CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA 175
 35 AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC 181
 GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA 187
 TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG 193
 AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT 199
 GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA 205
 40 AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT ACA AAT 211

	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
5	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
10	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG	1872
	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG	1920
	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA	1968
	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC	2064
15	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG	2112
	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC	2160
	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
20	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
25	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCG ATG	2784
30	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
35	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
40	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312

	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3360
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA	3408
	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504
5	AGC GTG GAA TTA CTC CTT ATG GAG GAA TAG	3534

6.6.5 DNA SEQUENCE ENCODING THE EG11751 CRYSTAL PROTEIN (SEQ ID NO:27)

	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
10	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
15	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
20	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
25	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
30	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
35	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	AGA GCT CCT ATG TTC TCT TGG ATA CAT CGT AGT GCT GAA TTT AAT AAT	1392
	ATA ATT GCA TCG GAT AGT ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
40	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584

	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
5	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG	1872
	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG	1920
	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA	1968
	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
10	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC	2064
	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG	2112
	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC	2160
	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
15	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
20	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
25	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAT GTG	2976
30	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
35	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312
	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3360
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA	3408
	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
40	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504

AGC GTG GAA TTA CTC CTT ATG GAG GAA TAG

3534

6.6.6 DNA SEQUENCE ENCODING THE EG11090 CRYSTAL PROTEIN (SEQ ID NO:29)

	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
5	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
10	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
15	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
20	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
25	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
30	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	AGA GCT CCT ATG TTC TCT TGG ATA CAT CGT AGT GCA ACT CTT ACA AAT	1392
	ACA ATT GAT CCA GAG AGA ATT AAT CAA ATA CCT TTA GTG AAA GGA TTT	1440
	AGA GTT TGG GGG GGC ACC TCT GTC ATT ACA GGA CCA GGA TTT ACA GGA	1488
35	GGG GAT ATC CTT CGA AGA AAT ACC TTT GGT GAT TTT GTA TCT CTA CAA	1536
	GTC AAT ATT AAT TCA CCA ATT ACC CAA AGA TAC CGT TTA AGA TTT CGT	1584
	TAC GCT TCC AGT AGG GAT GCA CGA GTT ATA GTA TTA ACA GGA GCG GCA	1632
	TCC ACA GGA GTG GGA GGC CAA GTT AGT GTA AAT ATG CCT CTT CAG AAA	1680
	ACT ATG GAA ATA GGG GAG AAC TTA ACA TCT AGA ACA TTT AGA TAT ACC	1728
40	GAT TTT AGT AAT CCT TTT TCA TTT AGA GCT AAT CCA GAT ATA ATT GGG	1776

	ATA AGT GAA CAA CCT CTA TTT GGT GCA GGT TCT ATT AGT AGC GGT GAA	1824
	CTT TAT ATA GAT AAA ATT GAA ATT ATT CTA GCA GAT GCA ACA TTT GAA	1872
	GCA GAA TCT GAT TTA GAA AGA GCA CAA AAG GCG GTG AAT GCC CTG TTT	1920
	ACT TCT TCC AAT CAA ATC GGG TTA AAA ACC GAT GTG ACG GAT TAT CAT	1968
5	ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA GAT GAA TTT TGT	2016
	CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA CAT GCG AAG CGA	2064
	CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC TTC AGA GGG ATC	2112
	AAT AGA CAA CCA GAC CGT GGC TGG AGA GGA AGT ACA GAT ATT ACC ATC	2160
	CAA GGA GGA GAT GAC GTA TTC AAA GAG AAT TAC GTC ACA CTA CCG GGT	2208
10	ACC GTT GAT GAG TGC TAT CCA ACG TAT TTA TAT CAG AAA ATA GAT GAG	2256
	TCG AAA TTA AAA GCT TAT ACC CGT TAT GAA TTA AGA GGG TAT ATC GAA	2304
	GAT AGT CAA GAC TTA GAA ATC TAT TTG ATC CGT TAC AAT GCA AAA CAC	2352
	GAA ATA GTA AAT GTG CCA GGC ACG GGT TCC TTA TGG CCG CTT TCA GCC	2400
	CAA AGT CCA ATC GGA AAG TGT GGA GAA CCG AAT CGA TGC GCG CCA CAC	2448
15	CTT GAA TGG AAT CCT GAT CTA GAT TGT TCC TGC AGA GAC GGG GAA AAA	2496
	TGT GCA CAT CAT TCC CAT CAT TTC ACC TTG GAT ATT GAT GTT GGA TGT	2544
	ACA GAC TTA AAT GAG GAC TTA GGT GTA TGG GTG ATA TTC AAG ATT AAG	2592
	ACG CAA GAT GGC CAT GCA AGA CTA GGG AAT CTA GAG TTT CTC GAA GAG	2640
	AAA CCA TTA TTA GGG GAA GCA CTA GCT CGT GTG AAA AGA GCG GAG AAG	2688
20	AAG TGG AGA GAC AAA CGA GAG AAA CTG CAG TTG GAA ACA AAT ATT GTT	2736
	TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT GTA AAC TCT CAA	2784
	TAT GAT AGA TTA CAA GTG GAT ACG AAC ATC GCA ATG ATT CAT GCG GCA	2832
	GAT AAA CGC GTT CAT AGA ATC CGG GAA GCG TAT CTG CCA GAG TTG TCT	2880
	GTG ATT CCA GGT GTC AAT GCG GCC ATT TTC GAA GAA TTA GAG GGA CGT	2928
25	ATT TTT ACA GCG TAT TCC TTA TAT GAT GCG AGA AAT GTC ATT AAA AAT	2976
	GGC GAT TTC AAT AAT GGC TTA TTA TGC TGG AAC GTG AAA GGT CAT GTA	3024
	GAT GTA GAA GAG CAA AAC AAC CAC CGT TCG GTC CTT GTT ATC CCA GAA	3072
	TGG GAG GCA GAA GTG TCA CAA GAG GTT CGT GTC TGT CCA GGT CGT GGC	3120
	TAT ATC CTT CGT GTC ACA GCA TAT AAA GAG GGA TAT GGA GAG GGC TGC	3168
30	GTA ACG ATC CAT GAG ATC GAA GAC AAT ACA GAC GAA CTG AAA TTC AGC	3216
	AAC TGT GTA GAA GAG GAA GTA TAT CCA AAC AAC ACA GTA ACG TGT AAT	3264
	AAT TAT ACT GGG ACT CAA GAA GAA TAT GAG GGT ACG TAC ACT TCT CGT	3312
	AAT CAA GGA TAT GAC GAA GCC TAT GGT AAT AAC CCT TCC GTA CCA GCT	3360
	GAT TAC GCT TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3408
35	GAG AAT CCT TGT GAA TCT AAC AGA GGC TAT GGG GAT TAC ACA CCA CTA	3456
	CCG GCT GGT TAT GTA ACA AAG GAT TTA GAG TAC TTC CCA GAG ACC GAT	3504
	AAG GTA TGG ATT GAG ATC GGA GAA ACA GAA GGA ACA TTC ATC GTG GAT	3552
	AGC GTG GAA TTA CTC CTT ATG GAG GAA	3579

7. REFERENCES

The following references, to the extent that they provide exemplary procedural or other details supplementary to those set forth herein, are specifically incorporated herein by reference.

- 5 U. S. Patent No. 4,554,101
- U. S. Patent No. 4,683,195
- U. S. Patent No. 4,683,202
- 10 U.S. Patent No. 4,702,914
- U. S. Patent No. 4,757,011
- 15 U. S. Patent No. 4,769,061
- U. S. Patent No. 4,940,835
- U. S. Patent No. 4,965,188
- 20 U. S. Patent No. 4,971,908
- U. S. Patent No. 5,004,863
- 25 U. S. Patent No. 5,015,580
- U. S. Patent No. 5,055,294
- U. S. Patent No. 5,128,130
- 30 U. S. Patent No. 5,176,995
- U. S. Patent No. 5,349,124
- 35 U. S. Patent No. 5,380,831
- U. S. Patent No. 5,384,253
- U. S. Patent No. 5,416,102
- 40 U. S. Patent No. 5,441,884
- U. S. Patent No. 5,449,681

- U. S. Patent No. 5,500,365.
- Intl. Pat. Appl. Publ. No. WO 91/10725, published Jul. 25, 1991.
- 5 Intl. Pat. Appl. Publ. No. WO 93/07278, published Apr. 15, 1993.
- Intl. Pat. Appl. Publ. No. WO 95/02058, published Jan. 19, 1995.
- Intl. Pat. Appl. Publ. No. WO 95/06730, published Mar. 9, 1995.
- 10 Intl. Pat. Appl. Publ. No. WO 95/30752, published Nov. 16, 1995.
- Intl. Pat. Appl. Publ. No. WO 95/30753, published Nov. 16, 1995.
- 15 Abdullah *et al.*, *Biotechnology*, 4:1087, 1986.
- Adelman *et al.*, *DNA*, 2/3:183-193, 1983.
- 20 Allen and Choun, "Large unilamellar liposomes with low uptake into the reticuloendothelial system," *FEBS Lett.*, 223:42-46, 1987.
- Altschul, Stephen F. *et al.*, "Basic local alignment search tool," *J. Mol. Biol.*, 215:403-410, 1990.
- 25 Arvidson *et al.*, *Mol. Biol.*, 3:1533-1534, 1989.
- Baum *et al.*, *Appl. Environ. Microbiol.*, 56:3420-3428, 1990.
- 30 Benbrook *et al.*, In: *Proceedings Bio Expo 1986*, Butterworth, Stoneham, MA, pp. 27-54, 1986.
- Bolivar *et al.*, *Gene*, 2:95, 1977.
- 35 Bytebier *et al.*, *Proc. Natl. Acad. Sci. USA*, 84:5345, 1987.
- Callis *et al.*, *Genes and Development*, 1:1183, 1987.
- 40 Campbell, "Monoclonal Antibody Technology, Laboratory Techniques in Biochemistry and Molecular Biology," Vol. 13, Burden and Von Knippenberg, Eds. pp. 75-83, Elsevier, Amsterdam, 1984.
- Capecchi, M.R., "High efficiency transformation by direct microinjection of DNA into cultured mammalian cells," *Cell* 22(2):479-488, 1980.
- 45 Cashmore *et al.*, *Gen. Eng. of Plants*, Plenum Press, New York, 29-38, 1983.

- Chambers *et al.*, *J. Bacteriol.*, 173:3966-3976, 1991.
- Chang *et al.*, *Nature*, 375:615, 1978.
- 5 Chau *et al.*, *Science*, 244:174-181, 1989.
- Clapp, D.W., "Somatic gene therapy into hematopoietic cells. Current status and future implications," *Clin. Perinatol.* 20(1):155-168, 1993.
- 10 Couvreur *et al.*, "Nanocapsules, a new lysosomotropic carrier," *FEBS Lett.*, 84:323-326, 1977.
- Couvreur, "Polyalkyleanoacrylates as colloidal drug carriers," *Crit. Rev. Ther. Drug*
 15 *Carrier Syst.*, 5:1-20, 1988.
- Crickmore *et al.*, *Abstr. 28th Annu. Meet. Soc. Invert. Pathol.*, Cornell University, Ithaca, NY, 1995.
- 20 Cristou *et al.*, *Plant Physiol*, 87:671-674, 1988.
- Curiel, D. T., Agarwal, S., Wagner, E., and Cotten, M., "Adenovirus enhancement of transferrin-polylysine-mediated gene delivery," *Proc. Natl. Acad. Sci. USA* 88(19):8850-8854, 1991.
- 25 Curiel, D. T., Wagner, E., and Cotten, M., Birnstiel, M. L., Agarwal, S., Li, C. M., Loechel, S., and Hu, P.C. high-efficiency gene transfer mediated by adenovirus coupled to DNA-polylysine complexes," *Hum. Gen. Ther.* 3(2):147-154, 1992.
- 30 Dhir *et al.*, *Plant Cell Reports*, 10:97, 1991.
- Eglitis, M. A., and Anderson, W.F., "Retroviral vectors for introduction of genes into mammalian cells," *Biotechniques* 6(7):608-614, 1988.
- 35 Eglitis, M. A., Kantoff, P. W., Kohn, D. B., Karson, E., Moen, R. C., Lothrop, C. D., Blaese, R. M., and Anderson, W. F., "Retroviral-mediated gene transfer into hemopoietic cells," *Adv. Exp. Med. Biol.* 241:19-27, 1988.
- Eichenlaub, R., *J. Bacteriol.*, 138(2):559-566, 1979.
- 40 Fiers *et al.*, *Nature*, 273:113, 1978.
- Fraley *et al.*, *Biotechnology*, 3:629, 1985.
- 45 Fraley *et al.*, *Proc. Natl. Acad. Sci. USA*, 80:4803, 1983.

- Fromm, M., Taylor, L. P., and Walbot, V., "Expression of genes transferred into monocot and dicot plant cells by electroporation," *Proc. Natl. Acad. Sci. USA* 82(17):5824-5828, 1985.
- 5 Fujimura *et al.*, *Plant Tissue Culture Letters*, 2:74, 1985.
- Fynan, E. F., Webster, R. G., Fuller, D. H., Haynes, J. R., Santoro, J. C., and Robinson, H. L., "DNA vaccines: protective immunizations by parenteral, mucosal, and gene
- 10 gun inoculations," *Proc. Natl. Acad. Sci. USA* 90(24):11478-11482, 1993.
- Gawron-Burke and Baum, *Genet. Engineer.*, 13:237-263, 1991.
- Gefter *et al.*, *Somat. Cell Genet.*, 3:231-236, 1977.
- 15 Gill *et al.*, *J. Biol. Chem.*, 270:27277-27282, 1995.
- Goding, "Monoclonal Antibodies: Principles and Practice," pp. 60-74. 2nd Edition, Academic Press, Orlando, FL, 1986.
- 20 Goeddel *et al.*, *Nature*, 281:544, 1979.
- Goeddel *et al.*, *Nucl. Acids Res.*, 8:4057, 1980.
- 25 Graham, F. L., and van der Eb, A. J., "Transformation of rat cells by DNA of human adenovirus 5," *Virology* 54(2):536-539, 1973.
- Green, *Nucl. Acids Res.* 16(1):369. 1988.
- 30 Grochulski *et al.*, *J. Mol. Biol.*, 254:447-464, 1995.
- Harlow, E. and Lane, D. "Antibodies: A Laboratory Manual," Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1988.
- 35 Harlow, E. and Lane, D. "Antibodies: A Laboratory Manual," Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1988.
- Henry-Michelland *et al.*, "Attachment of antibiotics to nanoparticles; Preparation, drug-release and antimicrobial activity *in vitro*," *Int. J. Pharm.*, 35:121-127, 1987.
- 40 Hess *et al.*, *J. Adv. Enzyme Reg.*, 7:149, 1968.
- Hilber, U. W., Bodmer, M., Smith, F. D., and Koller, W., "Biolistic transformation of conidia of *Botryotinia fuckeliana*," *Curr. Genet.* 25(2):124-127, 1994.
- 45

- Hitzeman *et al.*, *J. Biol. Chem.*, 255:2073, 1980.
- Höfte and Whiteley, *Microbiol. Rev.*, 53:242-255, 1989.
- 5 Holland *et al.*, *Biochemistry*, 17:4900, 1978.
- Honee *et al.*, *Mol. Microbiol.*, 5:2799-2806, 1991.
- 10 Hoover *et al.*, (Eds.), "Remington's Pharmaceutical Sciences," 15th Edition, Mack Publishing Co., Easton, PA, 1975.
- Horsch *et al.*, *Science*, 227:1229-1231, 1985.
- Horton *et al.*, *Gene*, 77:61-68, 1989.
- 15 Humason, "Animal Tissue Techniques," W. H. Freeman and Co., 1967.
- Itakura *et al.*, *Science*, 198:1056, 1977.
- 20 Jameson and Wolf, "The Antigenic Index: A Novel Algorithm for Predicting Antigenic Determinants," *Compu. Appl. Biosci.*, 4(1):181-6, 1988.
- Johnston, S. A., and Tang, D. C., "Gene gun transfection of animal cells and genetic immunization," *Methods Cell. Biol.* 43(A):353-365, 1994.
- 25 Jones, *Genetics*, 85:12 1977.
- Jorgensen *et al.*, *Mol. Gen. Genet.*, 207:471, 1987.
- 30 Keller *et al.*, *EMBO J.*, 8:1309-14, 1989.
- Kingsman *et al.*, *Gene*, 7:141, 1979.
- Klee *et al.*, *Bio/Technology*, 3:637, 1985.
- 35 Klein *et al.*, *Nature*, 327:70, 1987.
- Klein *et al.*, *Proc. Natl. Acad. Sci. USA*, 85:8502-8505, 1988.
- 40 Knight *et al.*, *J. Biol. Chem.*, 270:17765-17770, 1995.
- Kohler and Milstein, *Eur. J. Immunol.* 6:511-519, 1976.
- Kohler and Milstein, *Nature* 256:495-497, 1975.
- 45

- Krzyzek, R. A., "Genetic transformation of maize cells by electroporation of cells pretreated with pectin degrading enzymes," U. S. Patent No. 5,384,253, 01/24/95.
- Kuby, J., *Immunology 2nd Edition*, W. H. Freeman & Company, NY, 1994.
- 5 Kyte, J., and Doolittle, R. F., A simple method for displaying the hydropathic character of a protein," *J. Mol. Biol.* 157(1):105-132, 1982.
- Langridge *et al.*, *Proc. Natl. Acad. Sci. USA*, 86:3219-3223, 1989.
- 10 Lee *et al.*, *Biochem. Biophys. Res. Comm.*, 216:306-312, 1995.
- Lindstrom *et al.*, *Developmental Genetics*, 11:160, 1990.
- 15 Lorz *et al.*, *Mol. Gen. Genet.*, 199:178, 1985.
- Lu, L., Xiao, M., Clapp, D. W., Li, Z. H., and Broxmeyer, H. E., "High efficiency retroviral mediated gene transduction into single isolated immature and replatable CD34(3+) hematopoietic stem/progenitor cells from human umbilical cord blood," *J. Exp. Med.* 178(6):2089-2096, 1993.
- 20 Luo *et al.*, *Plant Mol. Biol. Report.*, 6:165, 1988.
- Maddock *et al.*, *Third Intl. Congr. Plant Mol. Biol.*, Abstr. No. 372, 1991.
- 25 Maloy *et al.*, "Microbial Genetics" 2nd Ed., Jones & Bartlett Publishers, Boston, MA, 1994.
- Maloy, S. R., "Experimental Techniques in Bacterial Genetics" Jones and Bartlett Prokop, A., and Bajpai, R. K. "Recombinant DNA Technology I" *Ann. N. Y. Acad. Sci.* vol. 646, 1991.
- 30 Maniatis *et al.*, "Molecular Cloning: a Laboratory Manual," Cold Spring Harbor Laboratory, Cold Spring Harbor, NY., 1982.
- 35 Marcotte *et al.*, *Nature*, 335:454, 1988.
- Masson *et al.*, *J. Biol. Chem.*, 270:20309-20315, 1995.
- 40 McCabe *et al.*, *Biotechnology*, 6:923, 1988.
- Mettus and Macaluso, *Appl. Environ. Microbiol.*, 56:1128-1134, 1990.
- 45 Nakamura *et al.*, (1987) *Enzyme Immunoassays: Heterogeneous and Homogeneous Systems*, Chapter 27.

- Neuhaus *et al.*, *Theor. Appl. Genet.*, 75:30, 1987.
- Odell *et al.*, *Nature*, 313:810, 1985.
- 5 Omirulleh *et al.*, *Plant Mol. Biol.*, 21:415-428, 1993.
- Pena *et al.*, *Nature*, 325:274, 1987.
- 10 Poszkowski *et al.*, *EMBO J.*, 3:2719, 1989.
- Potrykus *et al.*, *Mol. Gen. Genet.*, 199:183, 1985.
- Poulsen *et al.*, *Mol. Gen. Genet.*, 205:193-200, 1986.
- 15 Prokop, A., Bajpai, R.K., *Ann. N. Y. Acad. Sci.* 646, 1991
- Rogers *et al.*, In: "Methods For Plant Molecular Biology," A. Weissbach and H. Weissbach, eds., Academic Press Inc., San Diego, CA 1988.
- 20 Rogers *et al.*, *Methods Enzymol.*, 153:253-277, 1987.
- Sambrook, J. *et al.*, "Molecular Cloning: A Laboratory Manual," Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989.
- 25 Schnepf *et al.*, *J. Biol. Chem.*, 265:20923-20930, 1990.
- Segal, I. H., "Biochemical Calculations" 2nd Ed., John Wiley & Sons, New York, 1976.
- 30 Simpson, *Science*, 233:34, 1986.
- Spielmann *et al.*, *Mol. Gen. Genet.*, 205:34, 1986.
- Spoerel, *Methods Enzymol.*, 152:588-597, 1987.
- 35 Stinchcomb *et al.*, *Nature*, 282:39, 1979.
- Thompson *et al.*, *Genet. Engineer.*, 17:99-117, 1995.
- 40 Toriyama *et al.*, *Theor. Appl. Genet.*, 73:16, 1986.
- Tschemper *et al.*, *Gene*, 10:157, 1980.
- Uchimiya *et al.*, *Mol. Gen. Genet.*, 204:204, 1986.
- 45

- Van Tunen *et al.*, *EMBO J.*, 7:1257, 1988.
- 5 Vasil *et al.*, "Herbicide-resistant fertile transgenic wheat plants obtained by microprojectile bombardment of regenerable embryogenic callus," *Biotechnology*, 10:667-674, 1992.
- Vasil, *Biotechnology*, 6:397, 1988.
- Vodkin *et al.*, *Cell*, 34:1023, 1983.
- 10 Vogel *et al.*, *J. Cell Biochem.*, (Suppl.) 13D:312, 1989.
- Wagner, E., Zatloukal, K., Cotten, M., Kirlappos, H., Mechtler, K., Curiel, D.T., and Birnstiel, M. L., "Coupling of adenovirus to transferrin-polylysine/DNA complexes greatly enhances receptor-mediated gene delivery and expression of transfected genes," *Proc. Natl. Acad. Sci. USA* 89(13):6099-6103, 1992.
- 15 Weissbach and Weissbach, *Methods for Plant Molecular Biology*, (eds.), Academic Press, Inc., San Diego, CA, 1988.
- 20 Wenzler *et al.*, *Plant Mol. Biol.*, 12:41-50, 1989.
- Wolf *et al.*, "An Integrated Family of Amino Acid Sequence Analysis Programs," *Compu. Appl. Biosci.*, 4(1):187-91, 1988.
- 25 Wong, T. E., and Neumann, E., "Electric field mediated gene transfer," *Biochim. Biophys. Res. Commun.*, 107(2):584-587, 1982.
- Yamada *et al.*, *Plant Cell Rep.*, 4:85, 1986.
- 30 Yang *et al.*, *Proc. Natl. Acad. Sci. USA*, 87:4144-48, 1990.
- Zhou *et al.*, *Methods Enzymol.*, 101:433, 1983.

8. SEQUENCE LISTING

(1) GENERAL INFORMATION:

- 5 (i) APPLICANT: Malvar, Thomas
Gilmer, Amy Jelen
- (ii) TITLE OF INVENTION: HYBRID BACILLUS THURINGIENSIS DELTA-ENDOTOXINE
WITH NOVEL BROAD SPECTRUM INSECTICIDAL ACTIVITY
- 10 (iii) NUMBER OF SEQUENCES: 30
- (iv) CORRESPONDENCE ADDRESS:
15 (A) ADDRESSEE: Arnold, White & Durkee
(B) STREET: P.O. Box 4433
(C) CITY: Houston
(D) STATE: Texas
(E) COUNTRY: United States of America
(F) ZIP: 77210
- 20 (v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
25 (D) SOFTWARE: Patent In Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER: Unknown
(B) FILING DATE: Concurrently Herewith
30 (C) CLASSIFICATION: Unknown
- (viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Kitchell, Barbara S.
(B) REGISTRATION NUMBER: 33,928
35 (C) REFERENCE/DOCKET NUMBER: MOBT:009
- (ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: (512) 418-3000
(B) TELEFAX: (512) 474-7577

40

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
45 (A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

50

GGATAGCACT CATCAAAGGT ACC

23

55

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

5 GAAGATATCC AATTCGAACA GTTTCCTCC 27

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

10 (A) LENGTH: 28 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

CATATTCTGC CTCGAGTGTT GCAGTAAC 28

20 (2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

30 CCCGATCGGC CGCATGC 17

(2) INFORMATION FOR SEQ ID NO:5:

35 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

40 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CATTGGAGCT CTCCATG 17

45 (2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

55 GCACTACGAT GTATCC 16

(2) INFORMATION FOR SEQ ID NO:7:

60 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CATCGTAGTG CAACTCTTAC

20

10

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

15

(A) LENGTH: 39 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

20

CCAAGAAAAT ACTAGAGCTC TTGTTAAAAA AGGTGTTCC

39

(2) INFORMATION FOR SEQ ID NO:9:

25

(i) SEQUENCE CHARACTERISTICS:

30

(A) LENGTH: 3531 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ix) FEATURE:

35

(A) NAME/KEY: CDS
 (B) LOCATION: 1..3531

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

40

ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA
 Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
 1 5 10 15

48

AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT
 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
 20 25 30

96

45

TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT
 Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
 35 40 45

144

50

GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA
 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
 50 55 60

192

55

TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT
 Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
 65 70 75 80

240

60

GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC
 Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
 85 90 95

288

	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
	Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu	
	100 105 110	
5	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu	
	115 120 125	
10	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCG CTT ACA ACC GCT	432
	Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala	
	130 135 140	
15	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val	
	145 150 155 160	
20	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser	
	165 170 175	
25	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
	Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg	
	180 185 190	
30	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val	
	195 200 205	
35	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg	
	210 215 220	
40	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val	
	225 230 235 240	
45	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro	
	245 250 255	
50	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	
	260 265 270	
55	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	
	275 280 285	
60	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	
	290 295 300	
65	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln	
	305 310 315 320	
70	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	
	325 330 335	
75	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056

	Leu	Tyr	Gly	Thr	Met	Gly	Asn	Ala	Ala	Pro	Gln	Gln	Arg	Ile	Val	Ala	
				340					345					350			
5	CAA	CTA	GGT	CAG	GGC	GTG	TAT	AGA	ACA	TTA	TCG	TCC	ACT	TTA	TAT	AGA	1104
	Gln	Leu	Gly	Gln	Gly	Val	Tyr	Arg	Thr	Leu	Ser	Ser	Thr	Leu	Tyr	Arg	
			355					360					365				
10	AGA	CCT	TTT	AAT	ATA	GGG	ATA	AAT	AAT	CAA	CAA	CTA	TCT	GTT	CTT	GAC	1152
	Arg	Pro	Phe	Asn	Ile	Gly	Ile	Asn	Asn	Gln	Gln	Leu	Ser	Val	Leu	Asp	
		370					375					380					
15	GGG	ACA	GAA	TTT	GCT	TAT	GGA	ACC	TCC	TCA	AAT	TTG	CCA	TCC	GCT	GTA	1200
	Gly	Thr	Glu	Phe	Ala	Tyr	Gly	Thr	Ser	Ser	Asn	Leu	Pro	Ser	Ala	Val	
	385					390					395					400	
20	TAC	AGA	AAA	AGC	GGA	ACG	GTA	GAT	TCG	CTG	GAT	GAA	ATA	CCG	CCA	CAG	1248
	Tyr	Arg	Lys	Ser	Gly	Thr	Val	Asp	Ser	Leu	Asp	Glu	Ile	Pro	Pro	Gln	
					405					410					415		
25	AAT	AAC	AAC	GTG	CCA	CCT	AGG	CAA	GGA	TTT	AGT	CAT	CGA	TTA	AGC	CAT	1296
	Asn	Asn	Asn	Val	Pro	Pro	Arg	Gln	Gly	Phe	Ser	His	Arg	Leu	Ser	His	
				420					425					430			
30	GTT	TCA	ATG	TTT	CGT	TCA	GGC	TTT	AGT	AAT	AGT	AGT	GTA	AGT	ATA	ATA	1344
	Val	Ser	Met	Phe	Arg	Ser	Gly	Phe	Ser	Asn	Ser	Ser	Val	Ser	Ile	Ile	
			435				440						445				
35	AGA	GCT	CCA	ATG	TTT	TCT	TGG	ACG	CAC	CGT	AGT	GCA	ACC	CCT	ACA	AAT	1392
	Arg	Ala	Pro	Met	Phe	Ser	Trp	Thr	His	Arg	Ser	Ala	Thr	Pro	Thr	Asn	
		450					455					460					
40	ACA	ATT	GAT	CCG	GAG	AGG	ATT	ACT	CAA	ATA	CCA	TTG	GTA	AAA	GCA	CAT	1440
	Thr	Ile	Asp	Pro	Glu	Arg	Ile	Thr	Gln	Ile	Pro	Leu	Val	Lys	Ala	His	
	465					470				475						480	
45	ACA	CTT	CAG	TCA	GGT	ACT	ACT	GTT	GTA	AGA	GGG	CCC	GGG	TTT	ACG	GGA	1488
	Thr	Leu	Gln	Ser	Gly	Thr	Thr	Val	Val	Arg	Gly	Pro	Gly	Phe	Thr	Gly	
					485					490					495		
50	GGA	GAT	ATT	CTT	CGA	CGA	ACA	AGT	GGA	GGA	CCA	TTT	GCT	TAT	ACT	ATT	1536
	Gly	Asp	Ile	Leu	Arg	Arg	Thr	Ser	Gly	Gly	Pro	Phe	Ala	Tyr	Thr	Ile	
				500					505					510			
55	GTT	AAT	ATA	AAT	GGG	CAA	TTA	CCC	CAA	AGG	TAT	CGT	GCA	AGA	ATA	CGC	1584
	Val	Asn	Ile	Asn	Gly	Gln	Leu	Pro	Gln	Arg	Tyr	Arg	Ala	Arg	Ile	Arg	
			515					520					525				
60	TAT	GCC	TCT	ACT	ACA	AAT	CTA	AGA	ATT	TAC	GTA	ACG	GTT	GCA	CGT	GAA	1632
	Tyr	Ala	Ser	Thr	Thr	Asn	Leu	Arg	Ile	Tyr	Val	Thr	Val	Ala	Gly	Glu	
		530					535					540					
65	CGG	ATT	TTT	GCT	GGT	CAA	TTT	AAC	AAA	ACA	ATG	GAT	ACC	GGT	GAC	CCA	1680
	Arg	Ile	Phe	Ala	Gly	Gln	Phe	Asn	Lys	Thr	Met	Asp	Thr	Gly	Asp	Pro	
	545					550					555					560	
70	TTA	ACA	TTC	CAA	TCT	TTT	AGT	TAC	GCA	ACT	ATT	AAT	ACA	GCT	TTT	ACA	1728
	Leu	Thr	Phe	Gln	Ser	Phe	Ser	Tyr	Ala	Thr	Ile	Asn	Thr	Ala	Phe	Thr	
					565					570					575		
75	TTC	CCA	ATG	AGC	CAG	AGT	AGT	TTC	ACA	GTA	GGT	GCT	GAT	ACT	TTT	AGT	1776
	Phe	Pro	Met	Ser	Gln	Ser	Ser	Phe	Thr	Val	Gly	Ala	Asp	Thr	Phe	Ser	

	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	11544
	Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile	
	835 840 845	
5	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	11592
	Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu	
	850 855 860	
10	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	11640
	Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys	
	865 870 875 880	
15	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	11688
	Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu	
	885 890 895	
20	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	11736
	Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe	
	900 905 910	
	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	11784
	Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met	
	915 920 925	
25	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	11832
	Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu	
	930 935 940	
30	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	11880
	Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu	
	945 950 955 960	
35	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	11928
	Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn	
	965 970 975	
40	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	11976
	Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val	
	980 985 990	
	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	12024
	Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu	
	995 1000 1005	
45	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	12072
	Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys	
	1010 1015 1020	
50	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	12120
	Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr	
	1025 1030 1035 1040	
55	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	12168
	Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu	
	1045 1050 1055	
60	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	12216
	Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr	
	1060 1065 1070	

Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu
 115 120 125

5 Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala
 130 135 140

Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val
 145 150 155 160

10 Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser
 165 170 175

Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg
 180 185 190

15 Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val
 195 200 205

Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg
 210 215 220

20 Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val
 225 230 235 240

25 Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro
 245 250 255

Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val
 260 265 270

30 Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu
 275 280 285

Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr
 290 295 300

35 Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln
 305 310 315 320

40 Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro
 325 330 335

Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala
 340 345 350

45 Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg
 355 360 365

Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp
 370 375 380

50 Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val
 385 390 395 400

55 Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln
 405 410 415

Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His
 420 425 430

60 Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile

		435		440		445			
	Arg	Ala	Pro	Met	Phe	Ser	Trp	Thr	His
	450						455		
5	Thr	Ile	Asp	Pro	Glu	Arg	Ile	Thr	Gln
	465					470			475
	Thr	Leu	Gln	Ser	Gly	Thr	Thr	Val	Val
10					485				490
	Gly	Asp	Ile	Leu	Arg	Arg	Thr	Ser	Gly
				500					505
	Val	Asn	Ile	Asn	Gly	Gln	Leu	Pro	Gln
15								520	
	Tyr	Ala	Ser	Thr	Thr	Asn	Leu	Arg	Ile
	530						535		
20	Arg	Ile	Phe	Ala	Gly	Gln	Phe	Asn	Lys
	545					550			555
	Leu	Thr	Phe	Gln	Ser	Phe	Ser	Tyr	Ala
25						565			570
	Phe	Pro	Met	Ser	Gln	Ser	Ser	Phe	Thr
				580				585	
30	Ser	Gly	Asn	Glu	Val	Tyr	Ile	Asp	Arg
			595					600	
	Ala	Thr	Phe	Glu	Ala	Glu	Tyr	Asp	Leu
	610						615		
35	Asn	Ala	Leu	Phe	Thr	Ser	Ile	Asn	Gln
	625					630			635
	Thr	Asp	Tyr	His	Ile	Asp	Gln	Val	Ser
40					645				650
	Asp	Glu	Phe	Cys	Leu	Asp	Glu	Lys	Arg
				660					665
45	His	Ala	Lys	Arg	Leu	Ser	Asp	Glu	Arg
			675					680	
	Phe	Lys	Gly	Ile	Asn	Arg	Gln	Leu	Asp
	690						695		
50	Asp	Ile	Thr	Ile	Gln	Arg	Gly	Asp	Asp
	705					710			
	Thr	Leu	Pro	Gly	Thr	Phe	Asp	Glu	Cys
55								725	
	Lys	Ile	Asp	Glu	Ser	Lys	Leu	Lys	Ala
				740					745
60	Gly	Tyr	Ile	Glu	Asp	Ser	Gln	Asp	Leu
			755					760	

	Asn	Ala	Lys	His	Glu	Thr	Val	Asn	Val	Pro	Gly	Thr	Gly	Ser	Leu	Trp	
	770						775					780					
5	Pro	Leu	Ser	Ala	Gln	Ser	Pro	Ile	Gly	Lys	Cys	Gly	Glu	Pro	Asn	Arg	
	785					790					795					800	
	Cys	Ala	Pro	His	Leu	Glu	Trp	Asn	Pro	Asp	Leu	Asp	Cys	Ser	Cys	Arg	
					805					810						815	
10	Asp	Gly	Glu	Lys	Cys	Ala	His	His	Ser	His	His	Phe	Ser	Leu	Asp	Ile	
					820				825						830		
	Asp	Val	Gly	Cys	Thr	Asp	Leu	Asn	Glu	Asp	Leu	Gly	Val	Trp	Val	Ile	
15								840					845				
	Phe	Lys	Ile	Lys	Thr	Gln	Asp	Gly	His	Ala	Arg	Leu	Gly	Asn	Leu	Glu	
	850						855					860					
20	Phe	Leu	Glu	Glu	Lys	Pro	Leu	Val	Gly	Glu	Ala	Leu	Ala	Arg	Val	Lys	
	865					870					875					880	
	Arg	Ala	Glu	Lys	Lys	Trp	Arg	Asp	Lys	Arg	Glu	Lys	Leu	Glu	Trp	Glu	
					885					890					895		
25	Thr	Asn	Ile	Val	Tyr	Lys	Glu	Ala	Lys	Glu	Ser	Val	Asp	Ala	Leu	Phe	
				900					905					910			
	Val	Asn	Ser	Gln	Tyr	Asp	Gln	Leu	Gln	Ala	Asp	Thr	Asn	Ile	Ala	Met	
30								920					925				
	Ile	His	Ala	Ala	Asp	Lys	Arg	Val	His	Ser	Ile	Arg	Glu	Ala	Tyr	Leu	
	930						935					940					
35	Pro	Glu	Leu	Ser	Val	Ile	Pro	Gly	Val	Asn	Ala	Ala	Ile	Phe	Glu	Glu	
	945					950					955					960	
	Leu	Glu	Gly	Arg	Ile	Phe	Thr	Ala	Phe	Ser	Leu	Tyr	Asp	Ala	Arg	Asn	
					965					970					975		
40	Val	Ile	Lys	Asn	Gly	Asp	Phe	Asn	Asn	Gly	Leu	Ser	Cys	Trp	Asn	Val	
				980					985					990			
	Lys	Gly	His	Val	Asp	Val	Glu	Glu	Gln	Asn	Asn	Gln	Arg	Ser	Val	Leu	
45				995				1000					1005				
	Val	Val	Pro	Glu	Trp	Glu	Ala	Glu	Val	Ser	Gln	Glu	Val	Arg	Val	Cys	
							1015					1020					
50	Pro	Gly	Arg	Gly	Tyr	Ile	Leu	Arg	Val	Thr	Ala	Tyr	Lys	Glu	Gly	Tyr	
	1025					1030					1035					1040	
	Gly	Glu	Gly	Cys	Val	Thr	Ile	His	Glu	Ile	Glu	Asn	Asn	Thr	Asp	Glu	
					1045					1050					1055		
55	Leu	Lys	Phe	Ser	Asn	Cys	Val	Glu	Glu	Glu	Ile	Tyr	Pro	Asn	Asn	Thr	
					1060				1065					1070			
	Val	Thr	Cys	Asn	Asp	Tyr	Thr	Val	Asn	Gln	Glu	Glu	Tyr	Gly	Gly	Ala	
60					1075				1080					1085			

Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala
 1090 1095 1100

5 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg
 1105 1110 1115 1120

Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu
 1125 1130 1135

10 Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp
 1140 1145 1150

Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
 1155 1160 1165

15 Ser Val Glu Leu Leu Leu Met Glu Glu
 1170 1175

20 (2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 3531 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

25 (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 1..3531

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

35 ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA 48
 Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
 1 5 10 15

40 AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT 96
 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
 20 25 30

45 TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT 144
 Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu-Ser
 35 40 45

50 GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA - 192
 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
 50 55 60

55 TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT 240
 Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
 65 70 75 80

60 GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC 288
 Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
 85 90 95

ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA 336
 Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
 100 105 110

	TCT	TTT	AGA	GAG	TGG	GAA	GCA	GAT	CCT	ACT	AAT	CCA	GCA	TTA	AGA	GAA	384
	Ser	Phe	Arg	Glu	Trp	Glu	Ala	Asp	Pro	Thr	Asn	Pro	Ala	Leu	Arg	Glu	
			115					120					125				
5	GAG	ATG	CGT	ATT	CAA	TTC	AAT	GAC	ATG	AAC	AGT	GCC	CTT	ACA	ACC	GCT	432
	Glu	Met	Arg	Ile	Gln	Phe	Asn	Asp	Met	Asn	Ser	Ala	Leu	Thr	Thr	Ala	
		130					135					140					
10	ATT	CCT	CTT	TTT	GCA	GTT	CAA	AAT	TAT	CAA	GTT	CCT	CTT	TTA	TCA	GTA	480
	Ile	Pro	Leu	Phe	Ala	Val	Gln	Asn	Tyr	Gln	Val	Pro	Leu	Leu	Ser	Val	
	145					150					155					160	
15	TAT	GTT	CAA	GCT	GCA	AAT	TTA	CAT	TTA	TCA	GTT	TTG	AGA	GAT	GTT	TCA	528
	Tyr	Val	Gln	Ala	Ala	Asn	Leu	His	Leu	Ser	Val	Leu	Arg	Asp	Val	Ser	
				165						170					175		
20	GTG	TTT	GGA	CAA	AGG	TGG	GGA	TTT	GAT	GCC	GCG	ACT	ATC	AAT	AGT	CGT	576
	Val	Phe	Gly	Gln	Arg	Trp	Gly	Phe	Asp	Ala	Ala	Thr	Ile	Asn	Ser	Arg	
			180						185					190			
25	TAT	AAT	GAT	TTA	ACT	AGG	CTT	ATT	GGC	AAC	TAT	ACA	GAT	TAT	GCT	GTA	624
	Tyr	Asn	Asp	Leu	Thr	Arg	Leu	Ile	Gly	Asn	Tyr	Thr	Asp	Tyr	Ala	Val	
			195				200						205				
30	CGC	TGG	TAC	AAT	ACG	GGA	TTA	GAA	CGT	GTA	TGG	GGA	CCG	GAT	TCT	AGA	672
	Arg	Trp	Tyr	Asn	Thr	Gly	Leu	Glu	Arg	Val	Trp	Gly	Pro	Asp	Ser	Arg	
		210					215					220					
35	GAT	TGG	GTA	AGG	TAT	AAT	CAA	TTT	AGA	AGA	GAA	TTA	ACA	CTA	ACT	GTA	720
	Asp	Trp	Val	Arg	Tyr	Asn	Gln	Phe	Arg	Arg	Glu	Leu	Thr	Leu	Thr	Val	
	225					230					235				240		
40	TTA	GAT	ATC	GTT	GCT	CTG	TTC	CCG	AAT	TAT	GAT	AGT	AGA	AGA	TAT	CCA	768
	Leu	Asp	Ile	Val	Ala	Leu	Phe	Pro	Asn	Tyr	Asp	Ser	Arg	Arg	Tyr	Pro	
				245					250						255		
45	ATT	CGA	ACA	GTT	TCC	CAA	TTA	ACA	AGA	GAA	ATT	TAT	ACA	AAC	CCA	GTA	816
	Ile	Arg	Thr	Val	Ser	Gln	Leu	Thr	Arg	Glu	Ile	Tyr	Thr	Asn	Pro	Val	
			260						265					270			
50	TTA	GAA	AAT	TTT	GAT	GGT	AGT	TTT	CGA	GGC	TCG	GCT	CAG	GGC	ATA	GAA	864
	Leu	Glu	Asn	Phe	Asp	Gly	Ser	Phe	Arg	Gly	Ser	Ala	Gln	Gly	Ile	Glu	
			275					280					285				
55	AGA	AGT	ATT	AGG	AGT	CCA	CAT	TTG	ATG	GAT	ATA	CTT	AAC	AGT	ATA	ACC	912
	Arg	Ser	Ile	Arg	Ser	Pro	His	Leu	Met	Asp	Ile	Leu	Asn	Ser	Ile	Thr	
		290					295					300					
60	ATC	TAT	ACG	GAT	GCT	CAT	AGG	GGT	TAT	TAT	TAT	TGG	TCA	GGG	CAT	CAA	960
	Ile	Tyr	Thr	Asp	Ala	His	Arg	Gly	Tyr	Tyr	Tyr	Trp	Ser	Gly	His	Gln	
	305					310					315					320	
65	ATA	ATG	GCT	TCT	CCT	GTA	GGG	TTT	TCG	GGG	CCA	GAA	TTC	ACT	TTT	CCG	1008
	Ile	Met	Ala	Ser	Pro	Val	Gly	Phe	Ser	Gly	Pro	Glu	Phe	Thr	Phe	Pro	
				325						330					335		
70	CTA	TAT	GGA	ACT	ATG	GGA	AAT	GCA	GCT	CCA	CAA	CAA	CGT	ATT	GTT	GCT	1056
	Leu	Tyr	Gly	Thr	Met	Gly	Asn	Ala	Ala	Pro	Gln	Gln	Arg	Ile	Val	Ala	
				340					345					350			
75	CAA	CTA	GGT	CAG	GGC	GTG	TAT	AGA	ACA	TTA	TCG	TCC	ACT	TTA	TAT	AGA	1104

	Gln	Leu	Gly	Gln	Gly	Val	Tyr	Arg	Thr	Leu	Ser	Ser	Thr	Leu	Tyr	Arg	
			355					360					365				
5	AGA	CCT	TTT	AAT	ATA	GGG	ATA	AAT	AAT	CAA	CAA	CTA	TCT	GTT	CTT	GAC	115
	Arg	Pro	Phe	Asn	Ile	Gly	Ile	Asn	Asn	Gln	Gln	Leu	Ser	Val	Leu	Asp	
		370					375					380					
10	GGG	ACA	GAA	TTT	GCT	TAT	GGA	ACC	TCC	TCA	AAT	TTG	CCA	TCC	GCT	GTA	120
	Gly	Thr	Glu	Phe	Ala	Tyr	Gly	Thr	Ser	Ser	Asn	Leu	Pro	Ser	Ala	Val	
		385				390					395					400	
15	TAC	AGA	AAA	AGC	GGA	ACG	GTA	GAT	TCG	CTG	GAT	GAA	ATA	CCG	CCA	CAG	124
	Tyr	Arg	Lys	Ser	Gly	Thr	Val	Asp	Ser	Leu	Asp	Glu	Ile	Pro	Pro	Gln	
				405					410						415		
20	AAT	AAC	AAC	GTG	CCA	CCT	AGG	CAA	GGA	TTT	AGT	CAT	CGA	TTA	AGC	CAT	129
	Asn	Asn	Asn	Val	Pro	Pro	Arg	Gln	Gly	Phe	Ser	His	Arg	Leu	Ser	His	
				420					425					430			
25	GTT	TCA	ATG	TTT	CGT	TCA	GGC	TTT	AGT	AAT	AGT	AGT	GTA	AGT	ATA	ATA	134
	Val	Ser	Met	Phe	Arg	Ser	Gly	Phe	Ser	Asn	Ser	Ser	Val	Ser	Ile	Ile	
			435				440						445				
30	AGA	GCT	CCA	ATG	TTT	TCT	TGG	ACG	CAC	CGT	AGT	GCA	ACC	CCT	ACA	AAT	139
	Arg	Ala	Pro	Met	Phe	Ser	Trp	Thr	His	Arg	Ser	Ala	Thr	Pro	Thr	Asn	
		450					455					460					
35	ACA	ATT	GAT	CCG	GAG	AGG	ATT	ACT	CAA	ATA	CCA	TTG	GTA	AAA	GCA	CAT	144
	Thr	Ile	Asp	Pro	Glu	Arg	Ile	Thr	Gln	Ile	Pro	Leu	Val	Lys	Ala	His	
		465				470					475					480	
40	ACA	CTT	CAG	TCA	GGT	ACT	ACT	GTT	GTA	AGA	GGG	CCC	GGG	TTT	ACG	GGA	148
	Thr	Leu	Gln	Ser	Gly	Thr	Thr	Val	Val	Arg	Gly	Pro	Gly	Phe	Thr	Gly	
					485					490					495		
45	GGA	GAT	ATT	CTT	CGA	CGA	ACA	AGT	GGA	GGA	CCA	TTT	GCT	TAT	ACT	ATT	153
	Gly	Asp	Ile	Leu	Arg	Arg	Thr	Ser	Gly	Gly	Pro	Phe	Ala	Tyr	Thr	Ile	
				500					505					510			
50	GTT	AAT	ATA	AAT	GGG	CAA	TTA	CCC	CAA	AGG	TAT	CGT	GCA	AGA	ATA	CGC	158
	Val	Asn	Ile	Asn	Gly	Gln	Leu	Pro	Gln	Arg	Tyr	Arg	Ala	Arg	Ile	Arg	
			515					520					525				
55	TAT	GCC	TCT	ACT	ACA	AAT	CTA	AGA	ATT	TAC	GTA	ACG	GTT	GCA	GGT	GAA	163
	Tyr	Ala	Ser	Thr	Thr	Asn	Leu	Arg	Ile	Tyr	Val	Thr	Val	Ala	Gly	Glu	
		530					535					540					
60	CGG	ATT	TTT	GCT	GGT	CAA	TTT	AAC	AAA	ACA	ATG	GAT	ACC	GGT	GAC	CCA	168
	Arg	Ile	Phe	Ala	Gly	Gln	Phe	Asn	Lys	Thr	Met	Asp	Thr	Gly	Asp	Pro	
		545				550					555					560	
65	TTA	ACA	TTC	CAA	TCT	TTT	AGT	TAC	GCA	ACT	ATT	AAT	ACA	GCT	TTT	ACA	172
	Leu	Thr	Phe	Gln	Ser	Phe	Ser	Tyr	Ala	Thr	Ile	Asn	Thr	Ala	Phe	Thr	
					565				570						575		
70	TTC	CCA	ATG	AGC	CAG	AGT	AGT	TTC	ACA	GTA	GGT	GCT	GAT	ACT	TTT	AGT	177
	Phe	Pro	Met	Ser	Gln	Ser	Ser	Phe	Thr	Val	Gly	Ala	Asp	Thr	Phe	Ser	
				580					585					590			
75	TCA	GGG	AAT	GAA	GTT	TAT	ATA	GAC	AGA	TTT	GAA	TTG	ATT	CCA	GTT	ACT	182
	Ser	Gly	Asn	Glu	Val	Tyr	Ile	Asp	Arg	Phe	Glu	Leu	Ile	Pro	Val	Thr	

		595				600						605							
		GCA	ACA	CTC	GAG	GCT	GAA	TAT	AAT	CTG	GAA	AGA	GCG	CAG	AAG	GCG	GTG		1872
		Ala	Thr	Leu	Glu	Ala	Glu	Tyr	Asn	Leu	Glu	Arg	Ala	Gln	Lys	Ala	Val		
5		610						615					620						
		AAT	GCG	CTG	TTT	ACG	TCT	ACA	AAC	CAA	CTA	GGG	CTA	AAA	ACA	AAT	GTA		1920
		Asn	Ala	Leu	Phe	Thr	Ser	Thr	Asn	Gln	Leu	Gly	Leu	Lys	Thr	Asn	Val		
		625					630					635					640		
10		ACG	GAT	TAT	CAT	ATT	GAT	CAA	GTG	TCC	AAT	TTA	GTT	ACG	TAT	TTA	TCG		1968
		Thr	Asp	Tyr	His	Ile	Asp	Gln	Val	Ser	Asn	Leu	Val	Thr	Tyr	Leu	Ser		
						645					650					655			
15		GAT	GAA	TTT	TGT	CTG	GAT	GAA	AAG	CGA	GAA	TTG	TCC	GAG	AAA	GTC	AAA		2016
		Asp	Glu	Phe	Cys	Leu	Asp	Glu	Lys	Arg	Glu	Leu	Ser	Glu	Lys	Val	Lys		
					660					665					670				
		CAT	GCG	AAG	CGA	CTC	AGT	GAT	GAA	CGC	AAT	TTA	CTC	CAA	GAT	TCA	AAT		2064
20		His	Ala	Lys	Arg	Leu	Ser	Asp	Glu	Arg	Asn	Leu	Leu	Gln	Asp	Ser	Asn		
				675						680					685				
		TTC	AAA	GAC	ATT	AAT	AGG	CAA	CCA	GAA	CGT	GGG	TGG	GGC	GGA	AGT	ACA		2112
		Phe	Lys	Asp	Ile	Asn	Arg	Gln	Pro	Glu	Arg	Gly	Trp	Gly	Gly	Ser	Thr		
25		690						695					700						
		GGG	ATT	ACC	ATC	CAA	GGA	GGG	GAT	GAC	GTA	TTT	AAA	GAA	AAT	TAC	GTC		2160
		Gly	Ile	Thr	Ile	Gln	Gly	Gly	Asp	Asp	Val	Phe	Lys	Glu	Asn	Tyr	Val		
		705					710					715					720		
30		ACA	CTA	TCA	GGT	ACC	TTT	GAT	GAG	TGC	TAT	CCA	ACA	TAT	TTG	TAT	CAA		2208
		Thr	Leu	Ser	Gly	Thr	Phe	Asp	Glu	Cys	Tyr	Pro	Thr	Tyr	Leu	Tyr	Gln		
						725					730					735			
35		AAA	ATC	GAT	GAA	TCA	AAA	TTA	AAA	GCC	TTT	ACC	CGT	TAT	CAA	TTA	AGA		2256
		Lys	Ile	Asp	Glu	Ser	Lys	Leu	Lys	Ala	Phe	Thr	Arg	Tyr	Gln	Leu	Arg		
				740						745					750				
		GGG	TAT	ATC	GAA	GAT	AGT	CAA	GAC	TTA	GAA	ATC	TAT	TTA	ATT	CGC	TAC		2304
40		Gly	Tyr	Ile	Glu	Asp	Ser	Gln	Asp	Leu	Glu	Ile	Tyr	Leu	Ile	Arg	Tyr		
				755					760					765					
		AAT	GCA	AAA	CAT	GAA	ACA	GTA	AAT	GTG	CCA	GGT	ACG	GGT	TCC	TTA	TGG		2352
		Asn	Ala	Lys	His	Glu	Thr	Val	Asn	Val	Pro	Gly	Thr	Gly	Ser	Leu	Trp		
45		770						775					780						
		CCG	CTT	TCA	GCC	CAA	AGT	CCA	ATC	GGA	AAG	TGT	GGA	GAG	CCG	AAT	CGA		2400
		Pro	Leu	Ser	Ala	Gln	Ser	Pro	Ile	Gly	Lys	Cys	Gly	Glu	Pro	Asn	Arg		
		785					790					795					800		
50		TGC	GCG	CCA	CAC	CTT	GAA	TGG	AAT	CCT	GAC	TTA	GAT	TGT	TCG	TGT	AGG		2448
		Cys	Ala	Pro	His	Leu	Glu	Trp	Asn	Pro	Asp	Leu	Asp	Cys	Ser	Cys	Arg		
						805					810					815			
55		GAT	GGA	GAA	AAG	TGT	GCC	CAT	CAT	TCG	CAT	CAT	TTC	TCC	TTA	GAC	ATT		2496
		Asp	Gly	Glu	Lys	Cys	Ala	His	His	Ser	His	His	Phe	Ser	Leu	Asp	Ile		
					820					825					830				
60		GAT	GTA	GGA	TGT	ACA	GAC	TTA	AAT	GAG	GAC	CTA	GGT	GTA	TGG	GTG	ATC		2544
		Asp	Val	Gly	Cys	Thr	Asp	Leu	Asn	Glu	Asp	Leu	Gly	Val	Trp	Val	Ile		
				835					840					845					

	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	15-92
	Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu	
	850 855 860	
5	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2-10
	Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys	
	865 870 875 880	
10	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	8
	Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu	
	885 890 895	
15	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	1-36
	Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe	
	900 905 910	
20	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2-84
	Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met	
	915 920 925	
25	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	1-32
	Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu	
	930 935 940	
30	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	1-30
	Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu	
	945 950 955 960	
35	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	15-28
	Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn	
	965 970 975	
40	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	1-76
	Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val	
	980 985 990	
45	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	1-14
	Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu	
	995 1000 1005	
50	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	1-72
	Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys	
	1010 1015 1020	
55	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	1-20
	Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr	
	1025 1030 1035 1040	
60	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	1-58
	Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu	
	1045 1050 1055	
65	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	1-6
	Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr	
	1060 1065 1070	
70	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	1-64
	Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala	
	1075 1080 1085	

5 TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT 3312
 Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala
 1090 1095 1100
 5 GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA 3360
 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg
 1105 1110 1115 1120
 10 GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA 3408
 Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu
 1125 1130 1135
 15 CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT 3456
 Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp
 1140 1145 1150
 AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC 3504
 Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
 1155 1160 1165
 20 AGC GTG GAA TTA CTC CTT ATG GAG GAA 3531
 Ser Val Glu Leu Leu Leu Met Glu Glu
 1170 1175

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

30 (A) LENGTH: 1177 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

35 Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
 1 5 10 15
 40 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
 20 25 30
 Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
 35 40 45
 45 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
 50 55 60
 50 Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
 65 70 75 80
 Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
 85 90 95
 55 Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
 100 105 110
 Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu
 115 120 125
 60 Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala

	130		135		140
	Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val				
	145		150		155 160
5	Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser				
		165		170	175
10	Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg				
		180		185	190
	Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val				
		195		200	205
15	Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg				
		210		215	220
	Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val				
		225		230	235 240
20	Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro				
		245		250	255
	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val				
		260		265	270
25	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu				
		275		280	285
30	Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr				
		290		295	300
	Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln				
		305		310	315 320
35	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro				
		325		330	335
	Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala				
		340		345	350
40	Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg				
		355		360	365
45	Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp				
		370		375	380
	Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val				
		385		390	395 400
50	Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln				
		405		410	415
	Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His				
		420		425	430
55	Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile				
		435		440	445
60	Arg Ala Pro Met Phe Ser Trp Thr His Arg Ser Ala Thr Pro Thr Asn				
		450		455	460

Thr Ile Asp Pro Glu Arg Ile Thr Gln Ile Pro Leu Val Lys Ala His
 465 470 475 480
 5 Thr Leu Gln Ser Gly Thr Thr Val Val Arg Gly Pro Gly Phe Thr Gly
 485 490 495
 Gly Asp Ile Leu Arg Arg Thr Ser Gly Gly Pro Phe Ala Tyr Thr Ile
 500 505 510
 10 Val Asn Ile Asn Gly Gln Leu Pro Gln Arg Tyr Arg Ala Arg Ile Arg
 515 520 525
 Tyr Ala Ser Thr Thr Asn Leu Arg Ile Tyr Val Thr Val Ala Gly Glu
 530 535 540
 15 Arg Ile Phe Ala Gly Gln Phe Asn Lys Thr Met Asp Thr Gly Asp Pro
 545 550 555 560
 20 Leu Thr Phe Gln Ser Phe Ser Tyr Ala Thr Ile Asn Thr Ala Phe Thr
 565 570 575
 Phe Pro Met Ser Gln Ser Ser Phe Thr Val Gly Ala Asp Thr Phe Ser
 580 585 590
 25 Ser Gly Asn Glu Val Tyr Ile Asp Arg Phe Glu Leu Ile Pro Val Thr
 595 600 605
 Ala Thr Leu Glu Ala Glu Tyr Asn Leu Glu Arg Ala Gln Lys Ala Val
 610 615 620
 30 Asn Ala Leu Phe Thr Ser Thr Asn Gln Leu Gly Leu Lys Thr Asn Val
 625 630 635 640
 35 Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Thr Tyr Leu Ser
 645 650 655
 Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys
 660 665 670
 40 His Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Ser Asn
 675 680 685
 Phe Lys Asp Ile Asn Arg Gln Pro Glu Arg Gly Trp Gly Gly Ser Thr
 690 695 700
 45 Gly Ile Thr Ile Gln Gly Gly Asp Asp Val Phe Lys Glu Asn Tyr Val
 705 710 715 720
 50 Thr Leu Ser Gly Thr Phe Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln
 725 730 735
 Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg
 740 745 750
 55 Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr
 755 760 765
 60 Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp
 770 775 780

Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg
 785 790 795 800
 5 Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg
 805 810 815
 Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu Asp Ile
 820 825 830
 10 Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile
 835 840 845
 Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu
 850 855 860
 15 Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys
 865 870 875 880
 Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu
 885 890 895
 20 Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe
 900 905 910
 Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met
 915 920 925
 25 Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu
 930 935 940
 30 Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu
 945 950 955 960
 Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn
 965 970 975
 35 Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val
 980 985 990
 Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu
 995 1000 1005
 Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys
 1010 1015 1020
 45 Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr
 1025 1030 1035 1040
 Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu
 1045 1050 1055
 50 Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr
 1060 1065 1070
 Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala
 1075 1080 1085
 Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala
 1090 1095 1100
 60 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg

1105 1110 1115 1120

Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu
 1125 1130 1135

5 Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp
 1140 1145 1150

10 Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
 1155 1160 1165

Ser Val Glu Leu Leu Leu Met Glu Glu
 1170 1175

15

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

20 (A) LENGTH: 3531 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ix) FEATURE:

25 (A) NAME/KEY: CDS
 (B) LOCATION: 1..3531

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

30 ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA 48
 Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
 1 5 10 15

35 AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT 96
 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
 20 25 30

40 TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT 144
 Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
 35 40 45

45 GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA 192
 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
 50 55 60

50 TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT 240
 Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
 65 70 75 80

55 GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC 288
 Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
 85 90 95

60 ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA 336
 Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
 100 105 110

 TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA 384
 Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu
 115 120 125

	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala	
	130 135 140	
5	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val	
	145 150 155 160	
10	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser	
	165 170 175	
15	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
	Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg	
	180 185 190	
20	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT CAT GCT GTA	624
	Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp His Ala Val	
	195 200 205	
25	CGC TGG TAC AAT ACG GGA TTA GAG CGT GTA TGG GGA CCG GAT TCT AGA	672
	Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg	
	210 215 220	
30	GAT TGG ATA AGA TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	Asp Trp Ile Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val	
	225 230 235 240	
35	TTA GAT ATC GTT TCT CTA TTT CCG AAC TAT GAT AGT AGA ACG TAT CCA	768
	Leu Asp Ile Val Ser Leu Phe Pro Asn Tyr Asp Ser Arg Thr Tyr Pro	
	245 250 255	
40	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	
	260 265 270	
45	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	
	275 280 285	
50	GGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	Gly Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	
	290 295 300	
55	ATC TAT ACG GAT GCT CAT AGA GGA GAA TAT TAT TGG TCA GGG CAT CAA	960
	Ile Tyr Thr Asp Ala His Arg Gly Glu Tyr Tyr Trp Ser Gly His Gln	
	305 310 315 320	
60	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	
	325 330 335	
65	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
	Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala	
	340 345 350	
70	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg	
	355 360 365	
75	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152

	Arg	Pro	Phe	Asn	Ile	Gly	Ile	Asn	Asn	Gln	Gln	Leu	Ser	Val	Leu	Asp	
	370						375					380					
5	GGG	ACA	GAA	TTT	GCT	TAT	GGA	ACC	TCC	TCA	AAT	TTG	CCA	TCC	GCT	GTA	1200
	Gly	Thr	Glu	Phe	Ala	Tyr	Gly	Thr	Ser	Ser	Asn	Leu	Pro	Ser	Ala	Val	
	385					390					395					400	
10	TAC	AGA	AAA	AGC	GGA	ACG	GTA	GAT	TCG	CTG	GAT	GAA	ATA	CCG	CCA	CAG	1248
	Tyr	Arg	Lys	Ser	Gly	Thr	Val	Asp	Ser	Leu	Asp	Glu	Ile	Pro	Pro	Gln	
					405					410					415		
15	AAT	AAC	AAC	GTG	CCA	CCT	AGG	CAA	GGA	TTT	AGT	CAT	CGA	TTA	AGC	CAT	1296
	Asn	Asn	Asn	Val	Pro	Pro	Arg	Gln	Gly	Phe	Ser	His	Arg	Leu	Ser	His	
				420					425					430			
20	GTT	TCA	ATG	TTT	CGT	TCA	GGC	TTT	AGT	AAT	AGT	AGT	GTA	AGT	ATA	ATA	1344
	Val	Ser	Met	Phe	Arg	Ser	Gly	Phe	Ser	Asn	Ser	Ser	Val	Ser	Ile	Ile	
			435					440					445				
25	AGA	GCT	CCA	ATG	TTT	TCT	TGG	ACG	CAC	CGT	AGT	GCA	ACC	CCT	ACA	AAT	1392
	Arg	Ala	Pro	Met	Phe	Ser	Trp	Thr	His	Arg	Ser	Ala	Thr	Pro	Thr	Asn	
		450					455					460					
30	ACA	ATT	GAT	CCG	GAG	AGG	ATT	ACT	CAA	ATA	CCA	TTG	GTA	AAA	GCA	CAT	1440
	Thr	Ile	Asp	Pro	Glu	Arg	Ile	Thr	Gln	Ile	Pro	Leu	Val	Lys	Ala	His	
	465					470				475						480	
35	ACA	CTT	CAG	TCA	GGT	ACT	ACT	GTT	GTA	AGA	GGG	CCC	GGG	TTT	ACG	GGA	1488
	Thr	Leu	Gln	Ser	Gly	Thr	Thr	Val	Val	Arg	Gly	Pro	Gly	Phe	Thr	Gly	
					485					490					495		
40	GGA	GAT	ATT	CTT	CGA	CGA	ACA	AGT	GGA	GGA	CCA	TTT	GCT	TAT	ACT	ATT	1536
	Gly	Asp	Ile	Leu	Arg	Arg	Thr	Ser	Gly	Gly	Pro	Phe	Ala	Tyr	Thr	Ile	
				500					505					510			
45	GTT	AAT	ATA	AAT	GGG	CAA	TTA	CCC	CAA	AGG	TAT	CGT	GCA	AGA	ATA	CGC	1584
	Val	Asn	Ile	Asn	Gly	Gln	Leu	Pro	Gln	Arg	Tyr	Arg	Ala	Arg	Ile	Arg	
			515				520						525				
50	TAT	GCC	TCT	ACT	ACA	AAT	CTA	AGA	ATT	TAC	GTA	ACG	GTT	GCA	GGT	GAA	1632
	Tyr	Ala	Ser	Thr	Thr	Asn	Leu	Arg	Ile	Tyr	Val	Val	Val	Ala	Gly	Glu	
		530					535					540					
55	CGG	ATT	TTT	GCT	GGT	CAA	TTT	AAC	AAA	ACA	ATG	GAT	ACC	GGT	GAC	CCA	1680
	Arg	Ile	Phe	Ala	Gly	Gln	Phe	Asn	Lys	Thr	Met	Asp	Thr	Gly	Asp	Pro	
	545					550					555					560	
60	TTA	ACA	TTC	CAA	TCT	TTT	AGT	TAC	GCA	ACT	ATT	AAT	ACA	GCT	TTT	ACA	1728
	Leu	Thr	Phe	Gln	Ser	Phe	Ser	Tyr	Ala	Thr	Ile	Asn	Thr	Ala	Phe	Thr	
					565					570					575		
65	TTC	CCA	ATG	AGC	CAG	AGT	AGT	TTC	ACA	GTA	GGT	GCT	GAT	ACT	TTT	AGT	1776
	Phe	Pro	Met	Ser	Gln	Ser	Ser	Phe	Thr	Val	Gly	Ala	Asp	Thr	Phe	Ser	
				580					585					590			
70	TCA	GGG	AAT	GAA	GTT	TAT	ATA	GAC	AGA	TTT	GAA	TTG	ATT	CCA	GTT	ACT	1824
	Ser	Gly	Asn	Glu	Val	Tyr	Ile	Asp	Arg	Phe	Glu	Leu	Ile	Pro	Val	Thr	
			595					600					605				
75	GCA	ACA	TTT	GAA	GCA	GAA	TAT	GAT	TTA	GAA	AGA	GCA	CAA	AAG	GCG	GTG	1872
	Ala	Thr	Phe	Glu	Ala	Glu	Tyr	Asp	Leu	Glu	Arg	Ala	Gln	Lys	Ala	Val	

	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys	
	865 870 875 880	
5	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu	
	885 890 895	
10	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe	
	900 905 910	
15	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met	
	915 920 925	
20	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu	
	930 935 940	
25	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu	
	945 950 955 960	
30	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn	
	965 970 975	
35	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val	
	980 985 990	
40	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
	Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu	
	995 1000 1005	
45	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys	
	1010 1015 1020	
50	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr	
	1025 1030 1035 1040	
55	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
	Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu	
	1045 1050 1055	
60	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
	Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr	
	1060 1065 1070	
65	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
	Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala	
	1075 1080 1085	
70	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312
	Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala	
	1090 1095 1100	

GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA 3360
 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg 1120
 1105 1110 1115
 5 GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA 3408
 Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu 1135
 1125 1130
 10 CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT 3456
 Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp 1150
 1140 1145
 15 AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC 3504
 Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp 1165
 1155 1160
 20 AGC GTG GAA TTA CTC CTT ATG GAG GAA 3531
 Ser Val Glu Leu Leu Leu Met Glu Glu 1175
 1170

(2) INFORMATION FOR SEQ ID NO:14:

25 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1177 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

30 (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
 1 5 10 15
 35 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
 20 25 30
 40 Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
 35 40 45
 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
 50 55 60
 45 Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
 65 70 75 80
 Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
 85 90 95
 50 Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
 100 105 110
 55 Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu
 115 120 125
 Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala
 130 135 140
 60 Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val
 145 150 155 160

Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser
 165 170 175
 5 Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg
 180 185 190
 Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp His Ala Val
 195 200 205
 10 Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg
 210 215 220
 Asp Trp Ile Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val
 15 225 230 235 240
 Leu Asp Ile Val Ser Leu Phe Pro Asn Tyr Asp Ser Arg Thr Tyr Pro
 245 250 255
 20 Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val
 260 265 270
 Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu
 275 280 285
 25 Gly Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr
 290 295 300
 Ile Tyr Thr Asp Ala His Arg Gly Glu Tyr Tyr Trp Ser Gly His Gln
 305 310 315 320
 30 Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro
 325 330 335
 35 Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala
 340 345 350
 Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg
 355 360 365
 40 Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp
 370 375 380
 Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val
 385 390 395 400
 45 Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln
 405 410 415
 50 Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His
 420 425 430
 Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile
 435 440 445
 55 Arg Ala Pro Met Phe Ser Trp Thr His Arg Ser Ala Thr Pro Thr Asn
 450 455 460
 60 Thr Ile Asp Pro Glu Arg Ile Thr Gln Ile Pro Leu Val Lys Ala His
 465 470 475 480

Thr Leu Gln Ser Gly Thr Thr Val Val Arg Gly Pro Gly Phe Thr Gly
 485 490 495
 5 Gly Asp Ile Leu Arg Arg Thr Ser Gly Gly Pro Phe Ala Tyr Thr Ile
 500 505 510
 Val Asn Ile Asn Gly Gln Leu Pro Gln Arg Tyr Arg Ala Arg Ile Arg
 515 520 525
 10 Tyr Ala Ser Thr Thr Asn Leu Arg Ile Tyr Val Thr Val Ala Gly Glu
 530 535 540
 Arg Ile Phe Ala Gly Gln Phe Asn Lys Thr Met Asp Thr Gly Asp Pro
 545 550 555 560
 15 Leu Thr Phe Gln Ser Phe Ser Tyr Ala Thr Ile Asn Thr Ala Phe Thr
 565 570 575
 Phe Pro Met Ser Gln Ser Ser Phe Thr Val Gly Ala Asp Thr Phe Ser
 580 585 590
 20 Ser Gly Asn Glu Val Tyr Ile Asp Arg Phe Glu Leu Ile Pro Val Thr
 595 600 605
 25 Ala Thr Phe Glu Ala Glu Tyr Asp Leu Glu Arg Ala Gln Lys Ala Val
 610 615 620
 Asn Ala Leu Phe Thr Ser Ile Asn Gln Ile Gly Ile Lys Thr Asp Val
 625 630 635 640
 30 Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Asp Cys Leu Ser
 645 650 655
 Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys
 660 665 670
 35 His Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Pro Asn
 675 680 685
 40 Phe Lys Gly Ile Asn Arg Gln Leu Asp Arg Gly Trp Arg Gly Ser Thr
 690 695 700
 Asp Ile Thr Ile Gln Arg Gly Asp Asp Val Phe Lys Glu Asn Tyr Val
 705 710 715 720
 45 Thr Leu Pro Gly Thr Phe Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln
 725 730 735
 Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg
 740 745 750
 50 Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr
 755 760 765
 Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp
 770 775 780
 55 Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg
 785 790 795 800
 60 Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg

					805					810					815	
	Asp	Gly	Glu	Lys	Cys	Ala	His	His	Ser	His	His	Phe	Ser	Leu	Asp	Ile
				820					825					830		
5	Asp	Val	Gly	Cys	Thr	Asp	Leu	Asn	Glu	Asp	Leu	Gly	Val	Trp	Val	Ile
			835					840					845			
	Phe	Lys	Ile	Lys	Thr	Gln	Asp	Gly	His	Ala	Arg	Leu	Gly	Asn	Leu	Glu
10		850					855					860				
	Phe	Leu	Glu	Glu	Lys	Pro	Leu	Val	Gly	Glu	Ala	Leu	Ala	Arg	Val	Lys
	865					870					875					880
15	Arg	Ala	Glu	Lys	Lys	Trp	Arg	Asp	Lys	Arg	Glu	Lys	Leu	Glu	Trp	Glu
					885					890					895	
	Thr	Asn	Ile	Val	Tyr	Lys	Glu	Ala	Lys	Glu	Ser	Val	Asp	Ala	Leu	Phe
				900					905					910		
20	Val	Asn	Ser	Gln	Tyr	Asp	Gln	Leu	Gln	Ala	Asp	Thr	Asn	Ile	Ala	Met
			915					920					925			
	Ile	His	Ala	Ala	Asp	Lys	Arg	Val	His	Ser	Ile	Arg	Glu	Ala	Tyr	Leu
25		930					935					940				
	Pro	Glu	Leu	Ser	Val	Ile	Pro	Gly	Val	Asn	Ala	Ala	Ile	Phe	Glu	Glu
	945					950					955					960
30	Leu	Glu	Gly	Arg	Ile	Phe	Thr	Ala	Phe	Ser	Leu	Tyr	Asp	Ala	Arg	Asn
					965					970					975	
	Val	Ile	Lys	Asn	Gly	Asp	Phe	Asn	Asn	Gly	Leu	Ser	Cys	Trp	Asn	Val
				980					985					990		
35	Lys	Gly	His	Val	Asp	Val	Glu	Glu	Gln	Asn	Asn	Gln	Arg	Ser	Val	Leu
			995					1000					1005			
	Val	Val	Pro	Glu	Trp	Glu	Ala	Glu	Val	Ser	Gln	Glu	Val	Arg	Val	Cys
40		1010					1015					1020				
	Pro	Gly	Arg	Gly	Tyr	Ile	Leu	Arg	Val	Thr	Ala	Tyr	Lys	Glu	Gly	Tyr
	1025					1030					1035					1040
45	Gly	Glu	Gly	Cys	Val	Thr	Ile	His	Glu	Ile	Glu	Asn	Asn	Thr	Asp	Glu
					1045					1050					1055	
	Leu	Lys	Phe	Ser	Asn	Cys	Val	Glu	Glu	Glu	Ile	Tyr	Pro	Asn	Asn	Thr
					1060				1065					1070		
50	Val	Thr	Cys	Asn	Asp	Tyr	Thr	Val	Asn	Gln	Glu	Glu	Tyr	Gly	Gly	Ala
					1075				1080					1085		
	Tyr	Thr	Ser	Arg	Asn	Arg	Gly	Tyr	Asn	Glu	Ala	Pro	Ser	Val	Pro	Ala
55		1090					1095					1100				
	Asp	Tyr	Ala	Ser	Val	Tyr	Glu	Glu	Lys	Ser	Tyr	Thr	Asp	Gly	Arg	Arg
	1105					1110					1115					1120
60	Glu	Asn	Pro	Cys	Glu	Phe	Asn	Arg	Gly	Tyr	Arg	Asp	Tyr	Thr	Pro	Leu
					1125					1130					1135	

Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp
 1140 1145 1150

5 Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
 1155 1160 1165

Ser Val Glu Leu Leu Leu Met Glu Glu
 1170 1175

10

(2) INFORMATION FOR SEQ ID NO:15:

15 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

TATCCAATTC GAACGTCATC

20

25 (2) INFORMATION FOR SEQ ID NO:16:

30 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

35 TTTAGTCATC GATTAAATCA

20

(2) INFORMATION FOR SEQ ID NO:17:

40 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

ATAATAAGAG CTCCAATGTT

20

50

(2) INFORMATION FOR SEQ ID NO:18:

55 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

60

TACATCGTAG TGCAACTCTT

20

(2) INFORMATION FOR SEQ ID NO:19:

5 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
10 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

TCATGGAGAG CTCCTATGTT

20

15 (2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
20 (B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

25 TTAACAAGAG CTCCTATGTT

20

(2) INFORMATION FOR SEQ ID NO:21:

30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
35 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

40 ACTACCAGGT ACCTTTGATG

20

(2) INFORMATION FOR SEQ ID NO:22:

45 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

ACTACCGGT ACCTTTGATA

20

55 (2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18 base pairs
60 (B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

ATTTGAGTAA TACTATCC

18

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

ATTACTCAAA TACCATTGG

19

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 3534 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 1..3531

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA
Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
1 5 10 15

48

AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT
Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
20 25 30

96

TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT
Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
35 40 45

144

GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA
Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
50 55 60

192

TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT
Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
65 70 75 80

240

GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC
Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
85 90 95

288

ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA
Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
100 105 110

336

	TCT	TTT	AGA	GAG	TGG	GAA	GCA	GAT	CCT	ACT	AAT	CCA	GCA	TTA	AGA	GAA	384
	Ser	Phe	Arg	Glu	Trp	Glu	Ala	Asp	Pro	Thr	Asn	Pro	Ala	Leu	Arg	Glu	
			115					120					125				
5	GAG	ATG	CGT	ATT	CAA	TTC	AAT	GAC	ATG	AAC	AGT	GCC	CTT	ACA	ACC	GCT	432
	Glu	Met	Arg	Ile	Gln	Phe	Asn	Asp	Met	Asn	Ser	Ala	Leu	Thr	Thr	Ala	
		130					135					140					
10	ATT	CCT	CTT	TTT	GCA	GTT	CAA	AAT	TAT	CAA	GTT	CCT	CTT	TTA	TCA	GTA	480
	Ile	Pro	Leu	Phe	Ala	Val	Gln	Asn	Tyr	Gln	Val	Pro	Leu	Leu	Ser	Val	
		145				150					155					160	
15	TAT	GTT	CAA	GCT	GCA	AAT	TTA	CAT	TTA	TCA	GTT	TTG	AGA	GAT	GTT	TCA	528
	Tyr	Val	Gln	Ala	Ala	Asn	Leu	His	Leu	Ser	Val	Leu	Arg	Asp	Val	Ser	
				165						170					175		
20	GTG	TTT	GGA	CAA	AGG	TGG	GGA	TTT	GAT	GCC	GCG	ACT	ATC	AAT	AGT	CGT	576
	Val	Phe	Gly	Gln	Arg	Trp	Gly	Phe	Asp	Ala	Ala	Thr	Ile	Asn	Ser	Arg	
			180						185					190			
25	TAT	AAT	GAT	TTA	ACT	AGG	CTT	ATT	GGC	AAC	TAT	ACA	GAT	CAT	GCT	GTA	624
	Tyr	Asn	Asp	Leu	Thr	Arg	Leu	Ile	Gly	Asn	Tyr	Thr	Asp	His	Ala	Val	
			195					200					205				
30	CGC	TGG	TAC	AAT	ACG	GGA	TTA	GAG	CGT	GTA	TGG	GGA	CCG	GAT	TCT	AGA	672
	Arg	Trp	Tyr	Asn	Thr	Gly	Leu	Glu	Arg	Val	Trp	Gly	Pro	Asp	Ser	Arg	
		210					215					220					
35	GAT	TGG	ATA	AGA	TAT	AAT	CAA	TTT	AGA	AGA	GAA	TTA	ACA	CTA	ACT	GTA	720
	Asp	Trp	Ile	Arg	Tyr	Asn	Gln	Phe	Arg	Arg	Glu	Leu	Thr	Leu	Thr	Val	
		225				230					235					240	
40	TTA	GAT	ATC	GTT	TCT	CTA	TTT	CCG	AAC	TAT	GAT	AGT	AGA	ACG	TAT	CCA	768
	Leu	Asp	Ile	Val	Ser	Leu	Phe	Pro	Asn	Tyr	Asp	Ser	Arg	Thr	Tyr	Pro	
				245					250						255		
45	ATT	CGA	ACA	GTT	TCC	CAA	TTA	ACA	AGA	GAA	ATT	TAT	ACA	AAC	CCA	GTA	816
	Ile	Arg	Thr	Val	Ser	Gln	Leu	Thr	Arg	Glu	Ile	Tyr	Thr	Asn	Pro	Val	
			260						265					270			
50	TTA	GAA	AAT	TTT	GAT	GGT	AGT	TTT	CGA	GGC	TCG	GCT	CAG	GGC	ATA	GAA	864
	Leu	Glu	Asn	Phe	Asp	Gly	Ser	Phe	Arg	Gly	Ser	Ala	Gln	Gly	Ile	Glu	
			275					280					285				
55	AGA	AGT	ATT	AGG	AGT	CCA	CAT	TTG	ATG	GAT	ATA	CTT	AAC	AGT	ATA	ACC	912
	Arg	Ser	Ile	Arg	Ser	Pro	His	Leu	Met	Asp	Ile	Leu	Asn	Ser	Ile	Thr	
		290					295					300					
60	ATC	TAT	ACG	GAT	GCT	CAT	AGG	GGT	TAT	TAT	TAT	TGG	TCA	GGG	CAT	CAA	960
	Ile	Tyr	Thr	Asp	Ala	His	Arg	Gly	Tyr	Tyr	Tyr	Trp	Ser	Gly	His	Gln	
		305				310					315					320	
65	ATA	ATG	GCT	TCT	CCT	GTA	GGG	TTT	TCG	GGG	CCA	GAA	TTC	ACT	TTT	CCG	1008
	Ile	Met	Ala	Ser	Pro	Val	Gly	Phe	Ser	Gly	Pro	Glu	Phe	Thr	Phe	Pro	
				325					330						335		
70	CTA	TAT	GGA	ACT	ATG	GGA	AAT	GCA	GCT	CCA	CAA	CAA	CGT	ATT	GTT	GCT	1056
	Leu	Tyr	Gly	Thr	Met	Gly	Asn	Ala	Ala	Pro	Gln	Gln	Arg	Ile	Val	Ala	
				340				345						350			
75	CAA	CTA	GGT	CAG	GGC	GTG	TAT	AGA	ACA	TTA	TCG	TCC	ACT	TTA	TAT	AGA	1104

	Gln	Leu	Gly	Gln	Gly	Val	Tyr	Arg	Thr	Leu	Ser	Ser	Thr	Leu	Tyr	Arg	
			355					360					365				
5	AGA	CCT	TTT	AAT	ATA	GGG	ATA	AAT	AAT	CAA	CAA	CTA	TCT	GTT	CTT	GAC	1152
	Arg	Pro	Phe	Asn	Ile	Gly	Ile	Asn	Asn	Gln	Gln	Leu	Ser	Val	Leu	Asp	
			370				375					380					
10	GGG	ACA	GAA	TTT	GCT	TAT	GGA	ACC	TCC	TCA	AAT	TTG	CCA	TCC	GCT	GTA	1200
	Gly	Thr	Glu	Phe	Ala	Tyr	Gly	Thr	Ser	Ser	Asn	Leu	Pro	Ser	Ala	Val	
						390					395					400	
15	TAC	AGA	AAA	AGC	GGA	ACG	GTA	GAT	TCG	CTG	GAT	GAA	ATA	CCG	CCA	CAG	1248
	Tyr	Arg	Lys	Ser	Gly	Thr	Val	Asp	Ser	Leu	Asp	Glu	Ile	Pro	Pro	Gln	
					405					410					415		
20	AAT	AAC	AAC	GTG	CCA	CCT	AGG	CAA	GGA	TTT	AGT	CAT	CGA	TTA	AGC	CAT	1296
	Asn	Asn	Asn	Val	Pro	Pro	Arg	Gln	Gly	Phe	Ser	His	Arg	Leu	Ser	His	
				420					425					430			
25	GTT	TCA	ATG	TTT	CGT	TCA	GGC	TTT	AGT	AAT	AGT	AGT	GTA	AGT	ATA	ATA	1344
	Val	Ser	Met	Phe	Arg	Ser	Gly	Phe	Ser	Asn	Ser	Ser	Val	Ser	Ile	Ile	
			435				440						445				
30	AGA	GCT	CCA	ATG	TTT	TCT	TGG	ACG	CAC	CGT	AGT	GCA	ACC	CCT	ACA	AAT	1392
	Arg	Ala	Pro	Met	Phe	Ser	Trp	Thr	His	Arg	Ser	Ala	Thr	Pro	Thr	Asn	
		450					455					460					
35	ACA	ATT	GAT	CCG	GAG	AGG	ATT	ACT	CAA	ATA	CCA	TTG	GTA	AAA	GCA	CAT	1440
	Thr	Ile	Asp	Pro	Glu	Arg	Ile	Thr	Gln	Ile	Pro	Leu	Val	Lys	Ala	His	
		465				470					475					480	
40	ACA	CTT	CAG	TCA	GGT	ACT	ACT	GTT	GTA	AGA	GGG	CCC	GGG	TTT	ACG	GGA	1488
	Thr	Leu	Gln	Ser	Gly	Thr	Thr	Val	Val	Arg	Gly	Pro	Gly	Phe	Thr	Gly	
					485				490						495		
45	GGA	GAT	ATT	CTT	CGA	CGA	ACA	AGT	GGA	GGA	CCA	TTT	GCT	TAT	ACT	ATT	1536
	Gly	Asp	Ile	Leu	Arg	Arg	Thr	Ser	Gly	Gly	Pro	Phe	Ala	Tyr	Thr	Ile	
				500					505					510			
50	GTT	AAT	ATA	AAT	GGG	CAA	TTA	CCC	CAA	AGG	TAT	CGT	GCA	AGA	ATA	CGC	1584
	Val	Asn	Ile	Asn	Gly	Gln	Leu	Pro	Gln	Arg	Tyr	Arg	Ala	Arg	Ile	Arg	
			515				520						525				
55	TAT	GCC	TCT	ACT	ACA	AAT	CTA	AGA	ATT	TAC	GTA	ACG	GTT	GCA	GGT	GAA	1632
	Tyr	Ala	Ser	Thr	Thr	Asn	Leu	Arg	Ile	Tyr	Val	Thr	Val	Ala	Gly	Glu	
		530					535					540					
60	CGG	ATT	TTT	GCT	GGT	CAA	TTT	AAC	AAA	ACA	ATG	GAT	ACC	GGT	GAC	CCA	1680
	Arg	Ile	Phe	Ala	Gly	Gln	Phe	Asn	Lys	Thr	Met	Asp	Thr	Gly	Asp	Pro	
		545				550					555					560	
65	TTA	ACA	TTC	CAA	TCT	TTT	AGT	TAC	GCA	ACT	ATT	AAT	ACA	GCT	TTT	ACA	1728
	Leu	Thr	Phe	Gln	Ser	Phe	Ser	Tyr	Ala	Thr	Ile	Asn	Thr	Ala	Phe	Thr	
					565				570						575		
70	TTC	CCA	ATG	AGC	CAG	AGT	AGT	TTC	ACA	GTA	GGT	GCT	GAT	ACT	TTT	AGT	1776
	Phe	Pro	Met	Ser	Gln	Ser	Ser	Phe	Thr	Val	Gly	Ala	Asp	Thr	Phe	Ser	
				580					585					590			
75	TCA	GGG	AAT	GAA	GTT	TAT	ATA	GAC	AGA	TTT	GAA	TTG	ATT	CCA	GTT	ACT	1824
	Ser	Gly	Asn	Glu	Val	Tyr	Ile	Asp	Arg	Phe	Glu	Leu	Ile	Pro	Val	Thr	

	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu	
	850 855 860	
5	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys	
	865 870 875 880	
10	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu	
	885 890 895	
15	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe	
	900 905 910	
20	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met	
	915 920 925	
	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu	
	930 935 940	
25	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu	
	945 950 955 960	
30	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn	
	965 970 975	
35	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val	
	980 985 990	
40	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
	Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu	
	995 1000 1005	
	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys	
	1010 1015 1020	
45	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr	
	1025 1030 1035 1040	
50	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
	Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu	
	1045 1050 1055	
55	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
	Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr	
	1060 1065 1070	
60	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
	Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala	
	1075 1080 1085	

TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT 3312
 Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala
 1090 1095 1100

5 GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA 3360
 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg
 1105 1110 1115 1120

10 GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA 3408
 Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu
 1125 1130 1135

15 CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT 3456
 Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp
 1140 1145 1150

AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC 3504
 Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
 1155 1160 1165

20 AGC GTG GAA TTA CTC CTT ATG GAG GAA TAG 3534
 Ser Val Glu Leu Leu Leu Met Glu Glu
 1170 1175

25 (2) INFORMATION FOR SEQ ID NO:26:

30 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1177 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
 1 5 10 15

40 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
 20 25 30

Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
 35 40 45

45 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
 50 55 60

50 Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
 65 70 75 80

Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
 85 90 95

55 Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
 100 105 110

Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu
 115 120 125

60 Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala

130 135 140
 Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val
 145 150 155 160
 5 Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser
 165 170 175
 10 Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg
 180 185 190
 Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp His Ala Val
 195 200 205
 15 Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg
 210 215 220
 Asp Trp Ile Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val
 225 230 235 240
 20 Leu Asp Ile Val Ser Leu Phe Pro Asn Tyr Asp Ser Arg Thr Tyr Pro
 245 250 255
 25 Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val
 260 265 270
 Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu
 275 280 285
 30 Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr
 290 295 300
 Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln
 305 310 315 320
 35 Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro
 325 330 335
 40 Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala
 340 345 350
 Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg
 355 360 365
 45 Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp
 370 375 380
 Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val
 385 390 395 400
 50 Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln
 405 410 415
 55 Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His
 420 425 430
 Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile
 435 440 445
 60 Arg Ala Pro Met Phe Ser Trp Thr His Arg Ser Ala Thr Pro Thr Asn
 450 455 460

	Thr	Ile	Asp	Pro	Glu	Arg	Ile	Thr	Gln	Ile	Pro	Leu	Val	Lys	Ala	His	465	470	475	480
5	Thr	Leu	Gln	Ser	Gly	Thr	Thr	Val	Val	Arg	Gly	Pro	Gly	Phe	Thr	Gly	485	490	495	
	Gly	Asp	Ile	Leu	Arg	Arg	Thr	Ser	Gly	Gly	Pro	Phe	Ala	Tyr	Thr	Ile	500	505	510	
10	Val	Asn	Ile	Asn	Gly	Gln	Leu	Pro	Gln	Arg	Tyr	Arg	Ala	Arg	Ile	Arg	515	520	525	
15	Tyr	Ala	Ser	Thr	Thr	Asn	Leu	Arg	Ile	Tyr	Val	Thr	Val	Ala	Gly	Glu	530	535	540	
	Arg	Ile	Phe	Ala	Gly	Gln	Phe	Asn	Lys	Thr	Met	Asp	Thr	Gly	Asp	Pro	545	550	555	560
20	Leu	Thr	Phe	Gln	Ser	Phe	Ser	Tyr	Ala	Thr	Ile	Asn	Thr	Ala	Phe	Thr	565	570	575	
	Phe	Pro	Met	Ser	Gln	Ser	Ser	Phe	Thr	Val	Gly	Ala	Asp	Thr	Phe	Ser	580	585	590	
25	Ser	Gly	Asn	Glu	Val	Tyr	Ile	Asp	Arg	Phe	Glu	Leu	Ile	Pro	Val	Thr	595	600	605	
30	Ala	Thr	Phe	Glu	Ala	Glu	Tyr	Asp	Leu	Glu	Arg	Ala	Gln	Lys	Ala	Val	610	615	620	
	Asn	Ala	Leu	Phe	Thr	Ser	Ile	Asn	Gln	Ile	Gly	Ile	Lys	Thr	Asp	Val	625	630	635	640
35	Thr	Asp	Tyr	His	Ile	Asp	Gln	Val	Ser	Asn	Leu	Val	Asp	Cys	Leu	Ser	645	650	655	
	Asp	Glu	Phe	Cys	Leu	Asp	Glu	Lys	Arg	Glu	Leu	Ser	Glu	Lys	Val	Lys	660	665	670	
40	His	Ala	Lys	Arg	Leu	Ser	Asp	Glu	Arg	Asn	Leu	Leu	Gln	Asp	Pro	Asn	675	680	685	
45	Phe	Lys	Gly	Ile	Asn	Arg	Gln	Leu	Asp	Arg	Gly	Trp	Arg	Gly	Ser	Thr	690	695	700	
	Asp	Ile	Thr	Ile	Gln	Arg	Gly	Asp	Asp	Val	Phe	Lys	Glu	Asn	Tyr	Val	705	710	715	720
50	Thr	Leu	Pro	Gly	Thr	Phe	Asp	Glu	Cys	Tyr	Pro	Thr	Tyr	Leu	Tyr	Gln	725	730	735	
	Lys	Ile	Asp	Glu	Ser	Lys	Leu	Lys	Ala	Phe	Thr	Arg	Tyr	Gln	Leu	Arg	740	745	750	
55	Gly	Tyr	Ile	Glu	Asp	Ser	Gln	Asp	Leu	Glu	Ile	Tyr	Leu	Ile	Arg	Tyr	755	760	765	
60	Asn	Ala	Lys	His	Glu	Thr	Val	Asn	Val	Pro	Gly	Thr	Gly	Ser	Leu	Trp	770	775	780	

Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg
 785 790 795 800
 5 Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg
 805 810 815
 Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu Asp Ile
 820 825 830
 10 Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile
 835 840 845
 Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu
 850 855 860
 15 Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys
 865 870 875 880
 Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu
 885 890 895
 20 Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe
 900 905 910
 Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met
 915 920 925
 Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu
 930 935 940
 30 Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu
 945 950 955 960
 Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn
 965 970 975
 35 Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val
 980 985 990
 Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu
 995 1000 1005
 Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys
 1010 1015 1020
 45 Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr
 1025 1030 1035 1040
 Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu
 1045 1050 1055
 50 Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr
 1060 1065 1070
 Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala
 1075 1080 1085
 Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala
 1090 1095 1100
 60 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg

	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala 130 135 140	432
5	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val 145 150 155 160	480
10	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser 165 170 175	528
15	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg 180 185 190	576
20	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val 195 200 205	624
25	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg 210 215 220	672
30	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val 225 230 235 240	720
35	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro 245 250 255	768
40	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val 260 265 270	816
45	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu 275 280 285	864
50	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr 290 295 300	912
55	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln 305 310 315 320	960
60	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro 325 330 335	1008
65	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala 340 345 350	1056
70	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg 340 345 350	1104

	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp	
	370 375 380	
5	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val	
	385 390 395 400	
10	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln	
	405 410 415	
15	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
	Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His	
	420 425 430	
20	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile	
	435 440 445	
25	AGA GCT CCT ATG TTC TCT TGG ATA CAT CGT AGT GCT GAA TTT AAT AAT	1392
	Arg Ala Pro Met Phe Ser Trp Ile His Arg Ser Ala Glu Phe Asn Asn	
	450 455 460	
30	ATA ATT GCA TCG GAT AGT ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	Ile Ile Ala Ser Asp Ser Ile Thr Gln Ile Pro Leu Val Lys Ala His	
	465 470 475 480	
35	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	Thr Leu Gln Ser Gly Thr Thr Val Val Arg Gly Pro Gly Phe Thr Gly	
	485 490 495	
40	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
	Gly Asp Ile Leu Arg Arg Thr Ser Gly Gly Pro Phe Ala Tyr Thr Ile	
	500 505 510	
45	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	Val Asn Ile Asn Gly Gln Leu Pro Gln Arg Tyr Arg Ala Arg Ile Arg	
	515 520 525	
50	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	Tyr Ala Ser Thr Thr Asn Leu Arg Ile Tyr Val Thr Val Ala Gly Glu	
	530 535 540	
55	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	Arg Ile Phe Ala Gly Gln Phe Asn Lys Thr Met Asp Thr Gly Asp Pro	
	545 550 555 560	
60	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	Leu Thr Phe Gln Ser Phe Ser Tyr Ala Thr Ile Asn Thr Ala Phe Thr	
	565 570 575	
65	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
	Phe Pro Met Ser Gln Ser Ser Phe Thr Val Gly Ala Asp Thr Phe Ser	
	580 585 590	
70	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	Ser Gly Asn Glu Val Tyr Ile Asp Arg Phe Glu Leu Ile Pro Val Thr	
	595 600 605	

	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG Ala Thr Phe Glu Ala Glu Tyr Asp Leu Glu Arg Ala Gln Lys Ala Val 610 615 620	1872
5	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG Asn Ala Leu Phe Thr Ser Ile Asn Gln Ile Gly Ile Lys Thr Asp Val 625 630 635 640	1920
10	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Asp Cys Leu Ser 645 650 655	1968
15	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys 660 665 670	2016
	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC His Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Pro Asn 675 680 685	2064
20	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG Phe Lys Gly Ile Asn Arg Gln Leu Asp Arg Gly Trp Arg Gly Ser Thr 690 695 700	2112
25	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC Asp Ile Thr Ile Gln Arg Gly Asp Asp Val Phe Lys Glu Asn Tyr Val 705 710 715 720	2160
30	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA Thr Leu Pro Gly Thr Phe Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln 725 730 735	2208
35	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg 740 745 750	2256
	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr 755 760 765	2304
40	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp 770 775 780	2352
45	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg 785 790 795 800	2400
50	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg 805 810 815	2448
55	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu Asp Ile 820 825 830	2496
	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile 835 840 845	2544
60	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592

	Phe	Lys	Ile	Lys	Thr	Gln	Asp	Gly	His	Ala	Arg	Leu	Gly	Asn	Leu	Glu	
	850						855					860					
5	TTT	CTC	GAA	GAG	AAA	CCA	TTA	GTA	GGA	GAA	GCG	CTA	GCT	CGT	GTG	AAA	2640
	Phe	Leu	Glu	Glu	Lys	Pro	Leu	Val	Gly	Glu	Ala	Leu	Ala	Arg	Val	Lys	
	865					870				875					880		
10	AGA	GCG	GAG	AAA	AAA	TGG	AGA	GAC	AAA	CGT	GAA	AAA	TTG	GAA	TGG	GAA	2688
	Arg	Ala	Glu	Lys	Lys	Trp	Arg	Asp	Lys	Arg	Glu	Lys	Leu	Glu	Trp	Glu	
				885						890					895		
15	ACA	AAT	ATC	GTT	TAT	AAA	GAG	GCA	AAA	GAA	TCT	GTA	GAT	GCT	TTA	TTT	2736
	Thr	Asn	Ile	Val	Tyr	Lys	Glu	Ala	Lys	Glu	Ser	Val	Asp	Ala	Leu	Phe	
				900					905					910			
20	GTA	AAC	TCT	CAA	TAT	GAT	CAA	TTA	CAA	GCG	GAT	ACG	AAT	ATT	GCC	ATG	2784
	Val	Asn	Ser	Gln	Tyr	Asp	Gln	Leu	Gln	Ala	Asp	Thr	Asn	Ile	Ala	Met	
			915					920					925				
25	ATT	CAT	GCG	GCA	GAT	AAA	CGT	GTT	CAT	AGC	ATT	CGA	GAA	GCT	TAT	CTG	2832
	Ile	His	Ala	Ala	Asp	Lys	Arg	Val	His	Ser	Ile	Arg	Glu	Ala	Tyr	Leu	
						930		935				940					
30	CCT	GAG	CTG	TCT	GTG	ATT	CCG	GGT	GTC	AAT	GCG	GCT	ATT	TTT	GAA	GAA	2880
	Pro	Glu	Leu	Ser	Val	Ile	Pro	Gly	Val	Asn	Ala	Ala	Ile	Phe	Glu	Glu	
	945					950				955					960		
35	TTA	GAA	GGG	CGT	ATT	TTC	ACT	GCA	TTC	TCC	CTA	TAT	GAT	GCG	AGA	AAT	2928
	Leu	Glu	Gly	Arg	Ile	Phe	Thr	Ala	Phe	Ser	Leu	Tyr	Asp	Ala	Arg	Asn	
				965						970					975		
40	GTC	ATT	AAA	AAT	GGT	GAT	TTT	AAT	AAT	GGC	TTA	TCC	TGC	TGG	AAC	GTG	2976
	Val	Ile	Lys	Asn	Gly	Asp	Phe	Asn	Asn	Gly	Leu	Ser	Cys	Trp	Asn	Val	
				980				985						990			
45	AAA	GGG	CAT	GTA	GAT	GTA	GAA	GAA	CAA	AAC	AAC	CAA	CGT	TCG	GTC	CTT	3024
	Lys	Gly	His	Val	Asp	Val	Glu	Glu	Gln	Asn	Asn	Gln	Arg	Ser	Val	Leu	
				995				1000					1005				
50	GTT	GTT	CCG	GAA	TGG	GAA	GCA	GAA	GTG	TCA	CAA	GAA	GTT	CGT	GTC	TGT	3072
	Val	Val	Pro	Glu	Trp	Glu	Ala	Glu	Val	Ser	Gln	Glu	Val	Arg	Val	Cys	
				1010			1015					1020					
55	CCG	GGT	CGT	GGC	TAT	ATC	CTT	CGT	GTC	ACA	GCG	TAC	AAG	GAG	GGA	TAT	3120
	Pro	Gly	Arg	Gly	Tyr	Ile	Leu	Arg	Val	Thr	Ala	Tyr	Lys	Glu	Gly	Tyr	
	1025					1030				1035					1040		
60	GGA	GAA	GGT	TGC	GTA	ACC	ATT	CAT	GAG	ATC	GAG	AAC	AAT	ACA	GAC	GAA	3168
	Gly	Glu	Gly	Cys	Val	Thr	Ile	His	Glu	Ile	Glu	Asn	Asn	Thr	Asp	Glu	
				1045					1050						1055		
65	CTG	AAG	TTT	AGC	AAC	TGC	GTA	GAA	GAG	GAA	ATC	TAT	CCA	AAT	AAC	ACG	3216
	Leu	Lys	Phe	Ser	Asn	Cys	Val	Glu	Glu	Ile	Tyr	Pro	Asn	Asn	Thr		
				1060				1065					1070				
70	GTA	ACG	TGT	AAT	GAT	TAT	ACT	GTA	AAT	CAA	GAA	GAA	TAC	GGA	GGT	GCG	3264
	Val	Thr	Cys	Asn	Asp	Tyr	Thr	Val	Asn	Gln	Glu	Glu	Tyr	Gly	Gly	Ala	
				1075				1080					1085				
75	TAC	ACT	TCT	CGT	AAT	CGA	GGA	TAT	AAC	GAA	GCT	CCT	TCC	GTA	CCA	GCT	3312
	Tyr	Thr	Ser	Arg	Asn	Arg	Gly	Tyr	Asn	Glu	Ala	Pro	Ser	Val	Pro	Ala	

	1090	1095	1100	
	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA			3360
5	Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg	1110	1115 1120	
	1105			
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA			3408
	Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu	1125	1130 1135	
10	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT			3456
	Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp	1140	1145 1150	
15	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC			3504
	Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp	1155	1160 1165	
20	AGC GTG GAA TTA CTC CTT ATG GAG GAA TAG			3534
	Ser Val Glu Leu Leu Leu Met Glu Glu	1170	1175	

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1177 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

35	Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu	1 5 10 15
	Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly	20 25 30
40	Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser	35 40 45
45	Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile	50 55 60
	Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile	65 70 75 80
50	Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala	85 90 95
	Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu	100 105 110
55	Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu	115 120 125
60	Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala	130 135 140

Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val
 145 150 155 160
 5 Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser
 165 170 175
 Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg
 180 185 190
 10 Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val
 195 200 205
 Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg
 210 215 220
 15 Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val
 225 230 235 240
 20 Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro
 245 250 255
 Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val
 260 265 270
 25 Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu
 275 280 285
 Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr
 290 295 300
 30 Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln
 305 310 315 320
 35 Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro
 325 330 335
 Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala
 340 345 350
 40 Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg
 355 360 365
 Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp
 370 375 380
 45 Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val
 385 390 395 400
 50 Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln
 405 410 415
 Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His
 420 425 430
 55 Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile
 435 440 445
 Arg Ala Pro Met Phe Ser Trp Ile His Arg Ser Ala Glu Phe Asn Asn
 450 455 460
 60 Ile Ile Ala Ser Asp Ser Ile Thr Gln Ile Pro Leu Val Lys Ala His

Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg
 805 810 815
 5 Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu Asp Ile
 820 825 830
 Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile
 835 840 845
 10 Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu
 850 855 860
 Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys
 865 870 875 880
 15 Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu
 885 890 895
 20 Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe
 900 905 910
 Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met
 915 920 925
 25 Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu
 930 935 940
 Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu
 945 950 955 960
 30 Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn
 965 970 975
 35 Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val
 980 985 990
 Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu
 995 1000 1005
 40 Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys
 1010 1015 1020
 Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr
 1025 1030 1035 1040
 45 Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu
 1045 1050 1055
 50 Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr
 1060 1065 1070
 Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala
 1075 1080 1085
 55 Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala
 1090 1095 1100
 60 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg
 1105 1110 1115 1120

Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu
 1125 1130 1135
 5 Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp
 1140 1145 1150
 Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
 1155 1160 1165
 10 Ser Val Glu Leu Leu Leu Met Glu Glu
 1170 1175

(2) INFORMATION FOR SEQ ID NO:29:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 3579 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 1..3579

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

30	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu 1 5 10 15	48
35	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly 20 25 30	96
40	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser 35 40 45	144
45	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile 50 55 60	192
50	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile 65 70 75 80	240
55	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala 85 90 95	288
60	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu 100 105 110	336
	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu 115 120 125	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432

	Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala	
	130 135 140	
5	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val	480
	145 150 155 160	
10	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser	528
	165 170 175	
15	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg	576
	180 185 190	
20	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val	624
	195 200 205	
25	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg	672
	210 215 220	
30	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val	720
	225 230 235 240	
35	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro	768
	245 250 255	
40	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	816
	260 265 270	
45	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	864
	275 280 285	
50	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	912
	290 295 300	
55	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Trp Ser Gly His Gln	960
	305 310 315 320	
60	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	1008
	325 330 335	
65	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala	1056
	340 345 350	
70	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg	1104
	340 345 350	
75	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp	1152

	370	375	380	
5	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val 385 390 395 400	1200		
10	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln 405 410 415	1248		
15	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His 420 425 430	1296		
20	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile 435 440 445	1344		
25	AGA GCT CCT ATG TTC TCT TGG ATA CAT CGT AGT GCA ACT CTT ACA AAT Arg Ala Pro Met Phe Ser Trp Ile His Arg Ser Ala Thr Leu Thr Asn 450 455 460	1392		
30	ACA ATT GAT CCA GAG AGA ATT AAT CAA ATA CCT TTA GTG AAA GGA TTT Thr Ile Asp Pro Glu Arg Ile Asn Gln Ile Pro Leu Val Lys Gly Phe 465 470 475 480	1440		
35	AGA GTT TGG GGG GGC ACC TCT GTC ATT ACA GGA CCA GGA TTT ACA GGA Arg Val Trp Gly Gly Thr Ser Val Ile Thr Gly Pro Gly Phe Thr Gly 485 490 495	1488		
40	GGG GAT ATC CTT CGA AGA AAT ACC TTT GGT GAT TTT GTA TCT CTA CAA Gly Asp Ile Leu Arg Arg Asn Thr Phe Gly Asp Phe Val Ser Leu Gln 500 505 510	1536		
45	GTC AAT ATT AAT TCA CCA ATT ACC CAA AGA TAC CGT TTA AGA TTT CGT Val Asn Ile Asn Ser Pro Ile Thr Gln Arg Tyr Arg Leu Arg Phe Arg 515 520 525	1584		
50	TAC GCT TCC AGT AGG GAT GCA CGA GTT ATA GTA TTA ACA GGA GCG GCA Tyr Ala Ser Ser Arg Asp Ala Arg Val Ile Val Leu Thr Gly Ala Ala 530 535 540	1632		
55	TCC ACA GGA GTG GGA GGC CAA GTT AGT GTA AAT ATG CCT CTT CAG AAA Ser Thr Gly Val Gly Gly Gln Val Ser Val Asn Met Pro Leu Gln Lys 545 550 555 560	1680		
60	ACT ATG GAA ATA GGG GAG AAC TTA ACA TCT AGA ACA TTT AGA TAT ACC Thr Met Glu Ile Gly Glu Asn Leu Thr Ser Arg Thr Phe Arg Tyr Thr 565 570 575	1728		
65	GAT TTT AGT AAT CCT TTT TCA TTT AGA GCT AAT CCA GAT ATA ATT GGG Asp Phe Ser Asn Pro Phe Ser Phe Arg Ala Asn Pro Asp Ile Ile Gly 580 585 590	1776		
70	ATA AGT GAA CAA CCT CTA TTT GGT GCA GGT TCT ATT AGT AGC GGT GAA Ile Ser Glu Gln Pro Leu Phe Gly Ala Gly Ser Ile Ser Ser Gly Glu 595 600 605	1824		
75	CTT TAT ATA GAT AAA ATT GAA ATT ATT CTA GCA GAT GCA ACA TTT GAA Leu Tyr Ile Asp Lys Ile Glu Ile Ile Leu Ala Asp Ala Thr Phe Glu 610 615 620	1872		

	GCA GAA TCT GAT TTA GAA AGA GCA CAA AAG GCG GTG AAT GCC CTG TTT Ala Glu Ser Asp Leu Glu Arg Ala Gln Lys Ala Val Asn Ala Leu Phe 625 630 635 640	1920
5	ACT TCT TCC AAT CAA ATC GGG TTA AAA ACC GAT GTG ACG GAT TAT CAT Thr Ser Ser Asn Gln Ile Gly Leu Lys Thr Asp Val Thr Asp Tyr His 645 650 655	1968
10	ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA GAT GAA TTT TGT Ile Asp Gln Val Ser Asn Leu Val Asp Cys Leu Ser Asp Glu Phe Cys 660 665 670	2016
15	CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA CAT GCG AAG CGA Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys His Ala Lys Arg 675 680 685	2064
20	CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC TTC AGA GGG ATC Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Pro Asn Phe Arg Gly Ile 690 695 700	2112
25	AAT AGA CAA CCA GAC CGT GGC TGG AGA GGA AGT ACA GAT ATT ACC ATC Asn Arg Gln Pro Asp Arg Gly Trp Arg Gly Ser Thr Asp Ile Thr Ile 705 710 715 720	2160
30	CAA GGA GGA GAT GAC GTA TTC AAA GAG AAT TAC GTC ACA CTA CCG GGT Gln Gly Gly Asp Asp Val Phe Lys Glu Asn Tyr Val Thr Leu Pro Gly 725 730 735	2208
35	ACC GTT GAT GAG TGC TAT CCA ACG TAT TTA TAT CAG AAA ATA GAT GAG Thr Val Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln Lys Ile Asp Glu 740 745 750	2256
40	TCG AAA TTA AAA GCT TAT ACC CGT TAT GAA TTA AGA GGG TAT ATC GAA Ser Lys Leu Lys Ala Tyr Thr Arg Tyr Glu Leu Arg Gly Tyr Ile Glu 755 760 765	2304
45	GAT AGT CAA GAC TTA GAA ATC TAT TTG ATC CGT TAC AAT GCA AAA CAC Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr Asn Ala Lys His 770 775 780	2352
50	GAA ATA GTA AAT GTG CCA GGC ACG GGT TCC TTA TGG CCG CTT TCA GCC Glu Ile Val Asn Val Pro Gly Thr Gly Ser Leu Trp Pro Leu Ser Ala 785 790 795 800	2400
55	CAA AGT CCA ATC GGA AAG TGT GGA GAA CCG AAT CGA TGC GCG CCA CAC Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg Cys Ala Pro His 805 810 815	2448
60	CTT GAA TGG AAT CCT GAT CTA GAT TGT TCC TGC AGA GAC GGG GAA AAA Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg Asp Gly Glu Lys 820 825 830	2496
65	TGT GCA CAT CAT TCC CAT CAT TTC ACC TTG GAT ATT GAT GTT GGA TGT Cys Ala His His Ser His His Phe Thr Leu Asp Ile Asp Val Gly Cys 835 840 845	2544
70	ACA GAC TTA AAT GAG GAC TTA GGT GTA TGG GTG ATA TTC AAG ATT AAG Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile Phe Lys Ile Lys 850 855 860	2592

	ACG CAA GAT GGC CAT GCA AGA CTA GGG AAT CTA GAG TTT CTC GAA GAG Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu Phe Leu Glu Glu 865 870 875 880	2640
5	AAA CCA TTA TTA GGG GAA GCA CTA GCT CGT GTG AAA AGA GCG GAG AAG Lys Pro Leu Leu Gly Glu Ala Leu Ala Arg Val Lys Arg Ala Glu Lys 885 890 895	2688
10	AAG TGG AGA GAC AAA CGA GAG AAA CTG CAG TTG GAA ACA AAT ATT GTT Lys Trp Arg Asp Lys Arg Glu Lys Leu Gln Leu Glu Thr Asn Ile Val 900 905 910	2736
15	TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT GTA AAC TCT CAA Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe Val Asn Ser Gln 915 920 925	2784
20	TAT GAT AGA TTA CAA GTG GAT ACG AAC ATC GCA ATG ATT CAT GCG GCA Tyr Asp Arg Leu Gln Val Asp Thr Asn Ile Ala Met Ile His Ala Ala 930 935 940	2832
25	GAT AAA CGC GTT CAT AGA ATC CGG GAA GCG TAT CTG CCA GAG TTG TCT Asp Lys Arg Val His Arg Ile Arg Glu Ala Tyr Leu Pro Glu Leu Ser 945 950 955 960	2880
30	GTG ATT CCA GGT GTC AAT GCG GCC ATT TTC GAA GAA TTA GAG GGA CGT Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu Leu Glu Gly Arg 965 970 975	2928
35	ATT TTT ACA GCG TAT TCC TTA TAT GAT GCG AGA AAT GTC ATT AAA AAT Ile Phe Thr Ala Tyr Ser Leu Tyr Asp Ala Arg Asn Val Ile Lys Asn 980 985 990	2976
40	GGC GAT TTC AAT AAT GGC TTA TTA TGC TGG AAC GTG AAA GGT CAT GTA Gly Asp Phe Asn Asn Gly Leu Leu Cys Trp Asn Val Lys Gly His Val 995 1000 1005	3024
45	GAT GTA GAA GAG CAA AAC AAC CAC CGT TCG GTC CTT GTT ATC CCA GAA Asp Val Glu Glu Gln Asn Asn His Arg Ser Val Leu Val Ile Pro Glu 1010 1015 1020	3072
50	TGG GAG GCA GAA GTG TCA CAA GAG GTT CGT GTC TGT CCA GGT CGT GGC Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys Pro Gly Arg Gly 1025 1030 1035 1040	3120
55	TAT ATC CTT CGT GTC ACA GCA TAT AAA GAG GGA TAT GGA GAG GGC TGC Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr Gly Glu Gly Cys - 1045 1050 1055	3168
60	GTA ACG ATC CAT GAG ATC GAA GAC AAT ACA GAC GAA CTG AAA TTC AGC Val Thr Ile His Glu Ile Glu Asp Asn Thr Asp Glu Leu Lys Phe Ser 1060 1065 1070	3216
65	AAC TGT GTA GAA GAG GAA GTA TAT CCA AAC AAC ACA GTA ACG TGT AAT Asn Cys Val Glu Glu Glu Val Tyr Pro Asn Asn Thr Val Thr Cys Asn 1075 1080 1085	3264
70	AAT TAT ACT GGG ACT CAA GAA GAA TAT GAG GGT ACG TAC ACT TCT CGT Asn Tyr Thr Gly Thr Gln Glu Glu Tyr Glu Gly Thr Tyr Thr Ser Arg 1090 1095 1100	3312
75	AAT CAA GGA TAT GAC GAA GCC TAT GGT AAT AAC CCT TCC GTA CCA GCT	3360

Asn Gln Gly Tyr Asp Glu Ala Tyr Gly Asn Asn Pro Ser Val Pro Ala
 1105 1110 1115 1120
 5 GAT TAC GCT TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA 3408
 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg
 1125 1130 1135
 GAG AAT CCT TGT GAA TCT AAC AGA GGC TAT GGG GAT TAC ACA CCA CTA 3456
 10 Glu Asn Pro Cys Glu Ser Asn Arg Gly Tyr Gly Asp Tyr Thr Pro Leu
 1140 1145 1150
 CCG GCT GGT TAT GTA ACA AAG GAT TTA GAG TAC TTC CCA GAG ACC GAT 3504
 Pro Ala Gly Tyr Val Thr Lys Asp Leu Glu Tyr Phe Pro Glu Thr Asp
 1155 1160 1165
 15 AAG GTA TGG ATT GAG ATC GGA GAA ACA GAA GGA ACA TTC ATC GTG GAT 3552
 Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
 1170 1175 1180
 20 AGC GTG GAA TTA CTC CTT ATG GAG GAA 3579
 Ser Val Glu Leu Leu Leu Met Glu Glu
 1185 1190

25 (2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1193 amino acids
 (B) TYPE: amino acid
 30 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
 1 5 10 15
 40 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
 20 25 30
 Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
 35 40 45
 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile-
 50 55 60
 50 Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile 80
 65 70 75
 Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
 85 90 95
 55 Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
 100 105 110
 Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu
 115 120 125
 60 Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala

130 135 140
 Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val
 145 150 155 160
 5 Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser
 165 170 175
 Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg
 180 185 190
 10 Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val
 195 200 205
 15 Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg
 210 215 220
 Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val
 225 230 235 240
 20 Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro
 245 250 255
 Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val
 260 265 270
 25 Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu
 275 280 285
 30 Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr
 290 295 300
 Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln
 305 310 315 320
 35 Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro
 325 330 335
 40 Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala
 340 345 350
 Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg
 340 345 350
 45 Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp
 370 375 380
 Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val
 385 390 395 400
 50 Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln
 405 410 415
 55 Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His
 420 425 430
 Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile
 435 440 445
 60 Arg Ala Pro Met Phe Ser Trp Ile His Arg Ser Ala Thr Leu Thr Asn
 450 455 460

Thr Ile Asp Pro Glu Arg Ile Asn Gln Ile Pro Leu Val Lys Gly Phe
 465 470 475 480
 5 Arg Val Trp Gly Gly Thr Ser Val Ile Thr Gly Pro Gly Phe Thr Gly
 485 490 495
 Gly Asp Ile Leu Arg Arg Asn Thr Phe Gly Asp Phe Val Ser Leu Gln
 500 505 510
 10 Val Asn Ile Asn Ser Pro Ile Thr Gln Arg Tyr Arg Leu Arg Phe Arg
 515 520 525
 Tyr Ala Ser Ser Arg Asp Ala Arg Val Ile Val Leu Thr Gly Ala Ala
 530 535 540
 15 Ser Thr Gly Val Gly Gly Gln Val Ser Val Asn Met Pro Leu Gln Lys
 545 550 555 560
 20 Thr Met Glu Ile Gly Glu Asn Leu Thr Ser Arg Thr Phe Arg Tyr Thr
 565 570 575
 Asp Phe Ser Asn Pro Phe Ser Phe Arg Ala Asn Pro Asp Ile Ile Gly
 580 585 590
 25 Ile Ser Glu Gln Pro Leu Phe Gly Ala Gly Ser Ile Ser Ser Gly Glu
 595 600 605
 Leu Tyr Ile Asp Lys Ile Glu Ile Ile Leu Ala Asp Ala Thr Phe Glu
 610 615 620
 30 Ala Glu Ser Asp Leu Glu Arg Ala Gln Lys Ala Val Asn Ala Leu Phe
 625 630 635 640
 35 Thr Ser Ser Asn Gln Ile Gly Leu Lys Thr Asp Val Thr Asp Tyr His
 645 650 655
 Ile Asp Gln Val Ser Asn Leu Val Asp Cys Leu Ser Asp Glu Phe Cys
 660 665 670
 40 Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys His Ala Lys Arg
 675 680 685
 Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Pro Asn Phe Arg Gly Ile
 690 695 700
 45 Asn Arg Gln Pro Asp Arg Gly Trp Arg Gly Ser Thr Asp Ile Thr Ile
 705 710 715 720
 50 Gln Gly Gly Asp Asp Val Phe Lys Glu Asn Tyr Val Thr Leu Pro Gly
 725 730 735
 Thr Val Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln Lys Ile Asp Glu
 740 745 750
 55 Ser Lys Leu Lys Ala Tyr Thr Arg Tyr Glu Leu Arg Gly Tyr Ile Glu
 755 760 765
 Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr Asn Ala Lys His
 770 775 780
 60

Glu Ile Val Asn Val Pro Gly Thr Gly Ser Leu Trp Pro Leu Ser Ala
 785 790 795 800
 5 Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg Cys Ala Pro His
 805 810 815
 Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg Asp Gly Glu Lys
 820 825 830
 10 Cys Ala His His Ser His His Phe Thr Leu Asp Ile Asp Val Gly Cys
 835 840 845
 Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile Phe Lys Ile Lys
 850 855 860
 15 Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu Phe Leu Glu Glu
 865 870 875 880
 Lys Pro Leu Leu Gly Glu Ala Leu Ala Arg Val Lys Arg Ala Glu Lys
 885 890 895
 20 Lys Trp Arg Asp Lys Arg Glu Lys Leu Gln Leu Glu Thr Asn Ile Val
 900 905 910
 25 Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe Val Asn Ser Gln
 915 920 925
 Tyr Asp Arg Leu Gln Val Asp Thr Asn Ile Ala Met Ile His Ala Ala
 930 935 940
 30 Asp Lys Arg Val His Arg Ile Arg Glu Ala Tyr Leu Pro Glu Leu Ser
 945 950 955 960
 Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu Leu Glu Gly Arg
 965 970 975
 35 Ile Phe Thr Ala Tyr Ser Leu Tyr Asp Ala Arg Asn Val Ile Lys Asn
 980 985 990
 40 Gly Asp Phe Asn Asn Gly Leu Leu Cys Trp Asn Val Lys Gly His Val
 995 1000 1005
 Asp Val Glu Glu Gln Asn Asn His Arg Ser Val Leu Val Ile Pro Glu
 1010 1015 1020
 45 Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys Pro Gly Arg Gly-
 1025 1030 1035 1040
 Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr Gly Glu Gly Cys
 1045 1050 1055
 50 Val Thr Ile His Glu Ile Glu Asp Asn Thr Asp Glu Leu Lys Phe Ser
 1060 1065 1070
 Asn Cys Val Glu Glu Glu Val Tyr Pro Asn Asn Thr Val Thr Cys Asn
 1075 1080 1085
 Asn Tyr Thr Gly Thr Gln Glu Glu Tyr Glu Gly Thr Tyr Thr Ser Arg
 1090 1095 1100
 60 Asn Gln Gly Tyr Asp Glu Ala Tyr Gly Asn Asn Pro Ser Val Pro Ala

	1105		1110		1115		1120
	Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg						
		1125			1130		1135
5	Glu Asn Pro Cys Glu Ser Asn Arg Gly Tyr Gly Asp Tyr Thr Pro Leu						
		1140		1145			1150
	Pro Ala Gly Tyr Val Thr Lys Asp Leu Glu Tyr Phe Pro Glu Thr Asp						
10		1155		1160			1165
	Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp						
		1170		1175			1180
15	Ser Val Glu Leu Leu Leu Met Glu Glu						
	1185		1190				

All of the compositions and methods disclosed and claimed herein can be made and-executed without undue experimentation in light of the present disclosure. While the compositions and methods of this invention have been described in terms of preferred embodiments, it will be apparent to those of skill in the art that variations may be applied
5 to the compositions and methods and in the steps or in the sequence of steps of the method described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents which are both chemically and physiologically related may be substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and
10 modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention as defined by the appended claims.